Generic Cell Surface Staining Procedure

Materials:

COMPtrol Kit (Spherotech #CMIgP-30-2K) – Antibodies Stained according to manufacturer's directions

- 1. Add 50 ul beads (orange cap) to each comp tube
- 2. Add antibody to each tube (from Excel sheet)
- 3. Incubate for 15-30 min at RT
- 4. Add 50 ul of Blank beads to each tube and the unstained tube
- 5. Add 2 ml D-PBS to each tube and centrifuge at 1500 RPM for 5 min
- 6. Decant supernatant and resuspend in 300 ul D-PBS

ArC Amine Reactive Compensation Kit (ThermoFisher #A10628) – Viability Dye Stained according to manufacturer's directions

- 1. Add 1 drop of beads (green top tube) to tube
- 2. Add 3 ul Live/Dead Blue dye to tube
- 3. Incubate for 30 min at RT
- 4. Add 1 drop of Blank ARC beads to tube
- 5. Add 2 ml D-PBS to each tube and centrifuge at 1500 RPM for 5 min
- 6. Decant supernatant and resuspend in 300 ul D-PBS

Viability Dye

Live/Dead Fixable Blue Dead Cell Stain (ThermoFisher #L23105)

- 1. Add 50 µl DMSO to 1 vial of dye to reconstitute
- 2. Add 50 µl D-PBS (Ca2+ Mg2+ free) to vial
- 3. Use 1µl to stain each sample

*Note: This procedure will work with other colors of Live/Dead Fixable Dyes

Antibodies

Titrate antibodies prior to staining

Prepare antibody staining cocktail by multiplying the amount of each titrated antibody per sample by the number of samples + 2 extra samples. Add all antibodies into one mixture and then use that mixture to stain each sample.

<u>Cells</u>

Prepare cells by centrifuging out of media and resuspending at 1×10^7 cells/ml in D-PBS (Ca²⁺ Mg²⁺ free)

Staining Procedure:

- 1. Prepare antibody staining cocktail and viability dye
- 2. Stain compensation beads
- 3. Add 100 μ I cells (1 x 10⁶ cells) to 12 x 75 mm polystyrene tubes (Falcon #352058)
- 4. Add 1 μl Live/Dead Blue dye to cells and incubate for 10 min at room temperature, protect from light
- 5. Add antibody cocktail to tubes and incubate for 30 min at RT, protect from light
- 6. Add 2 ml D-PBS (Ca²⁺ Mg²⁺ free) to each tube and centrifuge at 1500 RPM (450g) for 5 min. *Note: A centrifuge with a swinging bucket rotor works much better than a microcentrifuge.
- 7. Decant supernatant and resuspend in 300 µl 1% ultrapure formaldehyde
- 8. Refrigerate until acquisition