User's Manual

CellVoyager CV7000

High-throughput Cytological Discovery System

IM 80H01C01-01E



IM 80H01C01-01E 14th Edition

Introduction

Thank you for purchasing the CellVoyager CV7000.

This user's manual describes the functions, operating procedures, and safety and handling precautions of the CV7000. Before you start to use the CV7000, please read this manual carefully to enable correct use of the system. For more information on the specifications, refer to the technical specifications that are published separately.

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After reading this manual, keep it in a handy place so that you can refer to it whenever necessary.

Notes

- Unauthorized reproduction or reprinting of this manual in whole or in part is prohibited.
- The information contained in this manual is subject to change without prior notice due to improvements in performance and functionality or for other reasons.
- This manual has been prepared with the utmost care; however, if you have any questions, or note any errors, please contact us.
- Follow the operating instructions to ensure that the system remains stable.

Trademarks

- CellVoyager and CSU are registered trademarks of Yokogawa Electric Corporation.
- Microsoft and Windows are registered trademarks or trademarks of Microsoft Corporation in the United States and other countries.
- The CellVoyager is using patents on High Content Screening and High Content Analysis of Thermo Fisher Scientific Inc.
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Relevant Documents

- ●IM 80H01C03-01E CV7000 Analysis Software User's Manual
- ●IM 80H01A16-01E Image Correction Software User's Manual
- ●IM 80H01A17-01E Postprocessor User's Manual

History

June 2012 1st edition issued.

September 2012 2nd edition issued.

Modification

Added the explanation about camera gain setting.

November 2012 3rd edition issued.

Modification

Added camera binning 2×2 for sCMOS.

Added setting items for source plate name and source plate information in the

"Start Measurement" screen when dispensing measurement.

Added the setting items for postprocessor function.

February 2013 4th edition issued.

Modification

Added the explanation about plate information.

October 2013 5th edition issued.

Modification

Added the AirBlow function in the dispensing setting file.

Added Plate Bottom for tip position.

Added the SUM function for Image Processing in the 3D Fluorescence Acquisition and Z-stack Bright-field/Phase-contrast Acquisition screens.

Added snapshot function in the measurement result sub screen.

Added search filter function in the Measurement Setting List screen.

Modified the screen for high-speed time-lapse setting.

November 2013 6th edition issued.

Modification

Added the explanation about QUAD-DM model.

Modified bright-field acquisition to match pixel position with each image of confocal or epifluorescent imaging.

Added the explanation for 384-tip rack model.

December 2013 7th edition issued.

Modification

Added the function to record CO2 concentration and temperature logs.

Added XY offsets in the "Add Acquisition Points" screen.

June 2014 8th edition issued.

Modification

Added the function to perform high-speed time-lapse imaging with multi dispensing.

Added the function to expansion the Acquisition Point Screen.

Changed the operability of field selection.

Added the explanation of slide glass holder.

November 2014 9th edition issued

Modification

Added binning 3x3 and 4x4.

Added the function to perform high-speed time-lapse Imaging working with normal Imaging.

January 2015 10th edition issued

Modification
Added procedure for registering and selecting fluorophore.
Added explanation of high precision autofocus mode.
Added explanation of performing autofocus during high-speed time-lapse
imaging.
Modified the explanation of slide glass holder.

April 2015 11th edition issued

Modification
Added "Relevant Documents"
Added explanations about partial tiling function.
Deleted explanations about Camera Setting (Camera gain).
Added cautions of sample setting
Added explanation about changes of CO ₂ concentration control

October 2015 12th edition issued

Modification					
Added explanations about time line copying function.					
Modified explanations about CO2 concentration setting of stage					
incubator.					
Added explanations about imaging forbidden area of using water					
immersion objective lens.					
Modified explanations about slide glass measurement using water					
immersion objective lens.					
Added explanations about display of number of times that dispenser					
syringe moves.					
Modified explanations about saving path of measurement files.					

June 2016 13th edition issued

Modification							
Added e	explanations	about	digital	phase	contrast	(DPC)	acquisition
function.							
Modified explanation about Stage Incubator setting.							
Added Well plate registration function.							
Added explanation about On-the-fly image correction function.							

August 2016 14th edition issued

Modification

Modified description for WEEE directive.

Added table of MS code

For Safe Use of This Equipment

To ensure safe and correct use of this product, be sure to follow the precautions below. If this product is used in a manner not specified by this manual, the protection provided by this product may be impaired. If you do not follow the precautions when handling the product, we do not guarantee the safety and product.

The following safety symbols are used for this product.

WARNING or 🖄 CAUTION

These symbols indicate that you need to handle the equipment with care. These symbols are displayed in locations where you need to refer to the user's manual to protect your safety and the safety of equipment.

These symbols in the manual are used in the following cases.

PROTECTIVE GROUND TERMINAL

This symbol indicates a protective ground terminal. Be sure to provide a ground connection before turning on the equipment to prevent an electric shock.

\sim ALTERNATING CURRENT

This symbol indicates an alternating current.

POWER ON

This symbol indicates POWER ON.



POWER OFF

This symbol indicates POWER OFF.



On the equipment, this symbol is displayed in the locations where you must refer to the manual to protect yourself and the equipment from serious accidents. In the manual, this symbol is placed near text that describes the precautions to help avoid hazardous situations that could result in a bodily injury or death of the user, such as an electric shock, or that could result in damage to the equipment and devices.

♠ CAUTION

On the equipment, this symbol is displayed in the locations where you must refer to the manual to protect yourself and the equipment from accidents. In the manual, this symbol is placed near text that describes the precautions to help avoid hazardous situations that could result in a minor bodily injury of the user, or that could result in damage to the equipment.

- Be sure to turn on the power to the equipment after confirming that the power-supply voltage of the equipment matches the voltage of the supplied power.
- This is a Class 1 laser product. However, the equipment houses a Class 3B laser, which is protected by the enclosure and the interlocks provided at openings. When using this product, heed the precautions explained in "Laser Products Handling Precautions."
- Inhalation of 5%CO₂ gas may be harmful to the human body. Be sure to provide ventilation, lead exhaust gas from the incubator to outside of the room, or take other appropriate measures.
- Do not use the equipment by wet hand. It may cause electric shock and suffer electrical or mechanical damages.
- Do not bring electrically charged objects near the signal terminals. Equipment failure may result.
- Do not pour volatile chemicals onto the display or keep it in contact with rubber or plastic products for an extended period of time.
- Do not give shock to this equipment.
- Do not block the vent openings.
- Should you notice smoke coming out of the main unit, foul smell, abnormal noise or any other abnormality, immediately turn off the power switch and cut off the power supply. Contact the Yokogawa dealer from which you have purchased the equipment or Yokogawa's service department to notify the abnormality.
- To stop the equipment immediately in case of emergency, press the EMERGENCY STOP button. To release the button, turn it clockwise.



- •Use of controls or adjustments or performance of procedures other than those specified herein may result in hazardous exposure.
- Use the I/O terminals of the equipment within the range of the specifications to prevent damage to the equipment.
- This product uses precision optics. Therefore, do not use the equipment in locations where there are large vibrations, a lot of dust, high humidity and high temperature (in places near heating equipment or exposed to direct sunlight), rapid changes in the temperature (dew condensation), or corrosive or combustible gases.
- Never touch any of the internal parts of this product. The optical system housed in the product may become dirty, damaged or out of calibration, etc., leading to equipment failure.
- If the equipment malfunctions, contact us without attempting to access the inside or take any other action to resolve the problem yourself.

WEEE (Waste Electrical and Electronic Equipment), Directive



(This directive is only valid in the EU.)

- This instrument complies with the WEEE Directive marking requirement. The marking above indicates that you must not discard this instrument in domestic household waste.
- Product Category

With reference to the instrument type in the WEEE directive Annex 1, this product is classified as a "Monitoring and Control instrument" product.

•When disposing this instrument in the EU, contact the distributer whom you bought it from. Do not dispose in domestic household waste.

Warning and Caution Labels

The safety warning and caution labels attached on this product are listed below.

Warning Label (1)



Warning Label (3)



Caution Label (1)



Caution Label (2)

CAUTION- CLASS 3B LASER RADIATION WHEN Open. Avoid exposure to the beam

Caution Label (3)



Warning Label (2)



Warning Label (4)



Warning Label (1)

This label warns you to be careful because there is a risk that your hand or clothes may get caught. This label warns you not to approach the spot during operation.

Warning Label (2)

This label indicates a risk that your hand may get caught. This label warns you not to insert your hand.

Warning Label (3)

This label indicates a risk of an electric shock and warns you not to touch the conductor.

Warning Label (4) This label indicates a risk of laser radiation. Do not stare into beam.

Caution Label (1)

This label warns that safety management is required to use the laser. This label indicates a risk of your eyes or skin being exposed to a class 3R laser beam if you open this part.

Caution Label (2)

This label warns that safety management is required to use the laser. This label indicates a risk of your eyes or skin being exposed to a class 3B laser beam if you open this part.

Caution Label (3)

This label indicates that it is necessary to refer to this manual.

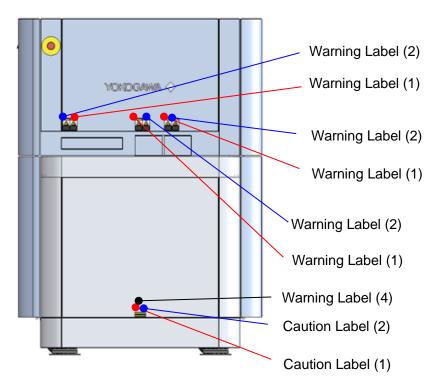
Locations of Warning Labels

Warning labels are placed in locations where safety management is required to use this product. The following shows the placed warning labels and their placement positions. If the equipment malfunctions, contact us without attempting to access the inside or take any other action to resolve the problem yourself.

Front Panel

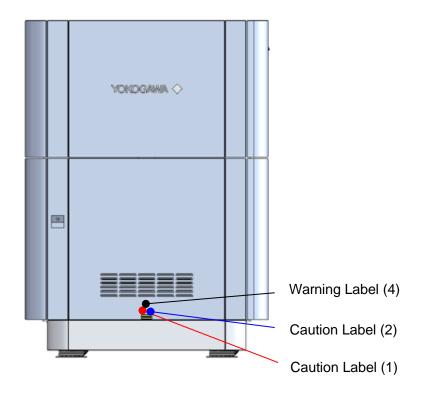


Right Side Panel

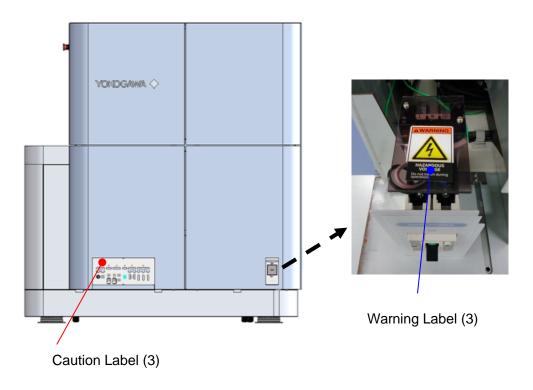


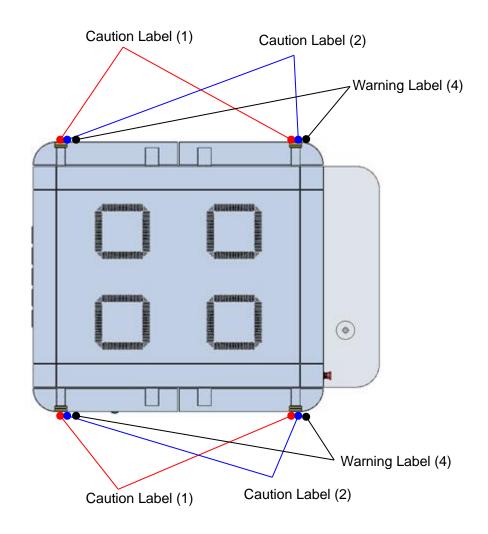


Left Side Panel



Rear Panel





Top Panel

Laser Product Handling Precautions

Laser products are classified based on their exposure emission limit determined by the wavelength and power characteristics of laser beam. A different set of common safety standards applies to each class of products. The product explained herein belongs to Class 1. However, the equipment houses a Class 3B laser, which is protected by the enclosure and the interlocks provided at openings.

Only service personnel can remove the covers on which a warning label is attached. If these covers are removed, class 3B laser beam will be emitted. Directly looking into Class 3B laser beam or beam reflected on a mirror may cause eye damage and is extremely dangerous. Exercise due caution when handling this product.

Safety Standards

A class 1 laser is safe under all conditions of normal use. This means the maximum permissible exposure cannot be exceeded. This equipment is designed in accordance with the IEC60825-1 Radiation safety standards for laser products and these lasers must be labeled with the following label, but are exempt from the requirements of the Laser Safety Program.

CLASS 1 LASER PRODUCT IEC60825-1 : 2007

Indemnity

- Yokogawa shall provide no warranty regarding this product, unless otherwise specified separately in "Warranty Terms."
- Yokogawa shall assume no responsibility for any loss suffered by a customer or third party as a result of use of this product, or any loss or indirect loss suffered by a customer or third party due to a defect in this product or any other problem not predictable by Yokogawa.

Laser Specification

Barcode Reader

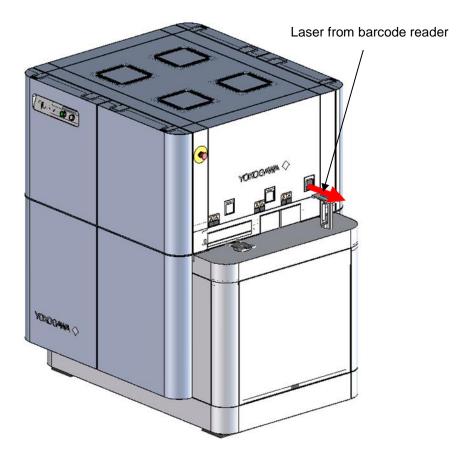
This product emits the laser below if it has barcode option.

Wavelength: 660 nm Output: 0.09 mW Pulse width: 200 μ s Beam divergence: parallel beam This laser beam is not for barcode reading. This laser beam is only for aiming beam and is only used at installation time.

Laser from barcode reader

(1) 2-barcode reader option

(2) 1-barcode reader option



Microscope

This product has lasers below in it.

- (1) 405 nm laser
 Wavelength: 405 ±5 nm
 Output: 100 mW
 Beam divergence: 0.4 mrad
 Laser type: Continuous
- (2) 488 nm laser
 Wavelength: 488 ±2 nm
 Output: 200 mW
 Beam divergence: 1.2 mrad
 Laser type: Continuous
- (3) 532 nm laser
 Wavelength: 532 ±2 nm
 Output: 200 mW
 Beam divergence: 1.3 mrad
 Laser type: Continuous
- (4) 561 nm laser
 Wavelength: 561 ±2 nm
 Output: 200 mW
 Beam divergence: 1.3 mrad
 Laser type: Continuous
- (5) 640 nm laser
 Wavelength: 640 +4/-5 nm
 Output: 100 mW
 Beam divergence: 1 mrad
 Laser type: Continuous
- (6) 785 nm laser
 Wavelength: 785 +15/-10 nm
 Output: 2.5mW
 Beam divergence: parallel beam
 Laser type: Continuous

Applicable Standards

CE Marking

•EMC Directives:

EN 61326-1 Class A, Table 1 (Basic immunity requirements)

Electrical equipment for measurement, control and laboratory use - EMC requirements -

Part 1: General requirements

Machinery Directive:

ISO12100

Safety of machinery

- General principles for design - Risk assessment and risk reduction

EN 13849-1

Safety of machinery

- Safety-related parts of control systems - Part 1: General principles for design

EN 61010-1

Safety requirements for electrical equipment for measurement, control, and laboratory use, Part 1: General requirements

EN 60825-1

Safety of laser products, Part 1: Equipment classification and requirements

EC DECLARATION OF CONFORMITY

YOKOGAWA

For

High-throughput Cytological Discovery System Model: CV7000

Manufactured by

Yokogawa Electric Corporation 2-9-32 Nakacho, Musashino-shi, Tokyo, 180-8750 Japan

Means of Conformity

The Product is in conformity with EC law as approximated by the Machinery Directive 2006/42/EC, based on Technical Documentation File No.2193 Issue 1, Revision 0, November 2013

> Standards used as guidance Machinery Directive

EN ISO 12100: 2010 EN ISO 13849-1: 2008 EN 60825-1: 2007 EN 61010-1: 2010 Refer to complete listing in Technical File

The Machinery Directive Technical File compiled from manufacturers documentation and held in the EU, on behalf of the manufacturer by

> TRaC Global Ltd 100 Frobisher Business Park, Leigh Sinton Road, Malvern, Worcestershire, WR14 1BX United Kingdom

Signature of Responsible Person:

Mr. Takayuki Kei Technical General Manager Life Science Headquarters R&D Department Yokogawa Electric Corporation

Takayuki Kei

Date:

21st November 2013

Document No.

TRA-015425-00 DofC Issue 1

Doc. No.: EEN302-C03

EC DECLARATION OF CONFORMITY

We Yokogawa Electric Corporation 2-9-32 Nakacho, Musashino-shi, Tokyo, 180-8750 Japan

declare under our sole responsibility that the product

CV7000 High-throughput Cytological Discovery System (See Appendix for the detailed type designation)

to which this declaration relates is in conformity with the following standards or other normative documents:

EN 61326-1: 2013 Class A, Table 1 (Basic immunity requirements) Electrical equipment for measurement, control and laboratory use – EMC requirements-Part 1: General requirements

following the provisions of EMC directive 2004/108/EC.

Subject products are manufactured and tested according to appropriate quality control procedures.

Tokyo, 31 July, 2015

Signature:

Takayuki Kei

Takayuki Kei R&D Section Manager Life Science Headquarters Yokogawa Electric Corporation

Yokogawa Electric Corporation

How to Use This Manual

This user manual consists of Chapters 1 to 14, the details of which are explained below.

Chapter	Title and content				
1	Overview of the Equipment				
	A functional overview of this equipment is explained.				
2	Before Use				
	Installation and wiring methods are explained.				
3	Starting and Shutting Down the Equipment				
	The name of each part and starting/shutdown of the equipment are				
	explained.				
4	Entering Well Plate Information Files				
	Entry of well plate information is explained.				
5	Using the Measurement Software				
	Functions and operations of the measurement software are explained.				
6 Explanation of Measurement Software Screens					
	Screens of the measurement software are explained.				
7	Setting Examples of Measurement Setting Files				
	Examples of measurement using this equipment are explained.				
8	Measurement				
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9	Checking Measured Images				
	Checking of captured images is explained.				
10	Saving Measured Files				
	Where to save captured images is explained.				
11	Troubleshooting				
	Troubleshooting methods for this equipment are explained.				
12	Maintenance and Inspection				
	Maintenance and inspection of this equipment are explained.				
13	Warranty				
	The terms of warranty applicable to this equipment are explained.				
14	General Specification				
	The general specification about CV7000 is explained.				

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1. Overview of the Equipment

1.1. About the CV7000

In basic research fields of medicine, biology, pharmacology, agriculture, etc., as well as applied research fields such as drug discovery, advancement of bio-research is making "live cell imaging" a general approach to observing live cells for a long period.

This equipment is a high-throughput cytological discovery system designed to let the users study various reactions of live cells both quickly and in detail to increase the efficiencies of drug development, compound evaluation, cell function study, etc.

Features of the CV7000

- The industry's most advanced live cell observation function The CV7000 is equipped with a confocal scanner unit CSU that uses the multi-scan system to minimize cell damage due to laser irradiation.
- The industry's fastest screening High-speed precision positioning, high-speed auto-focus and high-speed image acquisition technologies are combined to achieve high throughput. High-speed image acquisition is attained at less than 60 sec. for a 96 wellplate.
- 3. One of the industry's best optical technologies

The CV7000 adopts the high-resolution real confocal system using Nipkow disks with micro-lens array. The user can also freely change the observation mode among confocal, epifluorescence and phase contrast.

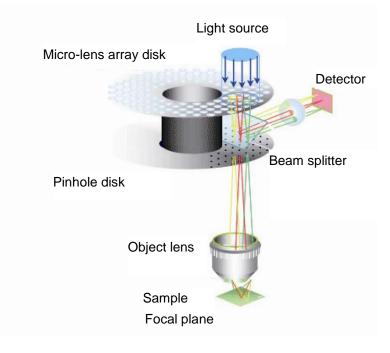
Fourfold wide-field images can be acquired by using the confocal scanner unit designed for wide-field imaging. (Compared to the CV6000)

1.2. Operating Principle of the Nipkow Disk System with

Micro-lens Array

Two disks, including the "pinhole array disk" having many pinholes arranged in a helical pattern, and the "micro-lens array disk" that condenses excited laser to individual pinholes, are operated jointly at high speed to perform multiple scans over the observation area with approximately 1,000 laser beams. Multiple beam scans are performed not only at high speed, but also with each beam exciting fluorochromes at high efficiency and very low laser intensity.

This results in an optimal live cell observation system where phototoxicity and fluorescence photobleaching are suppressed notably compared to any conventional system.



Operating Principle of the Nipkow Disk System with Micro-lens Array

2. Before Use This Equipment

2.1. Installing This Equipment

Installation Conditions

The following utilities are required.

Main unit

Power supply:230VAC 50Hz 2kVA max Grounding resistance:100 ohm or less CO₂:Purity 99.95% or higher 0.2L/min or more 0.3MPa Air:Cleanliness grade, Humidity and moisture content grade 6(dew point underpressure 10°C or below), Solid grain grade 2, oil grade 2, foul smell removed by active carbon filters, 5L/min or more 0.6MPa

Workstation

Power supply:100V-240VAC±10% 50/60Hz 1.5kVA max

Installation Location

Install this equipment in a location meeting the conditions specified below.

- Location where enough space is available Installation of this equipment requires a space of at least 2900 W x 3000 D x 2550 H (mm).
- Location subject to minimal mechanical vibration Install this equipment by selecting a location subject to minimal mechanical vibration.
- Level location

When installing this equipment, make sure the equipment does not tilt to the left or right, but remains level.

Location where the floor has sufficient strength

This equipment weighs 650 kg. Install it in a location where the floor is strong enough to withstand this load.

- Location subject to minimal lamp soot, steam, dust, corrosive gases, etc.
- Install this equipment in a location where height above sea level is less than 1000m.

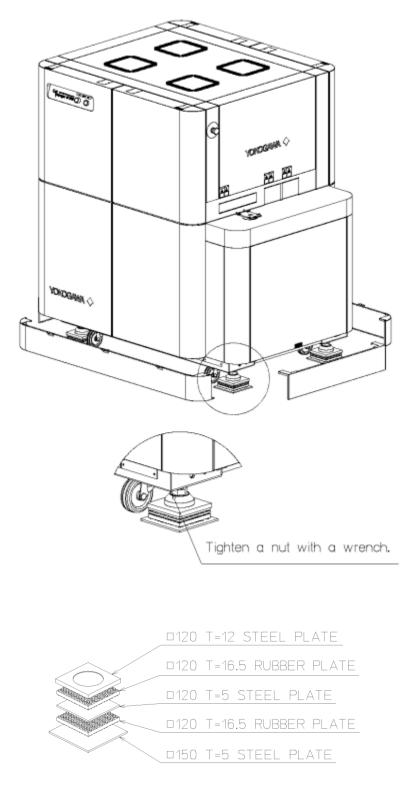
Note

Moving the equipment from a hot, humid location to a high-altitude location or otherwise subjecting the equipment to a sudden temperature shift may cause bedewing.

Installation Environment

Ambient operating temperature range: 15 to 30°C Ambient operating humidity range: 10 to 70%RH non-condensing

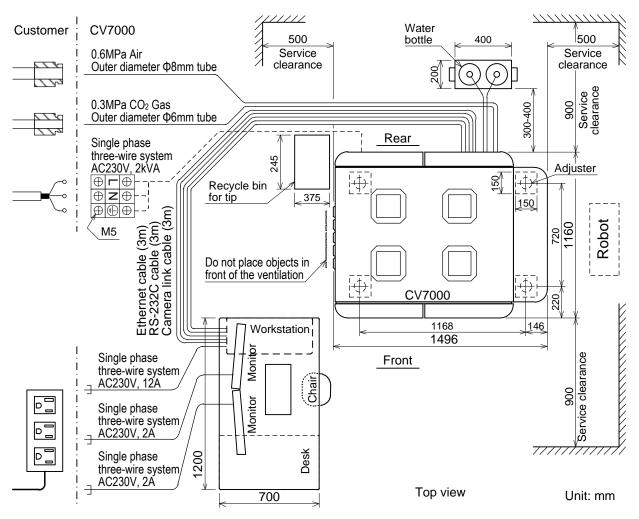
Main Unit installation



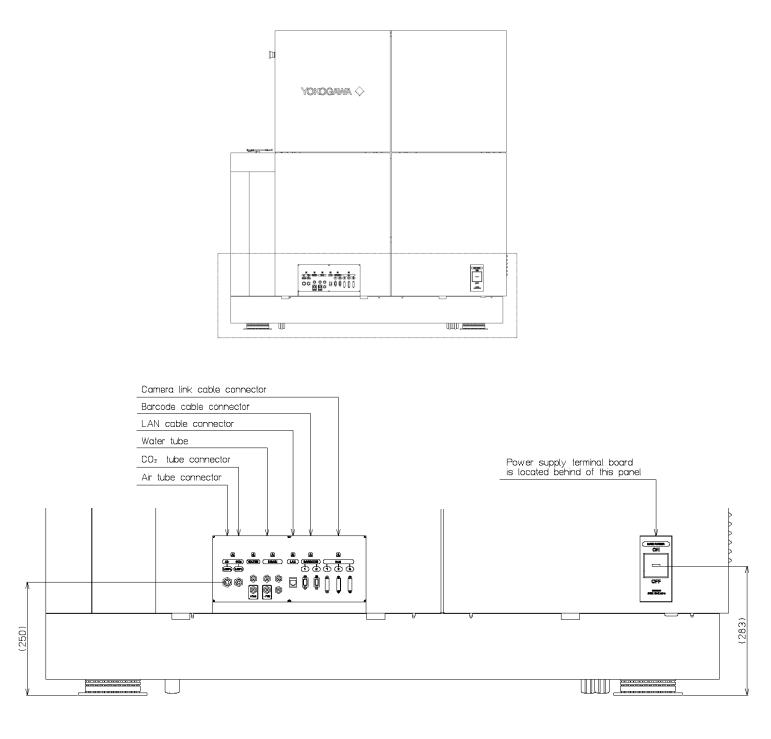
2.2. Connecting the System

Wiring the Peripherals

Connect the cables after confirming that the power switch on the equipment is turned off.



Connecter boards are located lower-left side of rear panel. Identification of connectors is shown as below.



- Pulling a cable wired to this equipment with a strong force may cause damage to the cable or connected terminals on the equipment. Provide each cable with an ample allowance so that the input terminals on the equipment will not receive a direct pulling force.
- Connect the gas cylinder(CO₂100%) firmly so that CO₂ will not leak. Install an alarm system as a precautionary measure.

Connection Procedure

Connecting the input power supply to the main unit

- 1) Remove the rear bottom cover of the equipment.
- 2) Remove the rear right cover of the equipment.
- 3) Remove the cover on the input terminals.
- 4) Connect a power cable (AWG16 or larger that can accommodate 230 VAC, 10 A is recommended) to the input terminals L, N and . Guide the power cable on the floor and through the power cable wiring aperture in the breaker assembly to connect to the input terminals. As for termination, attach at the end of each lead a crimp terminal or other appropriate terminal matching the terminal screw (M5) and securely connect it to the input terminal.
- 5) Guide cable clamps through the service apertures and clamp the power cable.
- 6) After the power cable has been connected, install the cover on the input terminals.
- 7) Install the rear right cover of the equipment.
- 8) Install the rear bottom cover of the equipment.

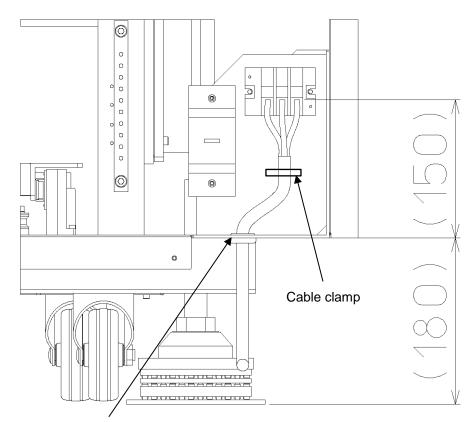
$riangle ext{ CAUTION } ext{$

- The power cable is not supplied. The customer must prepare an appropriate cable in accordance with National Wiring Regulations.
- Be sure to clamp the cable. If cable clamps are not used, the input terminals may receive a load and become loose, resulting in heating of the connection points, electric shock or other dangerous situation.
- The above procedure must be carried out by a qualified electrician.
- 9) Turn off the switch of the power distribution panel.
- 10) Connect the power cable to L, N and in on the power distribution panel. As for termination, attach at the end of each lead a crimp terminal or other appropriate terminal matching the terminal screw on the power distribution panel and securely connect it to the terminal on the panel.

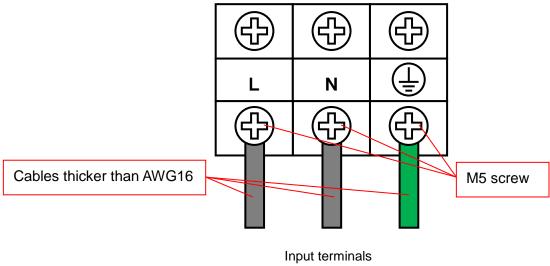
- If the polarities on the power distribution panel are not clear, always ask a qualified electrician or licensed electrician to check.
- Be sure to use the switch of the power distribution panel. If the power distribution panel has no switch, add a switch.
- If the terminal screws are not tightened firmly, the cables may come off or connection points may be heated and create a dangerous situation.
- •Be sure to use cables and crimp terminals of matching sizes.

THIS EQUIPMENT MUST BE EARTH.

Rated power supply voltage:230VAC Allowable power-supply voltage fluctuation range:207 to 253VAC Rated power frequency:50Hz Maximum power consumption:2kVA



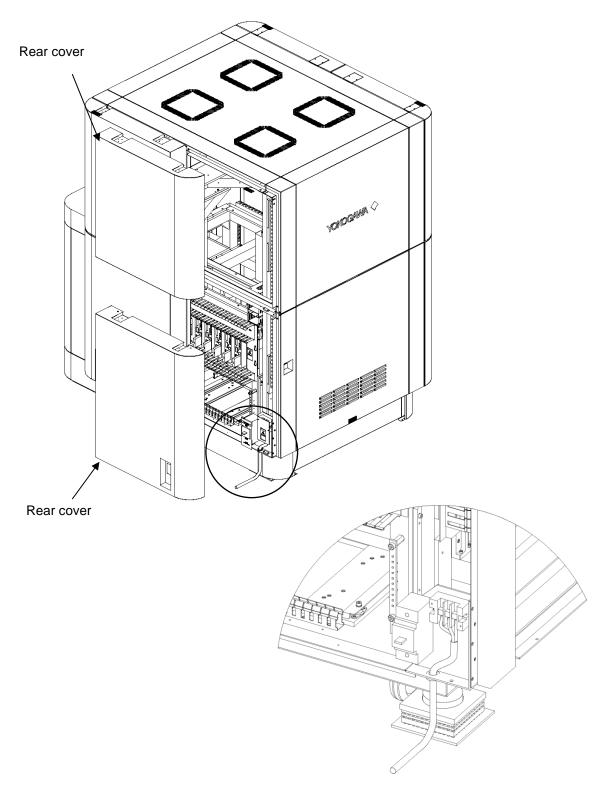
Aperture for wiring the power cable



(Rear view)

Parenthetic numbers represent reference dimensions.

Cable Installation



Removal of covers

Before connecting the power, read the following warnings. A failure to observe these warnings may result in electric shock or equipment damage.

Power supply

Be sure to turn on the power to the equipment after confirming that the power-supply voltage of the equipment matches the voltage of the supplied power.

Protective grounding

To prevent electric shock, be sure to provide protective grounding before turning on the power to this equipment.

Defects in protective functions

Do not cut the protective grounding wires running inside or outside this equipment or remove the connected wires from the protective grounding terminals. Doing so will put this equipment in a dangerous condition. Also check the protective functions to ensure absence of defects before operating this equipment.

Use in gas

Do not operate this equipment in a location where it may come in contact with flammable or explosive gases or vapors. Using this equipment in such environment is very dangerous.

Removing the case

Only Yokogawa's service personnel can remove the case. This equipment houses high-voltage parts and wires as well as a Class 3B laser, so it is dangerous to remove the case without due caution.

External connections

Connect each external device after confirming proper protective grounding.

Power Connection to the Workstation and Display

With both, connect the power cable after confirming that the power switch is turned off. Connect the plug on the other end of the power cable to a power outlet meeting the conditions specified below. For the power outlet, use a 3-pin power socket with protective grounding terminal.

Workstation Rated voltage : 230 VAC Rated current : 12 A Power-supply voltage fluctuation range : 207 to 253 VAC Rated power frequency : 50/60 Hz

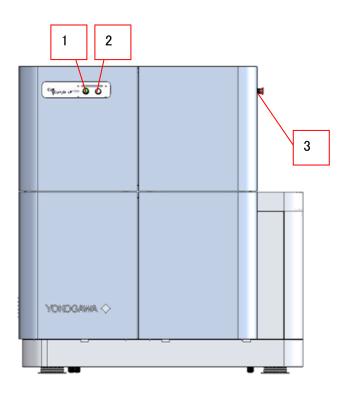
Display Rated voltage : 230 VAC Rated current : 2 A Power-supply voltage fluctuation range : 207 to 253 VAC Rated power frequency : 50 Hz Before connecting the power, read the following warnings. A failure to observe these cautions may result in electric shock or equipment damage.

- Connect the power cable after confirming that the supply voltage matches the rated power-supply voltage of this equipment.
- Connect the power cable after confirming that the power switch on this equipment is turned off.
- To prevent electric shock and fire, be sure to use the power cable supplied by Yokogawa.
- Be sure to provide protective grounding to prevent electric shock. Connect the power cable of this equipment to a 3-pin power outlet with protective grounding terminal. To connect the power cable to a 2-pin power outlet, use a 3-pin to 2-pin conversion adapter (usable only in Japan) and firmly connect the grounding wire of the conversion adapter to the protective grounding terminal of the power outlet.
- Do not use any extension cable without protective grounding wire. Use of such cable will disable the protective operations.

3. Starting and Shutting Down the Equipment

3.1. Name and Function of Each Part

Front View



1) POWER ON button

Use this button to start the system.

2) POWER OFF button

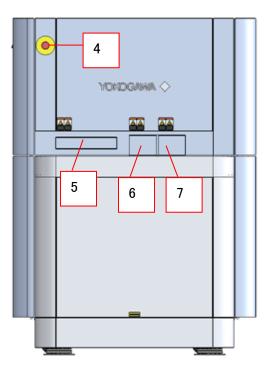
Use this button to shut down the system.

3) EMERGENCY STOP button

Use this button to stop the CV7000 immediately in case of emergency.

Right Side Panel

No barcode reader model



4) EMERGENCY STOP button

Use this button to stop the CV7000 immediately in case of emergency.

5) Assay plate loader

Use this to load the assay plate.

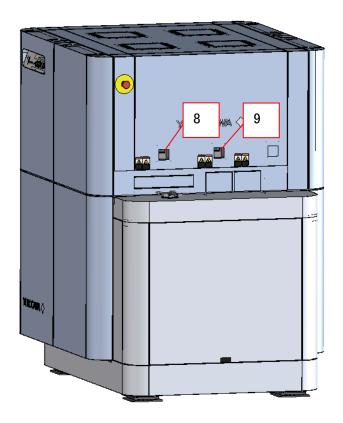
6) Source plate loader

Use this to load the source plate.

7) Tip rack loader

Use this to load a tip rack.

2-barcode reader model



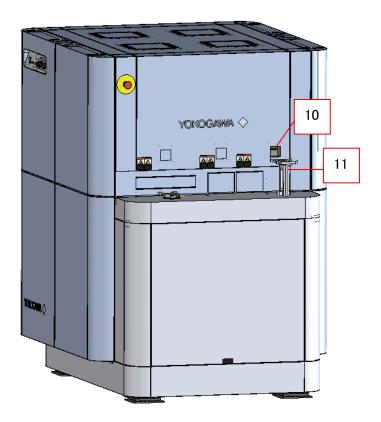
- Barcode reader for assay plate
 Use this to read barcode for the assay plate.
- 9) Barcode reader for source plate
 - Use this to read barcode for the source plate.



Class 1 laser is output from barcode reader. Do not stare into the beam.

• Do not disassemble barcode reader product. Laser emission from this product is not automatically stopped when it is disassembled.

1-barcode reader model



10) Barcode reader for assay plate and source plate

Use this to read barcodes for assay plate and source plate.

11) Reserve space

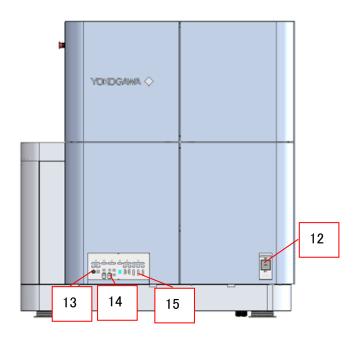
Use this to put on assay plate and source plate when reading the barcodes.



•Class 1 laser is output from barcode reader. Do not stare into the beam.

Do not disassemble barcode reader product. Laser emission from this product is not automatically stopped when it is disassembled.

Rear View



12) MAIN POWER breaker

For the main power.

13) Air connector and water connector

Air and water are supplied through these connectors.

14) LAN port

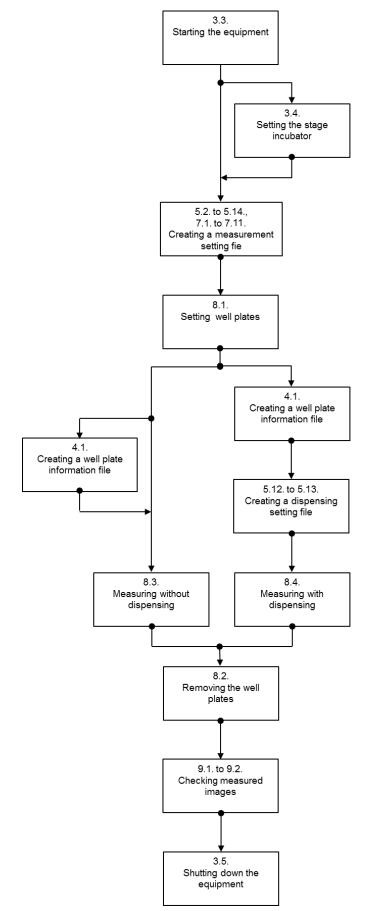
Connect to the system and workstation.

15) Camera port

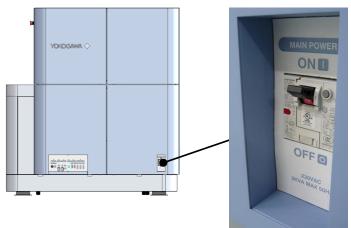
Connect the camera between the system and workstation.

3.2. Flowchart from Equipment Startup to Measurement

A flowchart from equipment startup to measurement is shown below.



3.3. Starting the Equipment



1) Turn on the MAIN POWER breaker on the rear side of the equipment.

2) Press the POWER ON button on the front side of the equipment. The POWER ON lamp becomes lit.



- 3) Start the measurement PC.
- 4) Click the icon below on the desktop of the measurement PC to start the application software.



5) The portal application software starts. Wait a while until the start is completed.

Reader Status: Initializing Reader Device Progression of Initialization:	"Ready" appears when the start is completed.
Connecting Device Initialize Larers Initialize Larup Initialize AT Initialize Stage Initialize Stage Initialize Stage Nationalize Dispenser Start Device	



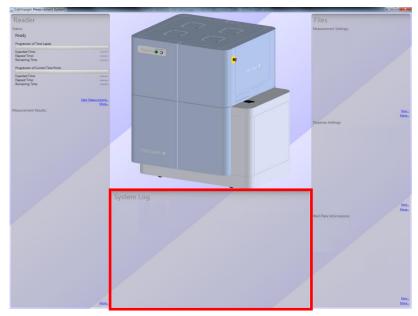
In case that following window appears and it says "The CSU is stopped", click "OK" (after clicking application software closes automatically).

CellVoyager Measurement System	Σ
The following error occurred:	
Reader device initialization erro: The CSU is stopped.	
CellVoyager Measurement System will not work correctly. Please exit this software and check device status.	
OK Cancel	

After application software closes, please re-start CV7000 (press POWER OFF button to shut down, and press POWER ON button to turn on).



If the **Q** mark appears in the System Log field, stop measurement and contact us.



Refer to 3.6 for the screen of the application software.

3.4. Setting the Stage Incubator

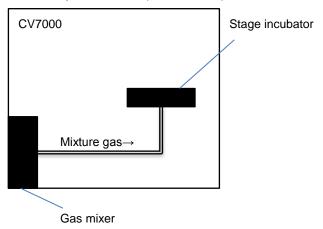
The stage incubator is set. (Stage incubator model only)



●CO₂ density control is different in CV7000 which is sold before March 2015 (conventional model) and after April 2015 (CV7000S)

- Conventional model (CV7000)
 Set CO₂ density and gas flow rate at outlet of gas mixer
- CV7000S

Set CO₂ density at stage incubator. Gas flow rate is fixed to optimal value (not settable)



 Hardware of CV7000 must be altered to change the CO₂ density control from conventional model to CV7000S (paid service)

Please contact dealer for detail

Preparations

- 1) Confirm the connection of the CO_2 cables.
- 2) Install a CO₂ detector to avoid carbon-dioxide toxicity.
- 3) Prepare the water supply bottle and the drainage bottle. Put pure water into the water supply bottle. (Pour off the water in the bottles regularly.)

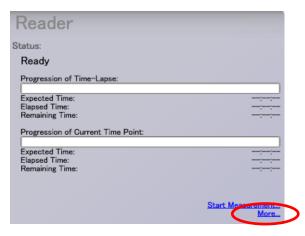


/ WARNING

In the case of replacing water in the bottles, make sure that the MAIN POWER breaker surely turns OFF to shut down CV7000. (Refer to 3.5) Be careful not to turn the breaker ON by oversight while at work.

Starting Stage Incubator

1) Click "More" at the bottom of the Status area on the Reader screen.



2) The "Device Console" tab opens in the top left of the "Reader Control" screen. (Refer to 6.4)

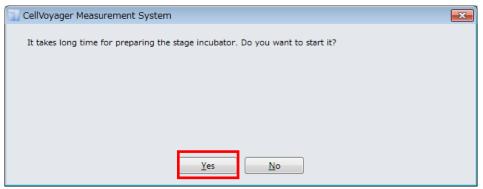
Input temperature, CO₂ density and gas flow rate (Only CV7000).

CV7000	enser Water CO2	
0 1 0 0 0		Control
	Get	Unload Well Plates Initialize Shut Down
Control Panel:	Alarm:	
Power On	Emergency Stop	Incubator
Hardware Status:	Access Gate	On Off Objective Lens © Water Immersion © Dry
Access Gate Open	O Tip Feeder Door	
O Tip Feeder Door Open	Temperature	Temperature: 37 🗘 °C
😑 Drive Unit Ready	Ventilation	CO2 Density: 5 🗘 %
Operation Possible	Air Supply	
Ready to Exchange Well Plate	Environment:	CO2 Flow Rate: 200 CO2 Flow Rate: 200
	Incubator	I confirmed that CO2 detector is working
CV7000S	penser Water CO2	Control
	Get	Unload Well Plates Initialize Shut Down
Control Panel:	Alarm:	Incubator
😑 Power On	Emergency Stop	Objective Lens
Hardware Status:	Access Gate	On Off Water Immersion O Dry
Access Gate Open	O Tip Feeder Door	
O Tip Feeder Door Open	Temperature	Temperature: 37 🗘 °C
😑 Drive Unit Ready	Ventilation	CO2 Density: 5 🗘 %
Operation Possible	Air Supply	
Ready to Exchange Well Plate	Environment:	I confirmed that CO2 detector is working
	ttable range	of CO ₂ density and gas flow rate is as
MEMO	lowing.	or CO ₂ density and gas now rate is as
	•	
- C	V7000	
ę	Settable rang	ge of CO ₂ density: $0.0 \sim 9.9\%$
(Settable rand	ge of gas flow rate: 0~500ml/min
· ·		je ol gas now late. Or soonin/min
- C	V7000S	
Ś	Settable rang	ge of CO ₂ denisty: $4.5 \sim 5.5\%$
5	Settable rang	ge of gas flow rate: Unsettable
		(fixed to optimal value)

3) Confirm that the CO₂ detector is operating. And then, check the "I confirmed that CO2 detector is working" checkbox.

Control Unload Well Plates Initialize Shut Down	Control Unload Well Plates Initialize Shut Down
Incubator On Off Objective Lens O Water Immersion O Dry	Incubator On Off Objective Lens O Water Immersion O Dry
Temperature: 37 🗘 *C CO2 Density: 5 🕉 %	Temperature: 37 ↓ *C CO2 Density: 5 ↓ %
CO2 Flow Rate: 200 C ml/min ✓ I confirmed that CO2 detector is working	☑ I confirmed that CO2 detector is working
CV7000	CV7000S

4) In case that following diagram is shown. Click "OK" and Stage Incubator preparation starts. (It takes about 10 minutes.)



5) Indicator of "Incubator" becomes green after water supplying to Stage Incubator finishes and temperature/ CO_2 density become stable. (It takes about 40 minutes.)

Status	Get
Control Panel:	Alarm:
😑 Power On	Emergency Stop
Hardware Status: Access Gate Open Tip Feeder Door Open Drive Unit Ready Operation Possible	 Access Gate Tip Feeder Door Temperature Ventilation Air Supply
Ready to Exchange Well Plate	Environment: Incubator

6) Present temperature and CO_2 density can be confirmed by selecting "Temperature" tab and "CO2" tab.

Status				Control
Target:	37 °C		Get	Target: 37 🗘 °C Set
Heater Temperat	ure:			Turn on/off Heater: Set
Objective 1	36.4 °C 🔵	Top of Assay Plate	36.4 °C 🔵	
Objective 2	36.4 °C 🥑	Middle of Assay Plate	36.4 °C 🔵	 Objective Top of Assay Plate
Objective 3	36.4 °C 🌖	Bottom of Assay Plate	36.4 °C 🔵	I TOP OF Assay Plate Middle of Assay Plate
Objective 4	36.4 °C 🔵	Source Plate Shuttle	36.4 °C 🔵	Bottom of Assay Plate
Objective 5	36.4 °C 🥚			Source Plate Shuttle
Objective 6	36.4 °C 🌖			Select All Deselect
		Dispenser Water CO2		
	emperature Tip		trol	
Status		Cor		
		Cor Get St	ntrol tage Incubator Supply On	Off
Status Stage Incubato	r	Cor Get St	age Incubator Supply On	Off 5 ♀ % Set

Finishing Stage Incubator

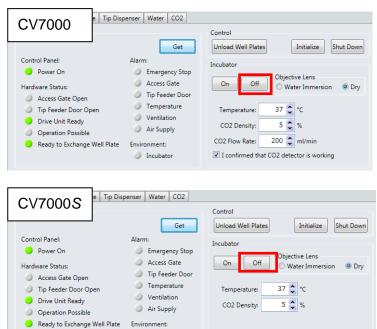
1) Click "More" at the bottom of the Status area on the Reader screen.

Reader	
Status:	
Ready	
Progression of Time-Lapse:	
Expected Time:	
Elapsed Time:	
Remaining Time:	
Progression of Current Time Point:	
Expected Time:	
Elapsed Time:	
Remaining Time:	
	Start Meaningment
	More

2) The "Device Console" tab opens in the top left of the "Reader Control" screen. (Refer to 6.4)

 \fbox{I} I confirmed that CO2 detector is working

Click "OFF".



Incubator

Confirming Log File for Stage Incubator

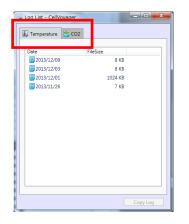
Each log file of temperature and CO2 concentration for stage incubator can be output.

1) To open log files for temperature and CO2 concentration, Select the "File" menu of the "Reader Control" screen and then click "Open Log List."

🐻 Re	eader Control - CellVoyager	
File	Edit View Measurement Help	
	Open Well Plate Information List	
	Open Measurement Setting List	
	Open Dispense Setting List	ser Tip
	Open Measurement Data List	
	Open Log List	
	Close	Emerg
	<u></u>	Access
	Hardware Status:	Tip Fee
	Access Gate Open	Tempe
	🥥 Tip Feeder Door Open 🥥	Interna
	Orive Unit Ready	Air Su

(2) The "Log List" screen is shown.

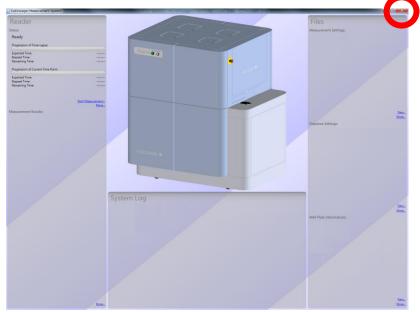
Select the "Temperature" tab or the "CO2" tab.



(3) Select the items you desire and click "Copy Log" to select destination to be output.

Temperature 📑 0	:02	
Date	FileSize	
2013/12/09	8 KB	
2013/12/03	8 KB	
2013/12/01	1024 KB	
2013/11/26	7 KB	
	(Copy Log

3.5. Shutting Down the Equipment



1) Close the screen of the application software.

- 2) Turn off the power of the measurement PC.
- 3) Press the POWER OFF button. The POWER ON lamp turns off.

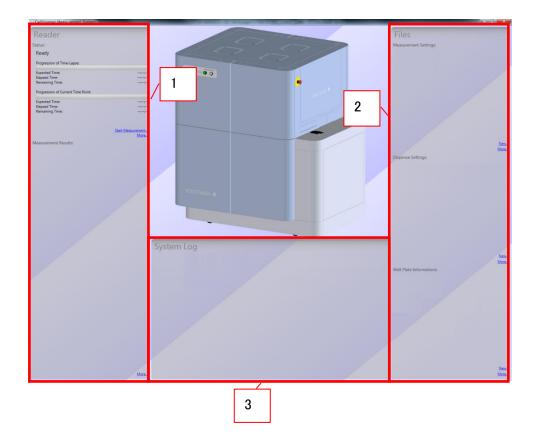


4) Turn off the MAIN POWER breaker on the rear side of the equipment.



3.6. Explanation of the Main Screen

The main screen of the portal application software is divided into the Reader area, Files area and log information display area.



1) Reader area (Refer to 6.1)

Well plates are set up for imaging and measured.

2) Files area (Refer to 6.1)

The past imaging history, dispenser settings and history of well plate information files are shown.

3) Log information display System logs are shown.

3.7. In Case of Emergency

This section explains how to stop the equipment immediately in case of emergency and how to reinitialize CV7000.

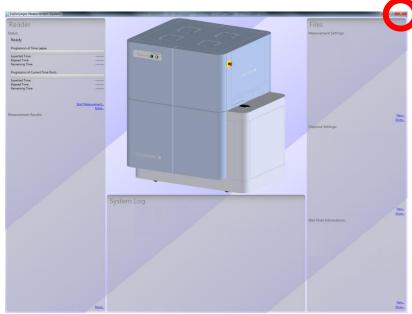
1) Press the EMERGENCY STOP button.



2) To release the EMERGENCY STOP button, turn it clockwise.



3) Close the application software. Additionally, turn off the power of the measurement PC.



4) Turn off the MAIN POWER breaker on the rear side of the equipment.

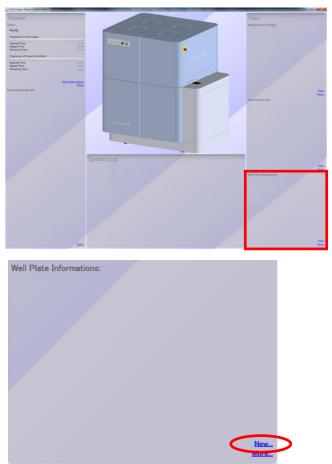


5) After 10 seconds, start CV7000. (Refer to 3.3.)

4. Entering Well Plate Information Files

It is not necessary to enter the well plate information files for measuring. (Entry is recommended.) But if dispensing is performed, this file must be created.

4.1. Creating a Well Plate Information File



1) Click "New" at the bottom of the Well Plate Informations area.

2) Select a desired well plate product. After a product has been selected, click "OK." (Refer to 4.3)

endor .	Name	Well Number	Shape	Bottom Material	Bottom Thickness	Usage
BD BD	#353219	96 wells	Round	Plastic	190 um	Source
BD BD	#353948	96 wells	Round	Plastic	880 um	Source
BD BD	#356663	384 wells	Rectangle	Plastic	380 um	Source
BD BD	#356936	384 wells	Rectangle	Plastic	380 um	Source
Corning	#3712	384 wells	Rectangle	Plastic	635 um	Source
Corning	#3904	96 wells	Round	Plastic	500 um	Source
PerkinElmer	#6004439	1536 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6005550	96 wells	Round	Plastic	190 um	Source
PerkinElmer	#6007430	384 wells	Rectangle	Plastic	190 um	Assay, Source
PerkinElmer	#6007439	384 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6007440	384 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6007450	384 wells	Rectangle	Plastic	190 um	Source
Greiner	#655090	96 wells	Round	Plastic	190 um	Source
Greiner	#655201	96 wells	Round	Plastic	990 um	Source
Greiner Greiner	#655896	96 wells	Round	Glass	175 um	Assay, Source
Greiner	#781091	384 wells	Rectangle	Plastic	190 um	Source
Greiner	#781896	384 wells	Rectangle	Glass	175 um	Source
					OK	Cancel

The screen for entering well plate information opens. (Refer to 4.3)

		#655896										
Dens	iity Unit:	mΜ	•									
	Usage:	Assay P	late 💿 Sor	urce Plate								
Des	cription:											
									-			
Name	Sample	olume Re	agent Name	Reagent	Volume	Graph Serie	s No. D	ose Amount	×			
	1	2	3	4	5	6	7	8	9	10	11	12
A												
в												
с						-						
D												
						-		-		-		
ε												
E F												

3) Enter the name of the well plate (file name).

```
Well Plate Name: #655896-20131003140723
```

Well Plate Type: #655896

4) Select "Usage."

Select "Assay Plate" if the well plate contains a cell sample to be measured. Select "Source Plate" if the well plate contains a reagent or other compound.

Usage: Assay Plate OSource Plate

5) Enter information regarding the well. (Refer to 4.2)

Item	Explanation	Remarks
Cell Name	Cell name	*1
Sample Volume	Amount of solution in the assay plate well (µI)	*2
Reagent Name	Reagent name. After imaging analysis, a density-dependent curve will be the drawn based on the reagent name entered here.	*3
Reagent Volume Amount of solution in the source plate well (µI)		*2
Graph Series No.	Graph number. When density-dependent curves are drawn after imaging analysis, wells of the same number are reflected as data points on the same graph.	*4
Dose Amount	Density of reagent in the assay plate. This information is reflected in the X-axis of the density-dependent curve after imaging analysis.	*4

Displayed Well Information Items

*1 : This information is not reflected in measurement or imaging analysis. (The field may remain blank.)

- *2 : This information must be entered if dispensing is performed.
- *3 : This information is displayed on the density-dependent curve. (Entry is recommended if CV7000 Analysis Software is installed.)
- *4 : This information must be entered properly to draw a desired density-dependent curve. (Entry is recommended if CV7000 Analysis Software is installed.)

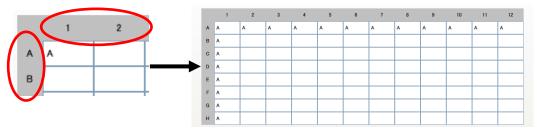
If the "Dose Amount" field is entered, select "Density Unit." (You can also use the default unit without entering a specific unit.)

Density Unit:	uМ	~
---------------	----	---

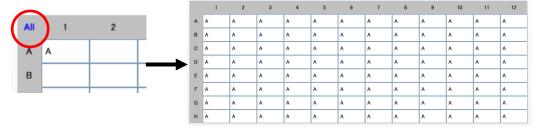
6) Click "Save."

4.2. How to Enter Well Information

Click the number or alphabet letter of the well to enter the number, character, or empty space at the beginning of the row or column of the well in the entire row or column.



Place the mouse pointer over the left upper corner of the well to display "All". Click to enter the number, character, or empty space you entered in cell A1 in all the remaining cells.



You can copy and paste the data you enter in the Excel worksheet.

Data entered in Excel worksheet

	-											
	1 1	2	3	4	5	6	7	8	9	10	11	12
A	0	1	3	10	30	100	300	1000	3000	1 0 0 0 0	30000	1 00000
в	0	1	3	10	30	100	300	1 0 0 0	3000	1 0 0 0 0	30000	1 00000
С	0	1	3	10	30	100	300	1 0 0 0	3000	1 0 0 0 0	30000	1 00000
D	0	1	3	10	30	100	300	1 0 0 0	3000	1 0000	30000	1 00000
E	0	1	3	10	30	100	300	1 0 0 0	3000	1 0 0 0 0	30000	1 00000
F	0	1	3	10	30	100	300	1 0 0 0	3000	1 0 0 0 0	30000	1 00000
G	0	1	3	10	30	100	300	1000	3000	1 0 0 0 0	30000	1 00000
Н	0	1	3	10	30	100	300	1000	3000	1 0 0 0 0	30000	1 00000

Paste

	1	2	3	4	5	6	7	8	9	10	11	12
А	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
в	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
С	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
D	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
Е	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
F	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
G	0	1	3	10	30	100	300	1000	3000	10000	30000	100000
н	0	1	3	10	30	100	300	1000	3000	10000	30000	100000

The following shows an entry example of well information.

• Entry Example of Reagent Names

A density-dependent curve is output for each reagent name you have entered. Leave the field blank for wells that are not used.

Cell Name	Sample	Volume	Reagent Na	me Rea	gent Volume	e Graph	Series No.	Dose A	mount 🔻			
	1	2	3	4	5	6	7	8	9	10	11	12
A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A
в	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A
С	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A
D	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A	Reagent A
E	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B
F	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B
G	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B
н	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B	Reagent B

Entry Example of Graph Series Numbers

When density-dependent curves are output after imaging analysis, wells of the same number are displayed as data points on the same graph. Enter "0" for wells that are not used.

Cell Na	ame	Sample \	Volume	Reagent Na	me Rea	igent Volume	Graph	Series No.	Dose A	mount 🔻			
	14	1	2	3	4	5	6	7	8	9	10	11	12
	А	1	1	1	1	1	1	1	1	1	1	1	1
	в	2	2	2	2	2	2	2	2	2	2	2	2
	С	3	3	3	3	3	3	3	3	3	3	3	3
	D	4	4	4	4	4	4	4	4	4	4	4	4
	Е	5	5	5	5	5	5	5	5	5	5	5	5
	F	6	6	6	6	6	6	6	6	6	6	6	6
	G	7	7	7	7	7	7	7	7	7	7	7	7
	н	8	8	8	8	8	8	8	8	8	8	8	8

• Entry Example of Dose Amounts

When density-dependent curves are output after imaging analysis, this information is reflected in the X-axis of each curve. Enter "0" for wells to which no reagent was added or wells that are not used.

Cell N	ame	Sample	Volume	Reagent Na	me Rea	igent Volume	Graph	Series No.	Dose A	mount 🔻			
		1	2	3	4	5	6	7	8	9	10	11	12
	A	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	в	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	с	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	D	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	Е	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	F	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	G	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100
	н	0.01	0.03	0.06	0.1	0.3	0.6	1	3	6	10	30	100

4.3. Well Plate Information File Screen

Well Plate Product Selection Screen

endor	Name	Well Number	Shape	Bottom Material	Bottom Thickness	Usage
BD BD	#353219	96 wells	Round	Plastic	190 um	Source
BD BD	#353948	96 wells	Round	Plastic	880 um	Source
BD BD	#356663	384 wells	Rectangle	Plastic	380 um	Source
BD BD	#356936	384 wells	Rectangle	Plastic	380 um	Source
Corning	#3712	384 wells	Rectangle	Plastic	635 um	Source
Corning	#3904	96 wells	Round	Plastic	500 um	Source
PerkinElmer	#6004439	1536 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6005550	96 wells	Round	Plastic	190 um	Source
PerkinElmer	#6007430	384 wells	Rectangle	Plastic	190 um	Assay, Source
PerkinElmer	#6007439	384 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6007440	384 wells	Rectangle	Plastic	190 um	Source
PerkinElmer	#6007450	384 wells	Rectangle	Plastic	190 um	Source
Greiner Greiner	#655090	96 wells	Round	Plastic	190 um	Source
Greiner	#655201	96 wells	Round	Plastic	990 um	Source
Greiner Greiner	#655896	96 wells	Round	Glass	175 um	Assay, Source
Greiner Greiner	#781091	384 wells	Rectangle	Plastic	190 um	Source
Greiner	#781896	384 wells	Rectangle	Glass	175 um	Source

- 1) Well plate manufacturer
- 2) Well plate type (model number)
- 3) Number of wells
- 4) Well shape
- 5) Material of well plate bottom
- 6) Thickness of well plate bottom
- 7) Well plate purpose

Well Plate Information File Screen

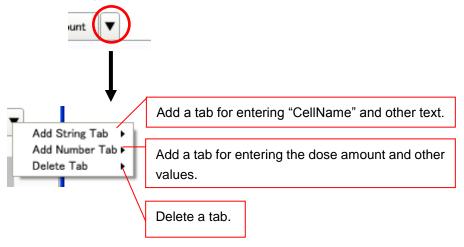
	ate Name: 🕴		2011082508543	33	2			1				
	sity Unit:	nM-	•		2							
3	Usage:	Q Assav	Plate 🔘 Sou	urce Plate								
	scription:		4									
			4			5						
		L										
Name	Sample V	olume	Reagent Name	Reagent	Volume	Graph Serie	s No. D	ose Amount	•			
_			-	-								
	1	2	3	4	5	6	7	8	9	10	11	12
A												
В												
С						6						
D						0						
E												
F												
G												
Н												
											Court	
									7		Save	Close
												/

- 1) Well plate information file name
- 2) Well plate type

(Clicking allows you to replace with other same well-number plate.)

- 3) Unit of dose amount (density of reagent in the assay plate)
- 4) Selection of well plate purpose (assay plate/source plate)
- 5) Memo
- 6) Well information

Selecting $\mathbf{\nabla}$ at the right edge of the tab opens the menu where you can add tabs for entering well information.



- 7) Save the well plate information file.
- 8) Close the well plate information file screen.

5. Using the Measurement Software

5.1. Measurement Software Functions

Time-lapse Setting

You can set a timeline to be used as the basis of measurement operation and processing or set time-lapse measurement for each plate. In time-lapse measurement for each plate, imaging of the same well plate is repeated at the specified interval.

You can add as many fluorescence imaging, software focus and other processes as desired for each timeline.

Well Plate Scan Setting

You can set an imaging well and imaging points.

To set an imaging well, select the assay plate well to be measured and save the setting information. To set imaging points, select imaging points in the well and movement pattern, and save the setting information. Imaging points are set in the following five modes.

Cell Count Function

The system moves through the imaging points to capture images repeatedly in the same well until the specified cell count is reached. When the number of repetitions reaches the specified value, imaging will stop and the system will move to the next well.

Cell Search Function

The system moves through the imaging points repeatedly until the specified count is reached, and outputs images of the specified number at the imaging point associated with the largest cell count, or imaging point closest to the specified cell count.

Tile Function

Images of the whole region of well (excluding edge areas or including edge areas) are captured with the specified object lens and tiled from the one corresponding to the top left of the well. A desired overlap between captured images can be specified in pixels.

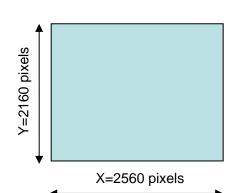
Partial Tile Function

Images of the partial region(s) of well (capable of plural regions setting) are captured with the specified object lens and tiled from the one corresponding to the top left of the well. A desired overlap between captured images can be specified in pixels. Fixed Position Function

Positions of imaging points are specified directly. You can specify desired imaging points or imaging points in a circular, rectangular or other pattern.

Magnification	Х	Y
4x	4160µm	3510µm
10x	1664µm	1404µm
20x	832µm	702µm
40x	416µm	351µm
60x	277µm	234µm

Magnification factors and corresponding view fields of the camera



View field of the camera in pixels

Action List

You can set various measurement operations including software focus, fluorescence imaging, bright field/phase contrast imaging, 3D imaging and dispensing.

Software Focus (Imaging Auto-focus)

Images are captured repeatedly at a specified wavelength while moving the Z position on the confocal plane to determine the Z position on the confocal plane at the position of the brightest image. The software focus setting has no imaging function. The software focus function is inter-dependent with fluorescence imaging and 3D imaging, so when the software focus function was used, you can perform fluorescence imaging and 3D imaging successively from the condition of the software focus function being executed.

Fluorescence Acquisition

Fluorescent light from the target is captured. Multiple cameras are used to capture images simultaneously at one or more fluorescent wavelengths. To acquire images for each wavelength during measurement where two or more wavelengths are used, add two or more fluorescence imaging sessions and select a desired measurement wavelength in each session. You can capture soft focus images by setting fluorescence imaging immediately after

implementing the software focus function. If software focus is skipped and only fluorescence imaging is set, auto-focus imaging can be performed.

Auto-focus Imaging

One shot imaging is performed without software focus.

Software focus imaging

One shot imaging is performed with software focus.

Epifluorescence Imaging

An optical circuit detouring around the confocal unit is used. The laser is used as the light source to capture images in the epifluorescence mode.

High-speed Time-lapse Imaging

Time-lapse measurement is performed for each well at an interval of several tens to several hundreds of milliseconds. In time-lapse measurement for each well, imaging of the same well is repeated at the specified interval. Time-lapse imaging of one well is completed before imaging of the next well is performed. During high-speed time-lapse imaging, dispensing can be performed at a specified timing. (Dispenser model only)

3D Fluorescence Acquisition

This is a function whereby a Z-position imaging area is specified and fluorescence images are captured over the entire Z plane by moving the Z position on the confocal plane. Fluorescent images with one or more wavelengths are captured. To acquire images for each wavelength during measurement where two or more wavelengths are used, add two or more 3D imaging sessions and select a desired measurement wavelength in each session. You can perform 3D imaging using the software focus plane as the reference, by setting 3D imaging immediately after implementing the software focus function. If software focus is skipped and only 3D imaging is set, 3D imaging can be performed using the auto-focus plane as the reference.

In regard to captured 3D images, a single image projected in the Z-axis direction also can be created.

Maximum

You can perform MIP on captured 3D images. MIP, or Maximum Intensity Projection, is a function to build a single image by putting together pixels associated with the largest signals among identical pixels from multiple image data captured on the confocal plane in the Z-axis direction. This way, an object that cannot fit a single confocal image can be fully depicted by capturing its 3D images and synthesizing them via MIP.

Minimum

You can perform MinIP on captured 3D images. MinIP, or Minimum Intensity Projection, is a function to create a single image by putting together pixels associated with the smallest signals among identical pixels from multiple image data captured on the confocal plane in the Z-axis direction. This function is effective for bright-field images captured by 3D imaging.

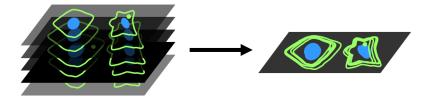
Average

You can perform AIP on captured 3D images. AIP, or Average Intensity Projection, is a function to create a single image by putting together pixels associated with the average signals among identical pixels from multiple data captured on the confocal plane in the Z-axis direction.

Average values are obtained by dividing the total pixel intensity by the number of Z images.

Sum

SUM is a function to create a single image by putting together pixels associated with the summation signals among identical pixels from multiple image data captured on the confocal plane in the Z-axis direction. To calculate summation intensity, the value which subtracted the background of the camera is summed from the 2nd sheet. Imageable intensity is up to 65525.



Z-axis projection

Bright-field/Phase-contrast Acquisition (Bright field/ phase contrast model only)

This is a function whereby imaging is performed based on the bright field and phase contrast by using a lamp as the light source. In the phase contrast mode, the object lens must be changed to one for phase contrast imaging.

Auto-focus Imaging

One shot imaging is performed without software focus.

High-speed Time-lapse Imaging

Time-lapse measurement is performed for each well at an interval of several tens to several hundreds of milliseconds. In time-lapse measurement for each well, imaging of the same well is repeated at the specified interval. Time-lapse imaging of one well is completed before imaging of the next well is performed. During high-speed time-lapse imaging, dispensing cannot be performed.

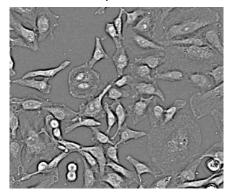
Z-Stack Bright-field/Phase-contrast Acquisition (Bright field/ phase contrast model only)

This is a function to capture Z-stack bright field and phase contrast images by moving the Z position on the focal plane. This is captured with other timing separated from fluorescence acquisition. In the phase contrast mode, the object lens must be changed to one for phase contrast imaging.

In regard to captured Z-stack images, a single image projected in the Z-axis direction also can be created in common with 3D Fluorescence Acquisition.

DPC (Digital Phase-contrast) Acquisition (Bright field model only)

This is a function to capture 2 Z position images by using a lamp as the light source and make phase contrast like image (Phase type) or fluorescence like image (Fluor type). This is captured with other timing separated from fluorescence acquisition.



Phase type DPC image

Fluor type DPC image

Dispensing Operation (Dispenser model only)

This is a function whereby reagent is dripped using a dispenser. Dispensing is performed for each plate, meaning that reagent is dispensed into all specified wells in one well plate.

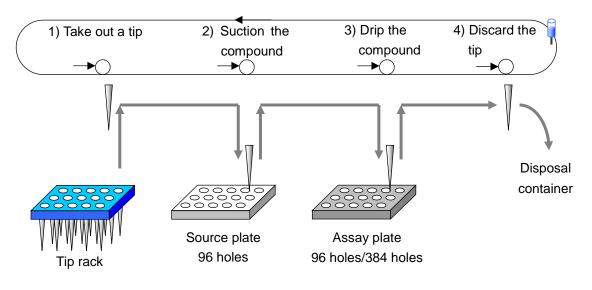
Since dispensing is not associated with any imaging function, separate measurement operations must be added if images are to be captured before and after dispensing.

Dispensing Function

Dispensing Mechanism

With the CellVoyager, dispensing refers to the operations such as suctioning a specified volume of a compound from the source plate that contains the compound and dripping the compound onto the assay plate. The device that performs dispensing is called "Dispenser."

The dispenser of the CellVoyager performs dispensing using disposable tips one by one. After each tip has been used, the dispenser replaces the tip with a new one and performs the next dispensing action. The figure below illustrates the operation flow of the dispenser.



Overview of Dispensing Mechanism

1) Taking out a tip

An unused tip is taken out from the tip rack. After all unused tips on the tip rack have been used, the tip rack is replaced with one containing unused tips.

2) Suctioning the compound

The edge of the tip is moved to above a specified well in the source plate containing the compound, and the compound is suctioned from the well by a specified volume.

3) Dripping the compound

The tip containing the compound is moved to above a specified well in the assay plate, and the compound is dripped into the well.

4) Discarding the tip

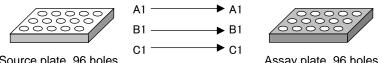
After all compound has been dripped onto the assay plate, the used tip is moved to the tip disposal location and is disposed of as a waste.

Dispensing Pattern

Since the number of wells in the source plate does not always match the number of wells in the assay plate, the following examples of combinations are to be considered. Only one source plate can be set, while up to four assay plates can be set.

1) 96-hole Source plate \rightarrow 96-hole Assay plate

This is the simplest pattern where the wells in one plate have a one-to-one association with the wells in the other plate.

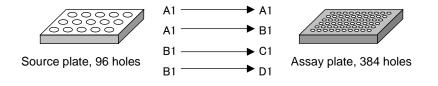


Source plate, 96 holes

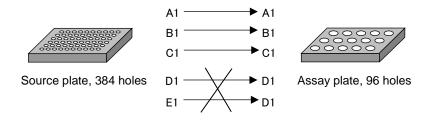


2) 96-hole Source plate \rightarrow 384-hole Assay plate

In this combination, one well in the source plate is associated with multiple wells in the assay plate.

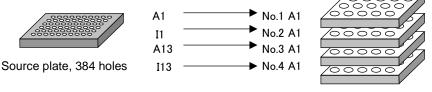


3) 384-hole Source plate \rightarrow 96-hole Assay plate Dispensing from the multiple wells in the source plate to one well in the assay plate cannot be performed.



4) 384-hole Source plate \rightarrow 96-hole Assay plate x 4

In this combination, one well in the source plate is associated with one well in one of the assay plates. This dispensing pattern is selectable when a large incubator (sold separately) is used.



Assay plate, 96 holes

Dispensing Actions

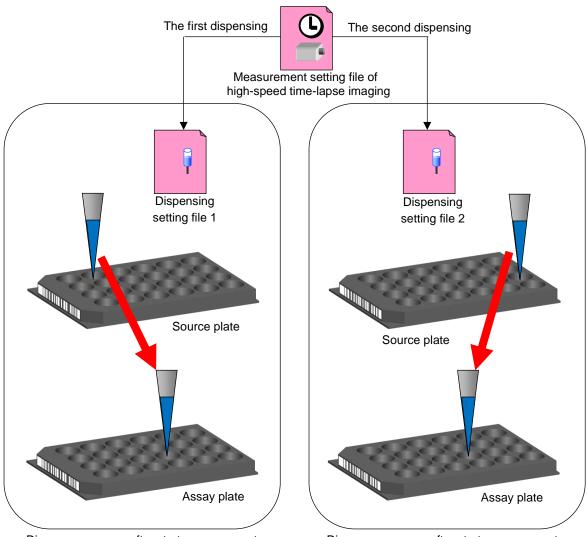
Ten steps of dispensing actions, including filling, dripping and agitation, can be set for each dispensing operation performed between source and assay plates. Detailed parameters such as the dripping speed and amount, and whether or not to implement agitation, can be set for each dispensing action.

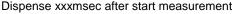
List of dispensing actions							
Dispensing action	Explanation						
	The liquid surface is detected. The tip is lowered within						
Liquid Surface	the well in the source plate until its edge contacts the						
	liquid, to detect the position of liquid surface in the well.						
Prewet	Solution is filled and then dripped within the well in the						
Flewel	source plate to wet the inside of the tip with the solution.						
	Solution is agitated within the well in the source plate.						
AspirateStir	Solution is suctioned and discharged for the specified						
	number of times.						
Airgap	Air is introduced into the tip before solution. This way, all						
Ліуар	solution can be dripped.						
Aspirate	Solution is filled from the well in the source plate.						
AspirateTiptouch	The tip is caused to contact the wall of the well to remove						
Aspirate riptoden	any water droplet attached at the edge of the tip.						
Dispense	Reagent is dropped onto the assay plate.						
	Dripped reagent is agitated in the well. Solution is						
DispenseStir	suctioned and then discharged within the well for the						
	specified number of times.						
DispenseTiptouch	The tip is caused to contact the wall of the well to remove						
Dispense riptoden	any water droplet attached at the edge of the tip.						
Air Blow	This allows to blow air to drop droplet adherent on the						
	tip top.						

of dia

Multi Dispensing

In high-speed time-lapse imaging, it is possible to dispense from wells from source plate to one well of assay plate. Disposable tips are exchanged after each dispensing is performed. Before you do multi dispensing, you make setting files of each dispensing. And when you start measurement, you assign these setting files and dispensing is done as the order which you assign them.





Dispense yyymsec after start measurement

Working of imaging with multi dispensing

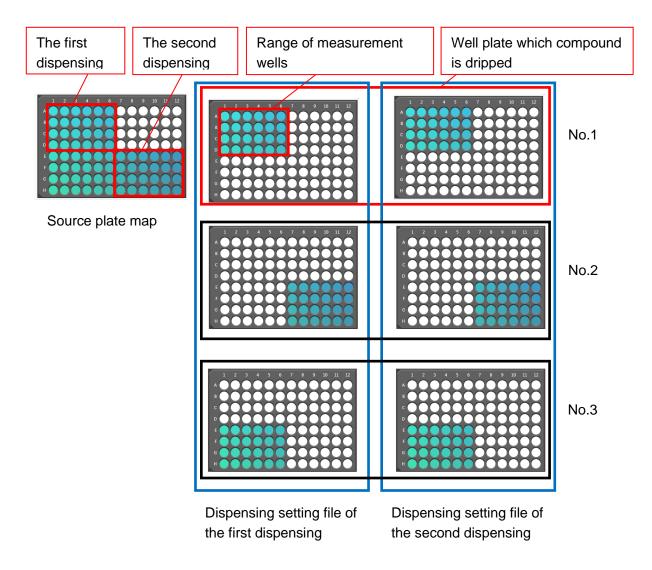
In high speed time lapse imaging, wells for imaging is decided as setting of imaging region in measurement setting file. Settings of multi dispensing need to meet following condition.

•Make dispensing setting file of each dispense.

- (For example, if you perform three times of dispensing, three dispensing setting files is needed)
- Well numbers of well-plate map of source plates must be set as same in all dispensing setting files.
- Well numbers of all well-plate maps of assay plates must be set as same in

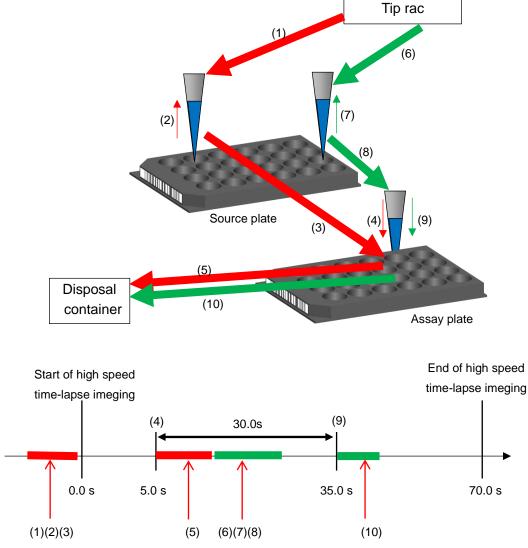
all dispensing setting files.

- •Wells which compound is dripped of each well-plate map number of assay plate must be set as same in all dispensing setting files.
- Number of well-plate map of assay plate must be set as same in all dispensing setting files.
- Measurement wells assigned in measurement setting file must be selected from the well-plate map of dispensing setting file.
- ●Only one well plate which compound is dripped can be assigned from well-plate map (No.1 No.4) of assay plate which is made in dispensing setting file. It is impossible to dispense to multiple assay plates.
- In case of multi dispensing with robot handling system, No. 1 well-plate map is applied as well-plate which compound is dripped from well-plate map of assay plate which is made in dispensing setting file.
- ●30 seconds of intervals are needed between procedures of multi dispensing



Assay plate map

Example of multi dispensing setting



Exapmle of measurement with multi dispensing

(1) Take out a tip from tip rack

(2) Suction the compound from source plate

(3) Move to assay plate

「Start high-speed time-lapse imaging」

(4) Drip the compound to assay plate 5 second after starting of high-speed time-lapse imaging

- (5) Discard the tip
- (6) Take out the new tip from tip rack
- (7) Suction the other compound from source plate
- (8) Move to assay plate

(9) Drip the compound to assay plate 35 second after starting of high-speed time-lapse imaging

(10) Discard the tip

[70 second after start imaging, stop high-speed time-lapse imaging]

Imaging channels

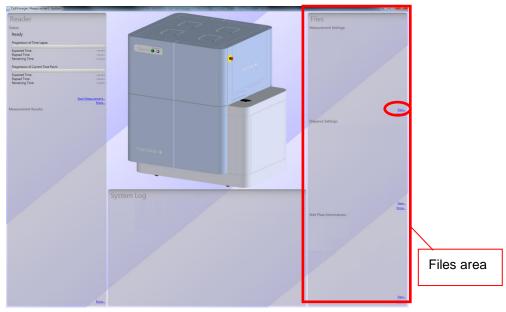
On the Image Setting screen, imaging channels are set. A combination of object lens, light source, filter wheel and camera settings is stored for each imaging channel.

Item		Explanation		
Ch		Number to identify the imaging channel		
Target		Specify the name of the target to be imaged.		
Method		Select the combination of filter and light source.		
Objective		Select the object lens.		
Acquisition		Select the filter name to be used for imaging. A filter name list is generated from the filters selected on the filter wheel.		
ExposureTime		Specification of exposure time. Unit : ms		
Binning		Select the binning. 1x1, 2x2, 3x3 or 4x4.		
LightSource		Select the laser as the light source. If the laser is selected, multiple wavelengths can be combined.		
Fluorophore		Select the used fluorophre. (It is needed for performing crosstalk correction by Image Correction Software)		
Preview	Color	Specify the display color for preview in the #rrggbb format. Default color is subject to filter setting selected by "Acquisition."		
	MinLevel	Specify the lower limit of input signals for generating a preview in a range of 0.0 to 1.0.		
	MaxLevel	Specify the upper limit of input signals for generating a preview in a range of 0.0 to 1.0.		

List of so	etting items	for each	impaina	channel
	sung liems	IUI Each	inaging	Channer

5.2. Displaying the Measurement setting File Screen

Creating a New Measurement Setting File

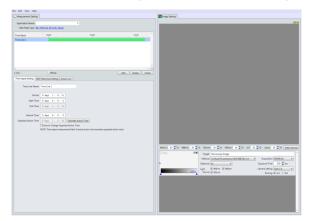


1) Click "New" in the Files area.

2) Select a desired well plate product and click "Create a New Measurement Setting."

Vendor	Name	Well Number	Shape	Bottom Material	Bottom Thickness	Usage
MUNC NUNC	#164588 (low bottom)	96 wells	Round	Glass	190 um	Assay, Source
MUNC	#1645xx	96 wells	Round	Glass	190 um	Assay, Source
PerkinElmer	#6007430	384 wells	Rectangle	Plastic	190 um	Assay, Source
Greiner Greiner	#655896	96 wells	Round	Glass	175 um	Assay, Source

The screen for editing a measurement setting files opens. (Refer to 6.2)



* If the plate you wish to use is not on the list, register the plate by referring to 5.16.

Editing a Measurement Setting File

1) Click "More" at the bottom of the Measurement Settings section in the Files area.



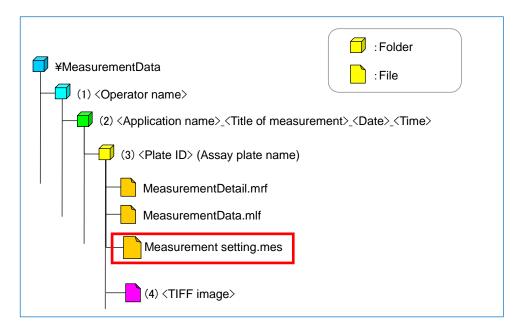
2) Select a desired measurement setting file and click "Open."



Click "Open External File" if you open the measurement setting file from the external folder you saved.



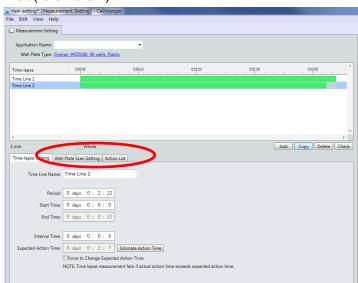
Select the file whose extension name of ".mes". The measurement setting file is shown.



5.3. Setting a Time Line

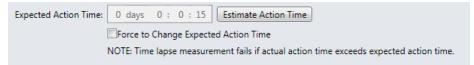
Each measurement setting file retains various setting information for measurement processing along a time line. For each time line, set the well movement pattern, imaging points and measurement actions such as fluorescence imaging and 3D imaging.

1) Set the items on the Well Plate Scan Setting tab (refer to 5.6) and Action List tab(refer to 5.7).

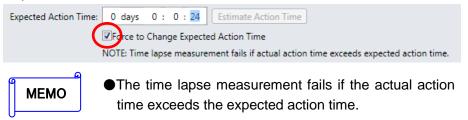


2) Click the Time-lapse Setting tab. (Refer to 6.2)

3) Click "Estimate Action Time" to display the expected time of measurement.



The Expected Action Time field can be entered if the Force to Change Expected Action Time checkbox is checked.



4) Enter the name of the time line. (You can use the default name.)

Time Line Name: Time Line 1

5) To perform time-lapse imaging (refer to 5.1) for each well plate, enter the imaging interval. Enter a value greater than the one shown in the Expected Action Time field.

Interval Time:	0	days	0	:	0	:	0	

6) Enter the start time of time line.

Start Time:	0 days	0:	0:	0	
-------------	--------	----	----	---	--

7) Enter the end time of the time line.

Period: 0 days	0 : 0 : 15
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8) Click "Add," and a new time line will be created. Select Time Line 2 and set the items on the Well Plate Scan Setting tab and Action List tab, as well as time-line timings, for Time Line 2.

0:00:00 Time-lapse 0:00:30 0:01:00 Time Line 1 Time Line 2 Delete time line Copy time line Add time line Whole 1 min Add Сору Delete Check Olick "Copy" button, and same time line as selected is MEMO copied to the last of list. In case that dispensing setting is included in selected

(Click "Copy" to copy time line. Click "Delete" to delete.)

9) Click "Check," and overlaps of time lines will be adjusted automatically. You can extend or shorten each time line using the scale bar.

time line, it is impossible to "Copy".

Time-lapse	0:00:00	0:00:30	0:01:00	
Time Line 1				
Time Line 2				
	Scale bar			
4				
min	Whole		Add Copy Delete	e Check
. min	whole		Add Copy Delete	Check

5.4. Entering the Application Name

Enter the application name such as "Granularity".

A desired name can be entered. (The field may remain blank.)



5.5. Setting the Imaging Channel

Set the object lens, light source, filter, CSU and sCMOS camera.



Image channel settings may be different by specifications. Please confirm your specification items for CV7000.

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1) Click "Add Channel" and specify the imaging channel number. (Refer to 6.2.)

		Create a new imaging channel.	
	405nm: 30 🗢 % 488nm: 30 🗙 %	561nm: 30 76 040nm: 30 76 UV: 30 76 Lamp: 30 8 Add Channel	\triangleright
	Ch 1	Target: Microscope Image	•
	/-	Method: Confocal Fluorescence 405/488/561/640 ni 🔹 Acquisition: BP445/45 🔹	
Dele	ete an imaging channel	hore: (Not Specified) Exposure Time: 250 🗘 ms	
Don	ete an inaging chamer	ctive: 10x •	
	10000 L	ght	

2) Enter the name of the imaging target.

Target: Ch1 Microscope Image

3) Select the combination of optical systems used for imaging.

Method: Confocal Fluorescence 405/488/640 nm

Optical system	Explanation
Confocal Fluorescence 405/488/561(or 532)nm	Confocal imaging using a light source
	of 405, 488 or 561(or 532) nm
Confocal Fluorescence 405/488/640nm	Confocal imaging using a light source
	of 405, 488 or 640 nm
Confocal Fluorescence 405/488/561/640nm	Confocal imaging using a light source
(QUAD-DM model only)	of 405, 488, 561 or 640nm
Epifluorescence 405/488/561(or 532)nm	Epifluorescence imaging using a light
	source of 405, 488 or 561(or 532) nm
Epifluoroscopco 405/488/640pm	Epifluorescence imaging using a light
Epifluorescence 405/488/640nm	source of 405, 488 or 640 nm
Epifluorescence 405/488/561/640nm	Epifluorescence imaging using a light
(QUAD-DM model only)	source of 405, 488, 561 or 640nm
Epifluorescence UV Lamp	LIV light source
(UV lamp model only)	UV light source

Brightfield	Brightfield imaging to match pixel
(Bright field model only)	position with epifluorescent imaging
Brightfield(Confocal path)	Brightfield imaging to match pixel
(Bright field model only)	position with confocal imaging
Phase Contrast (Phase contrast model only)	Phase contrast imaging to match pixel position with epifluorescent imaging.
Phase Contrast(Confocal path)	Phase contrast imaging to match
(Phase contrast model only)	pixel position with confocal imaging
Digital Phase Contrast	DPC imaging to match pixel position
(Bright field model only)	with epifluorescent imaging.

4) Select the fluorophore

Fluorophore:	MitoTracker Deep Red 🔹
Objective:	(Not Specified) Hoechist33342
Light	mKusabira Orange2
Source:	Azami Green
	Alexa Fluor 488
	MitoTracker Deep Red
	DAPI
	FusionRed_spectra

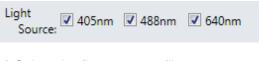


- If crosstalk correction will be performed by Image Correction Software, fluorephore must be selected,
- Refer to 5.15 about registration of fluorophore.

5) Select the object lens.

Objective: 10x 🔻	•
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6) Select the light source used for imaging.



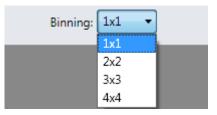
7) Select the fluorescence filter.

Acquisition:	BP445/45 🔹
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8) Set the exposure time. (11ms≦Exposure Time≦9999ms)

Exposure Time: 250 🗢 ms

9) Set camera binning.



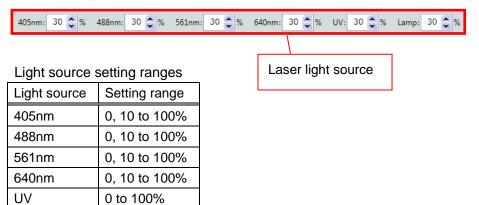
MEMO

Lamp

If multiple channels have been set, it is recommended to use the same binning setting for all channels. However CV7000 Analysis Software supports different binning setting on multiple channels, errors in recognition result can occur.

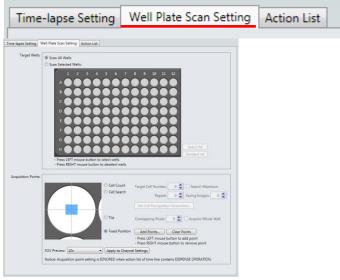
10) Set the light source output.

0 to 100%



5.6. Setting Acquisition Points in the Well

1) Set the well acquisition points. Click the Well Plate Scan Setting tab.

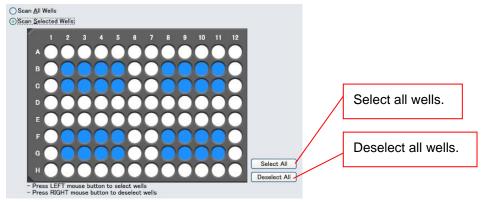


2) Select the wells to scan.

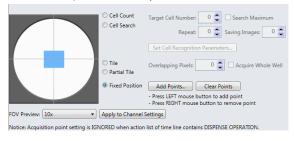
Scan All Wells	Scan all wells.	
Scan Selected Wells:	Select the wells	to scan.

When you select Scan Selected Wells, select the wells to scan. Left-click to select a well.

Right-click to deselect a well.



 Set acquisition points in the well. Acquisition points can be set in one of four modes including "Cell Count," "Cell Search," "Tile", "Partial Tile" and "Fixed Position." (Refer to 5.1)



Cell Count Function

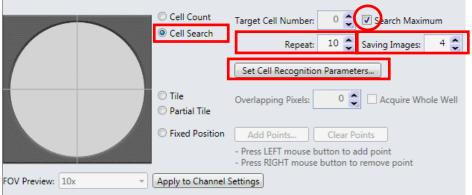
Cell Count Cell Search	Target Cell Number: 100 😜 🗌 Search Maximum Repeat: 10 📚 Saving Images: 0 📚	
	Set Cell Recognition Parameters	
○ Tile ○ Partial Tile	Overlapping Pixels: 0 💭 🗌 Acquire Whole Well	
© Fixed Position	Add Points Clear Points Press LEFT mouse button to add point	
- Press RIGHT mouse button to remove point FOV Preview: 10x Apply to Channel Settings		
Notice: Acquisition point setting is IGNORED when action list of time line contains DISPENSE OPERATION.		

- 1) Select "Cell Count." (Refer to 5.1 and 6.2)
- 2) Enter the value of "Target Cell Number" (specified cell count).
- 3) Enter the value of "Repeat" (maximum number of images to be captured).
- 4) Click "Set Cell Recognition Parameters." The screen for setting the cell recognition algorithm opens.

Set Cell Recognition Parameters			
Algorithm:	Connection		
Threshold:	1000 🗘		
Remove the fragments of the cell at the edge of the image			
Measure:	Mean Cell Diameter: 15 💭 um		
	Minimum Scaling Factor: 0.5 💭 (7.5 um)		
	Maximum Scaling Factor: 2 🗘 (30.0 um)		
OK Cancel			

5) Set the cell recognition algorithm. (Refer to 5.10)

Cell Search Function



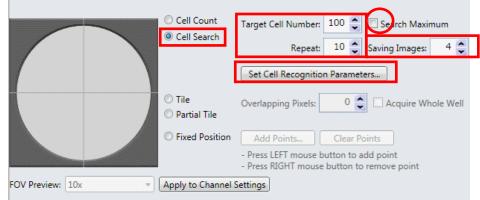
Save the image associated with the largest cell count

- 1) Select "Cell Search." (Refer to 5.1 and 6.2)
- 2) Select "Search Maximum."
- 3) Enter the value of "Repeat" (number of images to be captured)
- 4) Specify the value of "Saving Images" (number of images to be saved).
- 5) Click "Set Cell Recognition Parameters." The screen for setting the cell recognition algorithm opens.

1	Set Cell Recognition Parameters			
	Algorithm:	Connection		
	Threshold:	1000 🗘		
ł	Remove the fragments of the cell at the edge of the image			
	Measure:	Mean Cell Diameter: 15 🗣 um		
ł		Minimum Scaling Factor: 0.5 💭 (7.5 um)		
ł		Maximum Scaling Factor: 2 📚 (30.0 um)		
	OK Cancel			

6) Set the cell recognition algorithm. (Refer to 5.10)

Notice: Acquisition point setting is IGNORED when action list of time line contains DISPENSE OPERATION.



Save the image closest to the specified cell count

Notice: Acquisition point setting is IGNORED when action list of time line contains DISPENSE OPERATION.

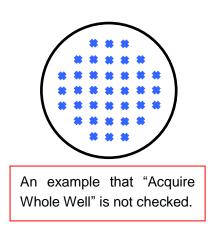
- 1) Select "Cell Search." (Refer to 5.1 and 6.2)
- 2) Unselect the "Search Maximum" check box.
- 3) Enter the value of "Target Cell Number" (specification of cell count).
- 4) Enter the value of "Repeat" (number of images to be captured).
- Specify the value of "Saving Images" (number of images to be saved). Images will be saved one by one, starting from the image closest to the specified cell count.
- 6) Click "Set Cell Recognition Parameters." The screen for setting the cell recognition algorithm opens.

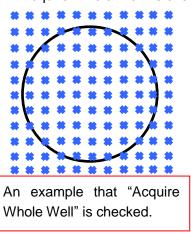
Set Cell Recognition Parameters				
Algorithm:	Connection •			
Threshold:	1000			
Remove the fragments of the cell at the edge of the image				
Measure:	Mean Cell Diameter: 15 🗣 um			
	Minimum Scaling Factor: 0.5 🗢 (7.5 um)			
	Maximum Scaling Factor: 2 🗘 (30.0 um)			
OK Cancel				

7) Set the cell recognition algorithm. (Refer to 5.10)

Cell	Target Cei Number:
© Tile © Parti © Fixed	al Tile Overlapping Pixels: O Clear Points - Press LEFT mouse button to add point - Press RIGHT mouse button to remove point
FOV Preview: 10x - Apply to	o Channel Settings

- 1) Select "Tile." (Refer to 5.1 and 6.2)
- 2) Enter the value of "Overlapping Pixels" (number of overlapping pixels of images). To have images overlap with each other, enter a positive value. To keep images apart, enter a negative value. (-100≦Overlapping Pixels≦100) Recommended value of Overlapping Pixels is "50".
- 3) The fields of whole well are acquired if "Acquire Whole Well" is checked.





Reference numbers of images captured by the tile function 1...

excluding edge areas (per well)			
Magnification	on 96 wells 384 wells		
10x	6 images	-	
20x	36 images	6 images	
40x	194 images	48 images	
60x	470 images	130 images	

Reference numbers of images captured by the tile function

including edge areas (per well)			
Magnification	96 wells	384 wells	
10x	20 images 6 images		
20x	80 images	20 images	
40x	304 images	80 images	
60x	696 images 168 images		

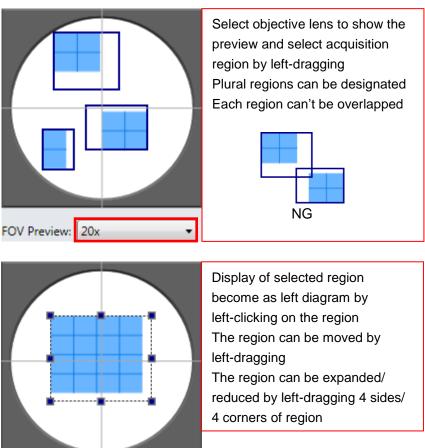
including edge areas (per -11

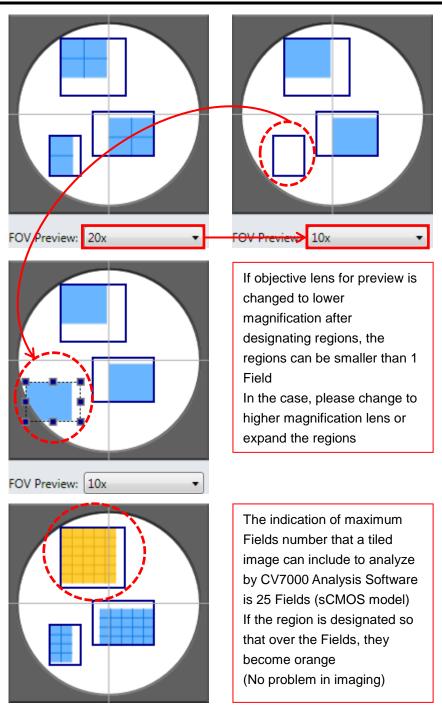
Partial Tile Function

Cell Count	Target Cell Number: 0 💭 🗆 Search Maximum
	Repeat: 0 Saving Images: 0 Set Cell Recognition Parameters
◯ Tile	Overlapping Pixels: 0 🖨 🗆 Acquire Whole Well
© Fixed Position	Add Points Clear Points - Press LEFT mouse button to add point - Press RIGHT mouse button to remove point
OV Preview: 20x Apply to Channel	·

- 1) Select "Partial Tile." (Refer to 5.1 and 6.2)
- Enter the value of "Overlapping Pixels" (number of overlapping pixels of images). To have images overlap with each other, enter a positive value. To keep images apart, enter a negative value. (-100≦Overlapping Pixels≦100) Recommended value of Overlapping Pixels is "50".
- 3) Designate acquisition region

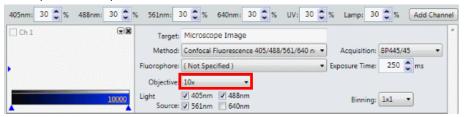
You can left-drag the well to specify desired acquisition region. Right-drag the selected region to delete the region.

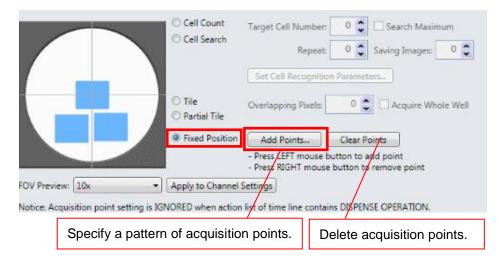




FOV Preview: 10x	FOV Preview. 60x W		
The restriction in imaging region can exist in some sample vessels when water immersion objective lens is used. In this case, prohibited area is displayed as pink. (refer to right of above diagram) Prohibited area s are unselectable as imaging region. However, if objective lens for preview is changed to water immersion type from dry type after designating regions, the regions can be in prohibited area. In the case, please reset the imaging regions.			

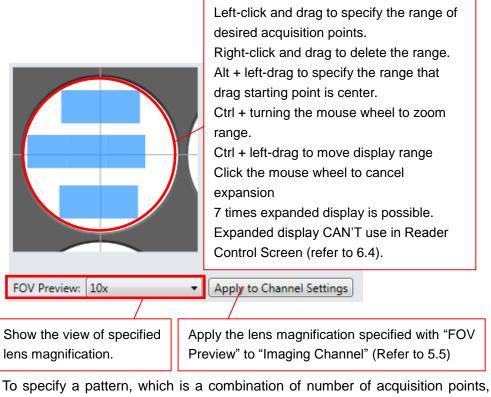
Objective lens selected in above procedure is to preview for setting region of tiling. Select actual objective lens used for measurement in channel setting window. (refer to 5.5).





- 1) Select "Fixed Position." (Refer to 5.1 and 6.2)
- 2) Specify the acquisition points.

You can left-click the well to specify desired acquisition points. Right-click an acquisition point to delete the point.

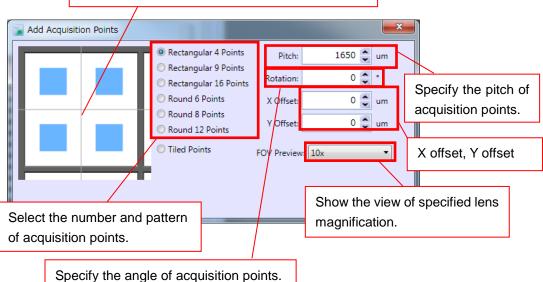


interval, etc., click "Add Points."

Fixed Position	Add Points	Clear Points
		button to add point e button to remove point

3) After clicking "Add Points," set the number, pattern, pitch and angle of imaging points. After all items have been set, click "Add Points" again.

Ctrl + turning the mouse wheel to zoom range.Ctrl + left-drag to move display rangeClick the mouse wheel to cancel expansion7 times expanded display is possible.



If selecting "Tiled Points," set as acquisition points the view field to be captured at a specified magnification factor in a tiled pattern. (Acquisition points can be added or deleted after clicking "Add Points.")

Add Acquisition Points			— X —
	 Rectangular 4 Points Rectangular 9 Points Rectangular 16 Points Round 6 Points Round 8 Points Round 12 Points Tiled Points 	Pitch: Rotation: X Offset: Y Offset: FOV Preview	3290 ♀ um 0 ♀ ° 0 ♀ um 0 ♀ um 10 ♀

The restriction in imaging region can exist in some sample vessels when water immersion objective lens is used. In this case, prohibited area is displayed as pink. (refer to right of above diagram)

Prohibited area is unselectable as imaging region. However, if objective lens for preview is changed to water immersion type from dry type after designating regions, the regions can be included in prohibited area. In the case, please reset the imaging regions.

Add Acquisition Points				×
	Rectangular 4 Points	Pitch:	7000	um
 Rectangular 9 Poin Rectangular 16 Poin 		Rotation:	0	•
		X Offset:	1000	um
	Round 6 Points	Y Offset:	0	um
	Round 8 Points Round 12 Points	FOV Preview:	60× W	
		TOV FIEVIEW.	002.00	
	Tiled Points			
			Add Points	Cancel

In case that select "Tiled Points" when imaging prohibited region exists, tiled region is selected inside acquirable region as below diagram.

 ○ Rectangular 4 Points ○ Rectangular 9 Points ○ Rectangular 16 Points ○ Rectangular 16 Points X Offset: 1000 C um 	<u>^</u>
 Round 6 Points Round 8 Points Round 12 Points FOV Preview: 60x W 	
Tiled Points	
Add Points Cancel	el

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Difference between the Tile Function and "Tiled Points"

The tile function lets you capture the entire view field of the well in a tiled pattern using the object lens selected when the imaging channel was set (refer to 5.5). You can also specify the number of pixels to overlap between tiled images.

With "Tiled Points," specify as acquisition points the view field to be captured at a specified lens magnification factor in a tiled pattern. You can also add or delete acquisition points.

5.7. Settings on the Action List Tab

Software Focus Setting

Refer to 5.1 for the software focus function.

1) Set the imaging channel. Under "Method," select "Confocal Fluorescence." (Refer to 5.5)

🗌 Ch 1 🔍 🖼	Target:	Microscope Image			
	Method:	Confocal Fluorescence 405/488/561/640 n 💌	Acquisition:	BP445/45	-
	Fluorophore:	(Not Specified) 🔹	Exposure Time:	250 🌲 ms	
	Objective:	10x •			
10000		🛿 405nm 📄 488nm 📄 561nm 📄 640nm	Binning:	1x1 •	

2) Click the Action List tab.

[Time-lapse Setting	Well Plate Scan Setting	Action List

3) Click "Software Focus."

Fuorescence	/ Ph isition Z-Stack BF / Ph Acquisition Acquisition DPC Acquisition Operation
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The screen for setting the software focus opens. (Refer to 6.2)

ACT: 0001	Target Cell	Shifting Distance:	5 😂 um
	Slicing Interval Ascending Distance (+)	Ascending Distance:	5 🤹 um
	Shifting Distance Descending Distance (-)	Descending Distance:	-5 🗘 um
Software	Bottom of Well Plate	Slicing Interval:	1 🤤 um
Focus	Software Focus Target:	X Offset:	0 🗘 um
		Y Offset:	0 😂 um
	Please select the target.	elect Test	

4) Click "Select."

Software Focus Target:	
Please select the target.	Select Test

5) The Select Channel screen opens. Select the target channel, and then click "OK."

Select Channel				
П	Target:	Microscope Image		
🔽 Ch 1	Acquisition:	BP445/45	Method:	Confocal Fluorescence 405/488/640 nm
			Objective:	
		I	.ight Source:	405nm
	Target:	Microscope Image		
Ch 2	Acquisition:	BP525/50	Method:	Confocal Fluorescence 405/488/640 nm
			Objective:	10x
11		l	ight Source:	488nm
You can select only a	a single chanr	el.		OK Cance

6) Set the imaging area based on software focus.

Target Cell	Shifting Distance:	5 🤤 um
Slicing Interval Ascending Distance (+)	Ascending Distance:	5 😂 um
Shifting Distance Descending Distance (-)	Descending Distance:	-5 🛟 um
Bottom of Well Plate	Slicing Interval:	1 🛟 um

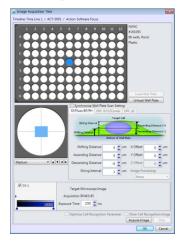
Item	Explanation		
Shifting Distance	Amount of shift from the auto-focus position		
Shifting Distance	Reference plane of software focus		
According Distance	Distance from the Shifting Distance position to the top		
Ascending Distance	plane of software focus		
Descending Distance	Distance from the Shifting Distance position to the		
Descending Distance	bottom plane of software focus		
Slicing Interval	Z step width		

In the above example, the plane 5 μ m above the auto-focus position is set as the reference plane of software focus, and images are captured in 1 μ m steps over the area between 5 μ m below and 5 μ m above this focal plane. From the total of 11 images captured, the one with the highest average brightness is output as the image in focus.

7) Display an imaging preview to check the imaging settings. Click "Test."



The Image Acquisition Test screen opens. (Refer to 6.2)



8) Click "Load Well Plate." The well plate is transferred to the reader. (Refer to 8.1 for the setting of a plate.)

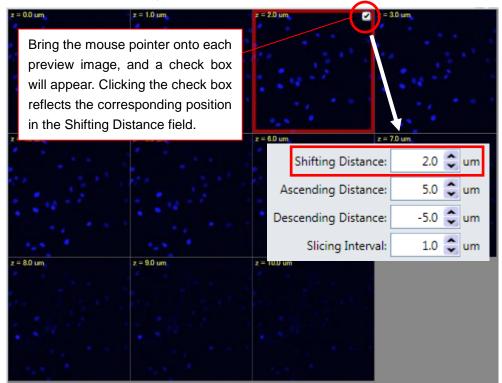


9) Click "Acquire Image" to display a preview.



10) A preview is displayed. (Refer to 5.9 and 6.2)

Bring the mouse pointer onto each preview image, and a check box will appear. Clicking the check box reflects the corresponding position in the Shifting Distance field, so click the check box of the image in best focus. The image of the highest brightness is shown with a red border, so look for a red bordered image.



11) After confirming the image selected on the preview screen, adjust the settings to appropriate values. If any of the settings has been changed, click "Acquire Image" to check the image on the preview screen again.

	01.101 D1.1		YOU I				
	Shifting Distance:	5 🤤 um	X Offset:	0 🗘	um	Set the offset set	ttings
	Ascending Distance:	5 🛟 um	Y Offset:	0 🗘	um		ungo.
┓╺┝	Descending Distance:	-5 🗘 um	Z Offset:	0 🗘	um		
	Slicing Interval:	1 🗘 um	Image Pro	cessing:			
			None		-		
			<u> </u>				
	Target: Microscope Ima	ge	C	change th	he sof	tware focus settings	S.
Ac	quisition: BP445/45	_					
Expos	ure Time: 250 🔶 ms						
	•						
					Char time.	ige the exposure	

12) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)

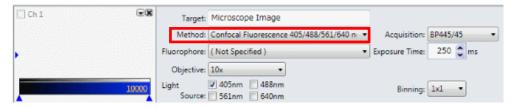


13) Click "OK."

Setting Fluorescence Acquisition

For the function of fluorescence acquisition, refer to 5.1.

1) Set the imaging channel. Under "Method," select "Confocal Fluorescence" or "Epifluorescence." (Refer to 5.5)



2) Click the Action List tab.

Time-lapse Setting	Well Plate Scan Setting	Action List

3) Click "Fluorescence Acquisition."

Software Focus Fluorescence Acquisition	3D Fluorescence Acquisition	BF / Ph Acquisition	Z-Stack BF / Ph Acquisition	DPC Acquisition	Dispense Operation
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The screen for setting fluorescence imaging opens. (Refer to 6.2)

ACT: 0001	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🗢 um
		Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 🗘 um
Acquisition	Please select the fluorescence targets.	Select Test Live Imaging: Off
		Connected Action: None

4) Click "Select."

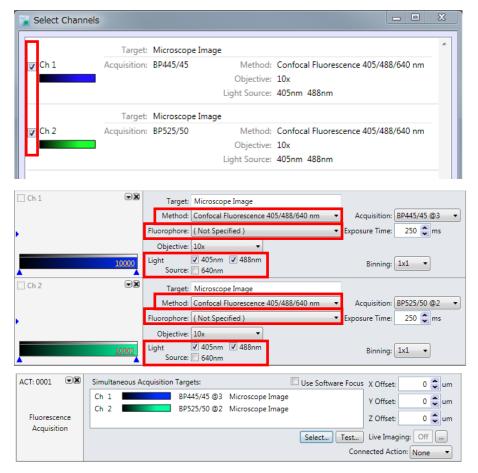
-				
ACT: 0001 📼	×	Simultaneous Acquisition Targets:	Use Software Focus	X Offset: 0 🗘 um
				Y Offset: 0 🗘 um
Fluorescence				Z Offset: 0 🗘 um
Acquisition		Please select the fluorescence targets.	Select Test	Live Imaging: Off
			Conr	nected Action: None 🔻

5) The Select Channels screen opens. Select the target channel, and then click "OK." (Multiple channels can be selected.)

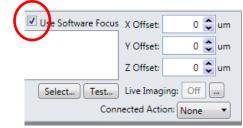
	Target:	Microscope Image		
h1	Acquisition:			Confocal Fluorescence 405/488/640 nm
			Objective:	10x
			Light Source:	405nm
	Target:	Microscope Image		
] ^C h 2	Acquisition:	BP525/50	Method:	Confocal Fluorescence 405/488/640 nm
			Objective:	10x
1			Light Source:	488nm

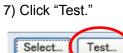
МЕМО

- If multiple channels are selected on the Select Channels screen, laser beams of multiple wavelengths are emitted simultaneously.
- •To emit laser beams of multiple wavelengths are emitted simultaneously, set the same optical system for all channels under "Method."
- If emitting laser beams of multiple wavelengths simultaneously and performing crosstalk correction by Image Correction Software, select fluorophore. Refer to 5.15 about registration of fluorophore.

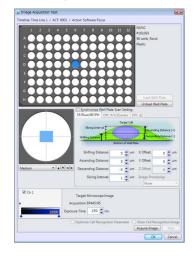


6) When "Use Software Focus" is checked, fluorescence images based on the software focused position are acquired. (Refer to 7.2)





The Image Acquisition Test screen opens. (Refer to 6.2)



8) Click "Load Well Plate." The well plate is transferred to the reader. (Refer to 8.1 for the setting of a plate.)

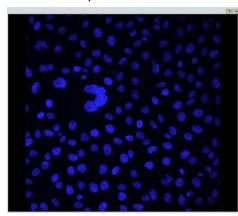


9) Click "Acquire Image" to display a preview.



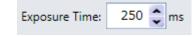
10) The preview screen opens. (Refer to 5.9 and 6.2)

Change the value of Z Offset while checking the preview screen to set the best focal plane of cells.





11) Adjust the exposure time.



- 12) After all items have been set, click "Stop" to stop the preview.
- 13) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)

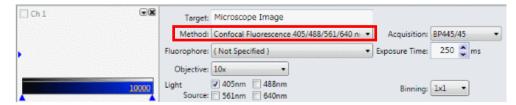
	Load Well Plate	
\mathbf{C}	Unload Well Plate	$\mathbf{)}$

14) Click "OK."

Setting 3D Fluorescence Acquisition

For the function of 3D fluorescence acquisition, refer to 5.1.

1) Set the imaging channel. Under "Method," select "Confocal Fluorescence" or "Epifluorescence." (Refer to 5.5)



2) Click the Action List tab.

Time-lapse Setting Well Plate Scan Setting Action List
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3) Click "3D Fluorescence Acquisition."

Software Focus Fluorescence Acquisition	3D Fluorescence Acquisition	BF / Ph Acquisition	Z-Stack BF / Ph Acquisition	DPC Acquisition	Dispense Operation
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The screen for setting 3D fluorescence acquisition opens. (Refer to 6.2)

ACT: 0001	Use Software Focus to Determine Shifting Distance Shifting Distance: 5 🗘 um	
	Slicing Interval	
	Shifting Distance -5 Cum	
3D	Bottom of Well Plate Slicing Interval: 1 🗘 um	
Fluorescence Acquisition	Simultaneous Acquisition Targets: X Offset: 0 🖕 um	
	Y Offset: 0 🗘 um	
	Image Processing:	
	Please select the fluorescence targets. Select Test	

 Specify the 3D imaging target. Click "Select."

Simultaneous Acquisition Targets:	
Please select the fluorescence targets.	Select Test

5) The Select Channel screen opens. Select the target channel, and then click "OK."

Select Channel					X
	Target:	Microscope Image			*
🔽 Ch 1	Acquisition:	BP445/45	Method:	Confocal Fluorescence 405/488/640 nm	
			Objective:		
			Light Source:	405nm	
	Target:	Microscope Image			
🔲 Ch 2	Acquisition:	BP525/50	Method:	Confocal Fluorescence 405/488/640 nm	
			Objective:	10x	
			Light Source:	488nm	
				\sim	
You can select only	a single chanr	nel.		ОК	ince

6) Uncheck "Use Software Focus to Determine Shifting Distance."

Use Software Focus to Determine Shifting Distance

7) Set the 3D imaging area.

Target Cell	Shifting Distance:	5 🗘 um
Slicing Interval Ascending Distance (+)	Ascending Distance:	5 😂 um
Shifting Distance Descending Distance (-)	Descending Distance:	-5 😂 um
Bottom of Well Plate	Slicing Interval:	1 😂 um

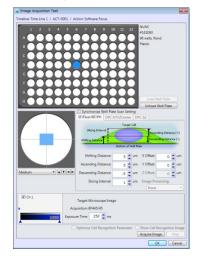
Item	Explanation
Shifting Distance	Amount of shift from the auto-focus position
	(Reference plane of 3D imaging)
Ascending Distance	Distance from the Shifting Distance position to the top
	plane of 3D imaging
Descending Distance	Distance from the Shifting Distance position to the
	bottom plane of 3D imaging
Slicing Interval	Z step width

In the above example, the plane 5 μ m above the auto-focus position is set as the reference plane of 3D imaging, and 11 images are captured in 1 μ m steps over the area between 5 μ m below and 5 μ m above this focal plane.

8) Click "Test."



The Image Acquisition Test screen opens. (Refer to 6.2)



 9) Click "Load Well Plate." The well plate is transferred to the reader. (Refer to 8.1 for the setting of a plate.)

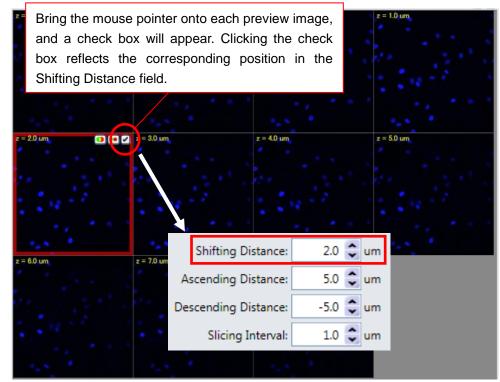


10) Click "Acquire Image" to display a preview.



11) A preview is displayed. (Refer to 5.9 and 6.2)

Bring the mouse pointer onto each preview image, and a check box will appear. Clicking the check box reflects the corresponding position in the Shifting Distance field, so click the check box of the image in best focus. The image of the highest brightness is shown with a red border, so look for a red bordered image.



12) An output method for captured Z images can be selected from the "Image Processing" items.

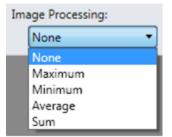
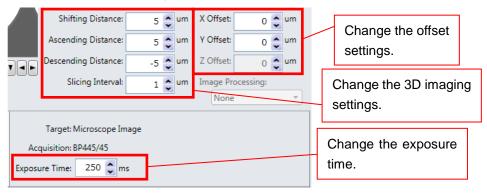


Image Processing	Explanation	
None	Acquired each Z image is saved to output folder.	
Maximum	MIP images are saved to output folder. (Refer to 5.1.)	
Minimum	MinIP images are saved to output folder. (Refer to 5.1.)	
Average	AIP images are saved to output folder. (Refer to 5.1.)	
Sum	SUM images are saved to output folder. (Refer to 5.1)	

13) Adjust the set values while checking the preview screen. If any of the settings has been changed, click "Acquire Image" to check the image on the preview screen again.



14) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)



15) Click "OK."



Selecting the "Use Software Focus to Determine Shifting Distance" check box lets you set the Shifting Distance value for the software focus plane so that 3D imaging can be performed by using the software focus plane as a reference. (Refer to 7.2)

se Software Focus to Determine Shifting Distance Shiftin	g Distance:	5 🔶 um
Target Cell Ascending Slicing Interval Ascending Distance (+)	g Distance:	5 🗘 um
	g Distance:	-5 🛟 um
Bottom of Well Plate Slici	ng Interval:	1 🛟 um
Simultaneous Acquisition Targets:	X Offset:	0 🗘 um
Ch 1 BP445/45 Microscope Image	Y Offset:	0 🛟 um
	Image P	rocessing:
Select Test	Non	e •

Setting Bright Field/Phase Contrast Acquisition

For the function of bright field/phase contrast acquisition, refer to 5.1.

1) Set the imaging channel. Under "Method," select "Brightfield" or "Phase Contrast." (Refer to 5.5)

Ch 1	Target:	Microscope Image	
	Method:	Brightfield 🔹	Acquisition: BP525/50
•			Exposure Time: 250 🔷 ms
	Objective:	10x •	
10000	Light Source:	🖉 Lamp	Binning: 1x1 •

2) Click the Action List tab.

Time-lapse Setting	Well Plate Scan Setting	Action List	

3) Click "Bright-field/Phase-contrast Acquisition."

Software Focus Fluorescence Acquisition Acquisition BF / Ph Acquisition	Z-Stack BF / Ph Acquisition	DPC Acquisition	Dispense Operation
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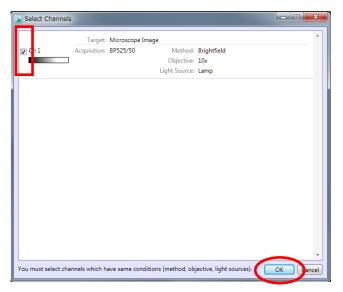
The screen for setting bright field/phase contrast imaging opens. (Refer to 6.2)

ACT: 0001	Acquisition Target:		X Offset: 0.0 🛟 um
Bright-field /			Y Offset: 0.0 🗘 um
Phase-contras Acquisition			Z Offset: 0.0 🗘 um
Acquisition	Please select the bright-field/phase-contrast target.	Select Test	Live Imaging: Off

4) Click "Select."

\sim	

5) The Select Channel screen opens. Select the target channel, and then click "OK."



6) Display an imaging preview to check the imaging settings. Click "Test."



The Image Acquisition Test screen opens. (Refer to 6.2)

				NUNC #161093 95 wells, Rom Plastic Uniced W		
	Skong Deterval	Target		Descend	ng Distar	
	Shifting Distance:	5 0	um	X Offset:	0	¢ um
	Ascending Distance	5 🗘	um	Y Offset:	0	C um
Medium • A •		-5 🗘	um	Z Offset:	0	C um
	Slicing Interval	1 🗘	um	Image Proce	ssing:	
				None		
	Target: Microscope Image Acquisition: 8P445/45 sosure Time: 250 😋 ms					
	Optimize Cell Recognition	Paramete	r (Show Cell Re Acquire Imag		on Image Stop
				OK		Cancel

7) Click "Load Well Plate." The well plate is transferred into the system. (Refer to 8.1 for the setting of a plate.)



8) Click "Acquire Image" to display a preview.



9) The preview screen opens. (Refer to 5.9 and 6.2)

Change the value of Z Offset while checking the preview screen to set the best focal plane of cells.



X Offset:	0.0 😂 um
Y Offset:	0.0 😂 um
Z Offset:	0.0 😂 um



In the phase contrast mode, correct phase-contrast images cannot be obtained except for the center of the well.

Exposure Time:	250 🗘	ms
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- 11) After all items have been set, click "Stop" to stop the preview.
- 12) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)

Load Well Plate
Unload Well Plate

13) Click "OK."

Setting Z-stack Bright-field/Phase-contrast Acquisition

For the function of Z-stack bright-field/phase-contrast acquisition, refer to 5.1.

1) Set the imaging channel. Under "Method," select "Brightfield" or "Phase Contrast." (Refer to 5.5)

Ch 1	Target:	Microscope Image	_	
	Method:	Brightfield 🔹	Acquisition:	BP525/50 -
•			Exposure Time:	250 😂 ms
	Objective:	10x •		
10000	Light Source:	🖉 Lamp	Binning:	1x1 •

2) Click the Action List tab.

[Time-lapse Setting	Well Plate Scan Setting	Action List	
Γ				

3) Click "Z-Stack Bright-field/Phase-contrast Acquisition."

Software Focus Fluorescence Acquisition	3D Fluorescence Acquisition	BF / Ph Acquisition	Z-Stack BF / Ph Acquisition	DPC Acquisition	Dispense Operation
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The screen for setting Z-stack bright-field/phase-contrast imaging opens. (Refer to 6.2)

ACT: 0001	Target Cell Shiftir	ng Distance:	5 🤤 um
	Slicing Interval Ascending Distance (+) Ascendir	ng Distance:	5 춫 um
		ng Distance:	-5 🛟 um
Z-Stack	Bottom of Well Plate Slic	ing Interval:	1 🤤 um
Bright-field /		_	
Phase-Contrast Acquisition	Acquisition Target:	X Offset:	0 🤤 um
Acquisition		Y Offset:	0 😂 um
		Image Pr	ocessing:
	Please select the bright-field/phase-contrast target. Select Test	None	•

4) Click "Select."

Simultaneous Acquisition Targets:	
Please select the bright-field/phase-contrast target.	Select Test

5) The Select Channel screen opens. Select the target channel, and then click "OK."

	Target:	Microscope Im	age		
🛛 Ch 1	Acquisition:	BP525/50	Method:	Brightfield	
			Objective:		
			Light Source:	Lamp	

6) Set the Z-stack imaging area.

Slicing Interval Ascending Distance (+) Ascending Distance: 5 🙄 um Shifting Distance Descending Distance (-) Descending Distance: -5 🙄 um Bottom of Well Plate Slicing Interval: 1 🙄 um	Target Cell	Shifting Distance: 5 📚 um
	Slicing Interval	Ascending Distance: 5 🗘 um
Bottom of Well Plate Slicing Interval: 1 🗘 um	Shifting Distance	•) Descending Distance: -5 🔷 um
	Bottom of Well Plate	Slicing Interval: 1 🗘 um

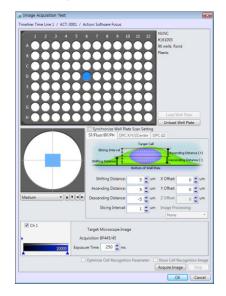
Item	Explanation		
Chifting Distance	Amount of shift from the auto-focus position		
Shifting Distance	(Reference plane of Z-stack imaging)		
According Distance	Distance from the Shifting Distance position to the top		
Ascending Distance	plane of Z-stack imaging		
Descending Distance	Distance from the Shifting Distance position to the		
Descending Distance	bottom plane of Z-stack imaging		
Slicing Interval	Z step width		

In the above example, the plane 5 μ m above the auto-focus position is set as the reference plane of Z-stack imaging, and 11 images are captured in 1 μ m steps over the area between 5 μ m below and 5 μ m above this focal plane.

7) Display an imaging preview to check the imaging settings. Click "Test."



The Image Acquisition Test screen opens. (Refer to 6.2)



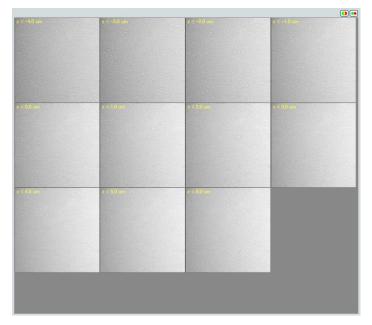
8) Click "Load Well Plate." The well plate is transferred into the system. (Refer to 8.1 for the setting of a plate.)



9) Click "Acquire Image" to display a preview.



10) The preview screen opens. (Refer to 5.9 and 6.2.)





In the phase contrast mode, correct phase-contrast images cannot be obtained except for the center of the well. 11) An output method for captured Z images can be selected from the "Image Processing" items.

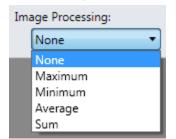
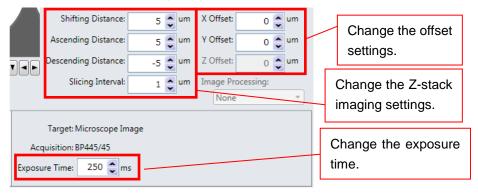


Image Processing	Explanation
None	Acquired each Z image is saved to output folder.
Maximum	MIP images are saved to output folder. (Refer to 5.1.)
Minimum	MinIP images are saved to output folder. (Refer to 5.1.)
Average	AIP images are saved to output folder. (Refer to 5.1.)
Sum	SUM images are saved to output folder. (Refer to 5.1)

12) Adjust the set values while checking the preview screen. If any of the settings has been changed, click "Acquire Image" to check the image on the preview screen again.



13) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)

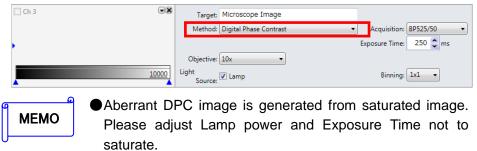


14) Click "OK."

Setting DPC Acquisition

Refer to 5.1 for the DPC acquisition function.

1) Set the imaging channel. Under "Method," select "Digital Phase Contrast." (Refer to 5.5)



2) Click the Action List tab.

	Time-lapse Setting	Well Plate Scan Setting	Action List
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3) Click "DPC Acquisition".

Eocus Acquisition	3D Iuorescence Acquisition	Z-Stack BF / Ph Acquisition	DPC Acquisition	Dispense Operation
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The screen for setting DPC Acquisition opens. (Refer to 6.2)

ACT: 0001 💌 🗷	Search Center of Z	Shifting Di Ascending Di Descending Di	istance:	10 🗘	um
DPC Acquisition	Bottom of Well Plate Acquisition Target:	Slicing I	X Offset: Y Offset:	2 0	um um um
	Please select a DPC target. Select	Test Im	ΔΖ: nage Type: Contrast:		• um

4) Click "Select".

Acquisition Target:	
Please select a DPC target.	Select Test

5) The Select Channel screen opens. Select the target channel, and then click "OK."

	Target:	Microscope Im	nage		
1	Acquisition:	BP525/50	Method:	Digital Phase Contrast	
			Objective:	10x	
			Light Source:	Lamp	

6) Set the automatic DPC reference position search.

Search Center of Z	Shifting Distance:	5 🛟 um
Ascending Distance (+) Slicing Interval	Ascending Distance:	10 🛟 um
Shifting Distance	Descending Distance:	-10 🛟 um
Bottom of Well Plate	Slicing Interval:	2 🧲 um

Item	Explanation			
Search Center of Z	Check in case of performing automatic DPC			
Search Center of Z	reference position search.			
	Amount of shift from the auto-focus position			
	(If "Search Center of Z" is checked, this Z position is			
Shifting Distance	reference plane of automatic DPC reference position			
	search. If unchecked, this Z position is reference			
	plane of DPC imaging)			
According Distance	Distance from the Shifting Distance position to the top			
Ascending Distance	plane of automatic DPC reference position search			
	Distance from the Shifting Distance position to the			
Descending Distance	bottom plane of automatic DPC reference position			
	search			
Slicing Interval	Z step width			



DPC image is created from multiple images of different Z position by image processing. DPC reference position means the reference Z position of these multiple images.

In the above example, the plane 5 μ m above the auto-focus position is set as the reference plane of automatic DPC reference position search, and bright field images are captured in 2 μ m steps over the area between 10 μ m below and 10 μ m above this focal plane. From the total of 11 images captured, the one with the most optimal for center of DPC acquisition is output as the image in focus.

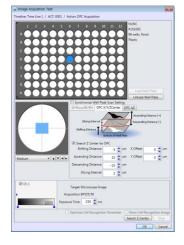
7) Select DPC image type to output.

Image Type:	Fluor type 🔹
Contrast:	Fluor type
Contrast:	Phase type

8) Display an imaging preview to check the imaging settings. Click "Test."



The Image Acquisition Test screen opens. (Refer to 6.2)



9) Click "Load Well Plate." The well plate is transferred to the reader. (Refer to 8.1 for the setting of a plate.)

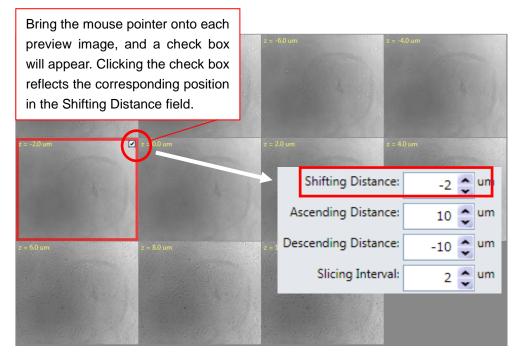


10) Click "Acquire Image" to display a preview.



11) A preview is displayed. (Refer to 5.9 and 6.2)

Bring the mouse pointer onto each preview image, and a check box will appear. Clicking the check box reflects the corresponding position in the Shifting Distance field, so click the check box of the image in best focus. The image of the most optimal for center of DPC acquisition is shown with a red border, so look for a red bordered image.



12) After confirming the image selected on the preview screen, adjust the settings to appropriate values. If any of the settings has been changed, click "Acquire Image" to check the image on the preview screen again.

	Search Z Center for	r DPC				
	Shifting Distance:	5 🍣 ur	X Offset:	0 춫 um	Set the offset se	ttings.
┓┫┣	Ascending Distance:	10 춫 ur	Y Offset:	0 🤤 um		
	Descending Distance:	-10 춫 ur				
	Slicing Interval:	2 춫 ur				
-						
Aco	Target: Microscope Ima uisition: BP525/50	ge	Cł	nange the sol	ftware focus settings	6.
	re Time: 250 🍨 ms					
				Char	nge the exposure	
				time.		

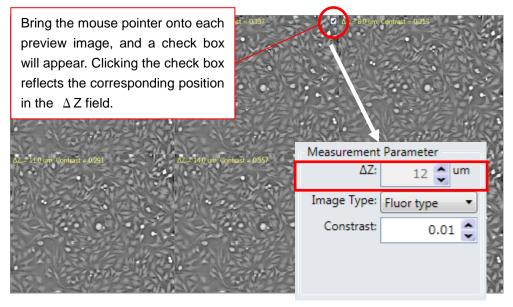
13) In case of changing DPC acquiring setting, select " ΔZ " tab. Check "Auto Contrast" and "Acquire Multiple ΔZ " and click "Acquire Image".

SF/Fluor/BF/PH DPC X/Y/ZCenter DPC ΔZ				
Z Center Measurement Parameter $\Delta Z: 50 \bigcirc um$ Image Type: Fluor type Constrast: 0.01 \bigcirc Refresh Preview	Test Davameter Auto Contrast Auto Contrast Acquire Multiple ΔΖ Maximum ΔΖ: 15 ↓ um Minimum ΔΖ: 3 ↓ um Step Length: 3 ↓ um			
Target: Microscope Image .cquisition: BP525/50 sure Time: 250 ms				
Optimize Cell Recognition Parameter Show Cell Recognition Image n test finished Acquire Image Stop				



●DPC image is created from multiple bright field images of different Z position by image processing. " △ Z" in this procedure means distance of these Z positions. 14) A preview is displayed. (Refer to 5.9 and 6.2)

Bring the mouse pointer onto each preview image, and a check box will appear. Clicking the check box reflects the corresponding position in the Δ Z field, so click the check box of the image in the most optimal DPC image.



15) To remove the well plate, click "Unload Well Plate." Remove the well plate from the stage. (Refer to 8.2)



16) Click "OK."

Setting Dispensing Operation

For the function of dispensing operation, refer to 5.1.

1) Click the Action List tab.



2) Click "Dispense Operation."

Software Focus Fluorescence Acquisition 3D Fluorescence Acquisition BF / Ph Acquisition Z-Stack BF / Ph Acquisition DPC Acquisition	Dispense Operation
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The screen for setting dispensing opens. (Refer to 6.2)

ACT: 0001	•×	Dispense Setting Name:	
Dispense Operation		1	Edit Dispense Setting Please enter dispense setting name.

3) Select the dispensing setting file.

Dispense Setting Name:	s		
Ed	it Dispense Setting		
Please enter disp	ense s Click here to open the screen for		
	selecting a dispensing setting file.		

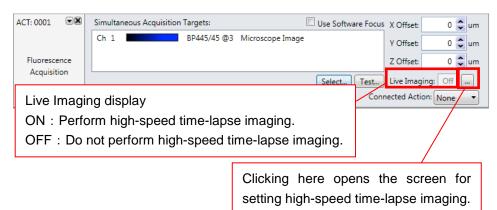
After selecting the dispensing setting file, click "Select."

Select a	dispense se	etting :					
Dispense	Setting						
🗾 Disp	ensing test						
Create [Dispense Setting	Modify Dispense Setting		Select		Cancel	
Create	a new disper	sing setting file.	E	dit a dispe	ensing	setting	, file.
(Refer	to 5.12)						
			,				
Dispense	Setting Name:	Dispensing test					
		Edit	Dispens	e Setting			
	Edit the dis	pensing setting fi	le to h	ave been :	selec	ted.	

5.8. Setting High-speed Time-lapse Imaging

For the function of high-speed time-lapse imaging, refer to 5.1.

- 1) On the Action List tab, set a "Fluorescence Acquisition" task. (Refer to 5.7)
- 2) Open the screen for setting high-speed time-lapse imaging.



The screen for setting high-speed time-lapse imaging opens. (Refer to 6.2)

🚡 S	et Live Imaging Pi	arameters		
10	No Live Imaging			
ົ ເ	Jse Live Imaging:			
	Period:	10000 🗘 ms		
	Interval:	261 🗘 ms		
		Dispense Setting Name	Timing	Add
	Dispense Setting:			Change
	Dispense setting.			Delete
				Liquid Volume
		Perform AF during live imaging		
	Test Interval Value			OK Cancel

3) Select "Use Live Imaging."

No Live Imaging

Use Live Imaging:

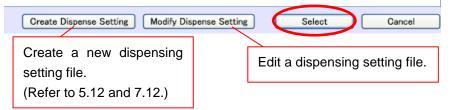
 4) Enter the value of "Period" (period of high-speed time-lapse imaging). (Interval≦Period)

Period:	30000	0	ms
1 6110 61	20000	~	

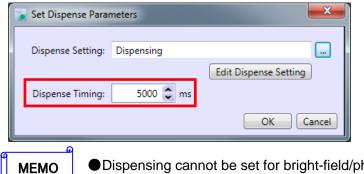
5) Set dispensing setting. (Set this item if dispensing is performed.) Click "Add."

Dispense Setting:	Dispense Setting Name	Timing	Add
			Change
			Delete
			Liquid Volume
🚡 Set Dispense F	Parameters		
Dispense Setti	ng: Dispensing		
Dispense Timi	ng: 5000 🗘 ms	Edit Dispense Setting	
		OK Cancel	
Click to	display the screer	n to select dispensing	setting file.

Select a dispensing setting file and then click "Select."



Enter the value of "Dispense Timing" (timing at which to drip reagent). (1000ms≦Dispense Timing)



Dispensing cannot be set for bright-field/phase-contrast acquisition.

 Up to three dispensing setting files can be assigned. (Refer to 5.1 for detail of multi dispensing.) 6) Enter the value of "Interval" (interval of high-speed time-lapse imaging). After entering, click "Test Interval Value."

ء 📷	Set Live Imaging Pa	rameters	X				
0	O No Live Imaging						
۰	Use Live Imaging:						
	Period:	10000 🗘 ms					
	interval:	261 🗘 ms					
		Dispense Setting Name Timing	Add				
	Dispense Setting:		Change				
	bispense settingi		Delete				
			Liquid Volume				
Ι.	Perform AF during live imaging						
	Test Interval Value)	OK Cancel				

A message screen is displayed if the entered value for "Interval" is shorter than allowed. Re-enter "Interval" according to the message.

	Period:	30000 🗘 ms			
	Interval:	182 🗘 ms			
	CellVoyager Mea	isurement System		×	
	The interval value	ue must be 315 msec or gre	ater.		
	Re-ent	er "Interval" lo	nger than 315ms.		
			ОК		
ľ	MEMO	paramete	value for "Interva rs such as exposure storage, autofocus	e time, binning, tra	

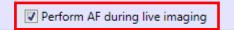
7) Check "Perform AF during live imaging" to perform autofocus during high-speed time-lapse imaging.

ء 😿	Set Live Imaging Pa	arameters		X		
0	No Live Imaging					
0	Use Live Imaging:		ms g Name Timing Add Change Delete Liquid Volume			
	Period:	10000 🗘 ms				
	Interval:	261 🗘 ms				
		Dispense Setting Name	Timing	Add		
	Dispense Setting:			Change		
	bispense setting.			Delete		
				Liquid Volume		
		Perform AF during live imagin	g 🖌			
	Test Interval Value	Interval: 261 ms Dispense Setting Name Timing Add Setting: Delete Liquid Volume Perform AF during live imaging				

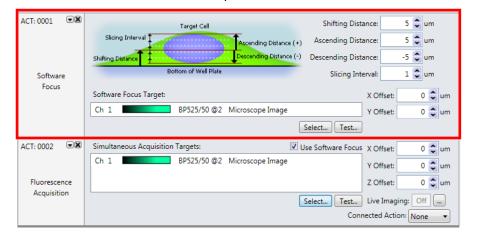
MEMO

Autofocus during high-speed time-lapse imaging cannot be performed if "Interval" isn't long enough. (According to acquisition condition, about over than 5000 ms.)

In this case, red box appears around "Perform AF during live imaging". Uncheck "Perform AF during live imaging" or set longer value to "Interval".



• Following setting is recommended when autofocus during high-speed time-lapse imaging is performed. Also, following setting must be set in combination. - Set software focus at previous "Action List".



- Check "Force undersurface detection on AF" at bottom of "Action List".



Optimize measurement sequence to shorten time for Force undersurface detection on AF

🚡 S	et Live Imaging Pa	arameters		X		
0	No Live Imaging					
Use Live Imaging:						
	Period: 30000 🗘 ms					
	Interval:	400 🗘 ms				
		Dispense Setting Name	Timing	Add		
		Dispensing	5000 ms	Change		
	Dispense Setting:			Delete		
	Liquid Volume			Liquid Volume		
Perform AF during live imaging						
	Test Interval Value			OK Cancel		

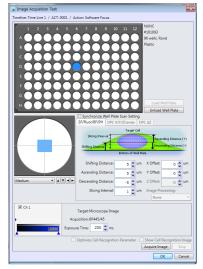
Example of a high-time lapse setting

In the above example, time-lapse imaging is performed at an imaging interval of 400 ms and reagent is dropped 5 seconds after the start of imaging. The system moves to the next well after capturing for a period of 30 seconds.

8) After all items have been set, click "OK."

ОК	Cancel	
ACT: 0001	Simultaneous Acquisition Targets: 🔲 Use Software Focus	X Offset: 0 🗘 um
	Ch 1 BP445/45 @3 Microscope Image	Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 🗘 um
Acquisition	Select) Test	Live Imaging: On 🛄
	The Live Imaging setting changes to "ON."	ected Action: None 🔹

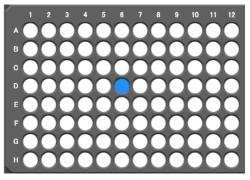
5.9. Setting the Preview Screen



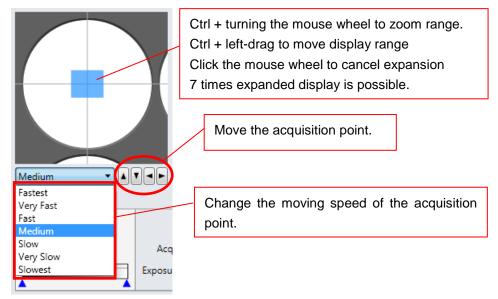
Preview screen (Refer to 6.2)

1) Select the well whose preview will be displayed.

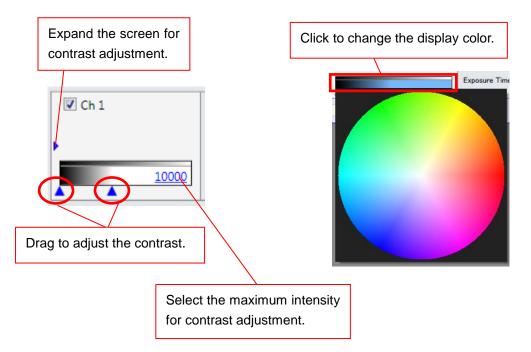
Click a desired well to be used for the image-acquisition test.



2) Specify the acquisition point in the well. Click the arrow buttons to move the acquisition point. Click on the well, and the clicked point will be specified as the imaging point.



3) Adjust the contrast on the preview screen. Click the contrast bar to change the display color for preview.



5.10. Setting the Cell Recognition Algorithm

🚡 Set Cell R	ecognition Parameters
Algorithm:	Connection
Threshold:	1000 🗘
	Remove the fragments of the cell at the edge of the image
Measure:	Mean Cell Diameter: 15 📚 um
	Minimum Scaling Factor: 0.5 💙 (7.5 um)
	Maximum Scaling Factor: 2 💙 (30.0 um)
	OK Cancel

Cell recognition algorithm setting screen

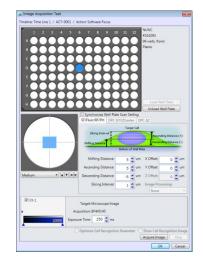
Item		Explanation
•		Sets the algorithm to count the number of
		cells.
	Binary per Mean Area	Algorithm to obtain the number of cells by
		binarizing cell images and dividing the total
		area of the recognized cells by the mean
		cell size.
	Connection	Algorithm to binarize cell images, perform
		cell labeling, and count the number of cells
		that fall within the range between the
		entered minimum and maximum scaling
		factors relative to the cell mean size.
	Connection and Binary per	Algorithm to add the number of cells
	Mean Area	obtained by the Connection algorithm and
		the number of cells obtained by the Binary
		per Mean Area algorithm in relation to the
		number of cells that exceeds the maximum
		scaling factor relative to the cell mean
		size.
Th	reshold	Threshold to binarize cell images.
Re	move the fragments of the	Check the checkbox to exclude the cells at
cell at the edge of the image		the edges of the image in the cell count.
Mean Cell Diameter		Specifies the mean cell diameter.
		Unit:µm
Mir	nimum Scaling Factor	Sets the minimum scaling factor.
Maximum Scaling Factor		Sets the maximum scaling factor.

Items displayed on the cell recognition algorithm setting scre	en
--	----

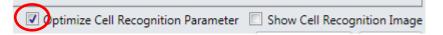
1) Select the cell recognition algorithm.

Algorithm:	Connection	~
, "Bourerun		· ·

- 2) Select the following check box if you do not want the cells at edges of the image to be recognized.
- Remove the fragments of the cell at the edge of the image
- 3) Do not set anything under "Threshold" and "Measure," and click "OK."
- 4) The preview screen opens based on the Action List tab settings to which the cell recognition algorithm has been applied. Adjust the well position, imaging position, etc. (Refer to 5.9)



5) Select the "Optimize Cell Recognition Parameter" (automatically optimize the set values of the recognition algorithm) checkbox, and then click "Acquire Image."



6) After checking the preview screen, click "Stop." When the preview screen closes, the "Threshold" and "Measure" parameters on the cell recognition algorithm setting screen have been automatically set to optimal values. To manually set the parameters, unselect the "Optimize Cell Recognition Parameter" check box and adjust the "Threshold" and "Measure" parameters while checking the preview screen.

🝺 Set Cell Re	ecognition Parameters
Algorithm:	Connection
Threshold:	687 🗘
	Remove the fragments of the cell at the edge of the image
Measure:	Mean Cell Diameter: 12 🔹 um
	Minimum Scaling Factor: 0.5 📚 (6.0 um)
	Maximum Scaling Factor: 2 2 (24.0 um)
	OK Cancel

5.11. Saving the Measurement Setting File

1) In the menu of the measurement setting file, click "File" and then select "Save As."

🝺 new setting* [Measurement Setting] - Cel				
File	Edit View	Help		
	Save	Ctrl+S		
	Save As	Ctrl+Shift+S		
	Close		<u>96, 9(</u>	

 Enter the name under which to save the measurement setting file, and click "Save the Measurement Setting."

Measurement Setting List - CellVoyager				
nter a measurement setting na	me:			
Measurement Setting	Well Plate Type	Application D	Date	
	Measurement Setting Name:	Save the	e Measurement Setting Cancel	
asurement Setting Name	Sample Test	Sa	ve the Measurement Settin	g
0	L			

5.12. Creating a Dispensing Setting File

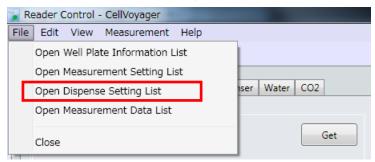
1) Click "More" in the Reader area to open the Reader Control screen.

Reader	
Status:	
Ready	
Progression of Time-Lapse:	
Expected Time: Elapsed Time: Remaining Time:	
Progression of Current Time Point:	
Expected Time:	
Elapsed Time: Remaining Time:	
	Start Measurement

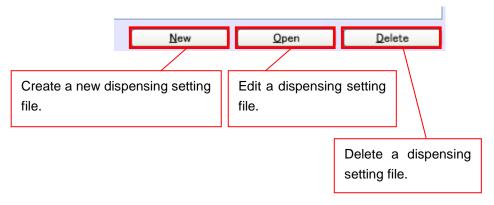
The Reader Control screen opens. (Refer to 6.4)

Argument of Tree Lague	
Image: standard sta	
Image: Contraction	
Prode	
Ar Nach (1) Ar Nach (2) Ar Nach (
Number Store In Transito Ore In Transito Ore In Transito Ore	
Trins the form The set of the set o	
Are to forme A	
Maxement Charles II I I I I I I I I I I I I I I I I I	
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apende Time	atten 0∑ \$ Ston 0∑ \$ 60m 0∑ \$ 00 0∑ \$ Long 0∑
period Tome	
naining Time gywsion of Current Time Pont:	
ogression of Current Time Point:	
sected Times second	
pad Time:	

2) Click "Open Dispense Setting List" in the File menu.



3) The Dispense Setting List screen opens. Click "New."



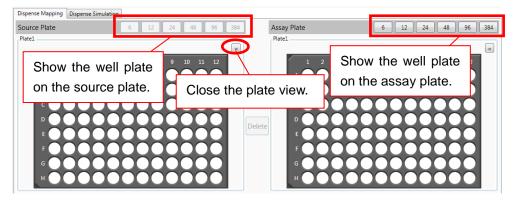
The Dispense Setting screen appears. (Refer to 6.3)

new file [Dispense Setting] - CellVoyager			(in) (i) ×
File View Simulation			
Dispense Setting	Dispense Mapping Dispense Si		
Sourantikat Assynthet Sen	^r Source Plate	Assay Plate	6 12 24 46 96 36t
Basic Disperse Setting			
😗 🗔 Prevet			
🔁 🗌 AspirateStir			
🚺 🗔 Airgap			
🔂 🛄 Aspirate			
🖪 🗌 AspirateTiptouch			
1 Dispense			
8 🖸 DispenseStr			
🔁 🛄 DispenseTiptouch			
📴 🗌 ArBow			

4) In the menu, click View -> Basic Dispense Setting or Advanced Dispense Setting, and select the dispensing setting mode. To configure the advanced setting for the dispensing operation, select Advanced Dispense Setting.

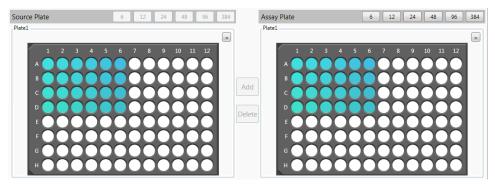
<u>F</u> ile	⊻iew	Simulation	
Dis	Dispense Mapping		
Se	Dispense Simulation		
Plat	✓ Basic Dispense Setting		
Plat	Advanced Dispense Setting		
D1 -	4 / 4 /		

5) Display the Dispense Mapping tab and then display source plate and assay plate.





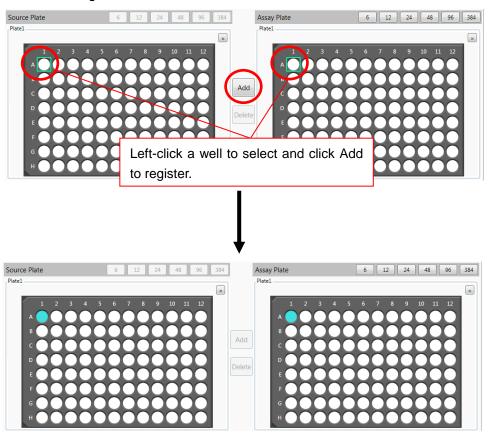
- Only one source plate and up to four assay plates can be set.
- Dispensing to multiple assay plates is possible when a large incubator (sold separately) is used.
- 6) Link the wells on the source plate and assay plate.



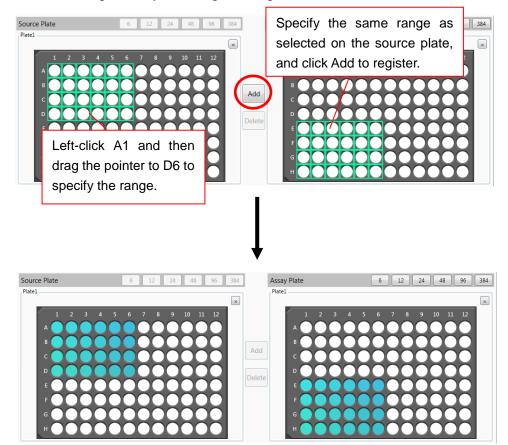


Dispensing to an assay plate from one well on the source plate is possible up to four wells.

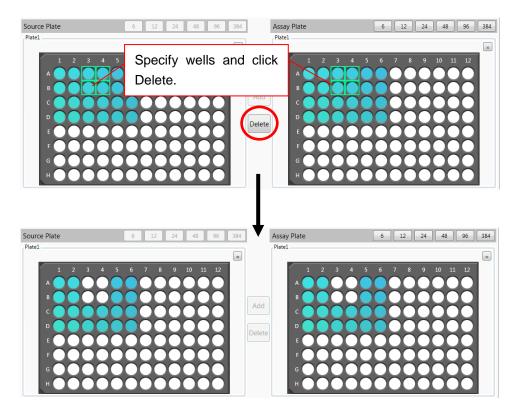
•When linking individual wells



•When linking wells by selecting the range



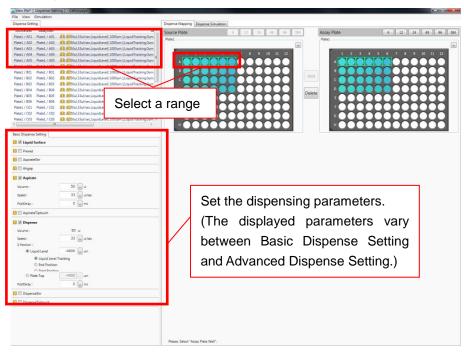
When deleting wells



7) Set the dispensing parameters. (Refer to 5.13)

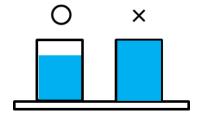
When the association is set, the setting items for individual wells are displayed on the "Dispense Setting" tab.

Select a range of wells in the source plate, or select a range of wells on the "Dispense Setting" tab by dragging with the mouse while holding down the Shift key, and then enter the dispensing parameters. The same parameters can be set to all wells in the selected range. The dispensing parameters to be entered vary between "Basic Dispense Setting" and "Advanced Dispense Setting."



✓!\ CAUTION

• Do NOT put solution up to each well top of Assay plate and Source plate.





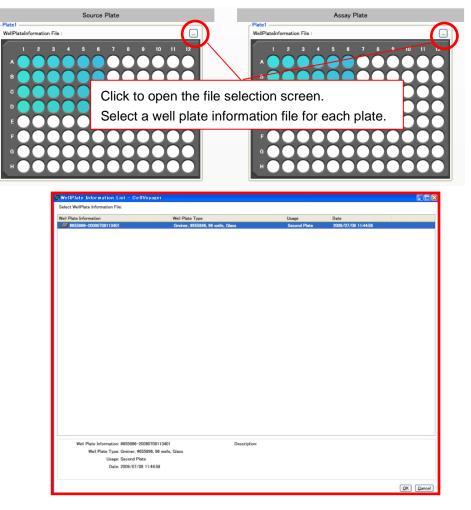
●Indication of reagent volume is 20% - 40% of well volume in Assay Plate and 50% - 80% of well volume in Source Plate.

8) Perform dispensing simulation.

Click the "Dispense Simulation" tab.

Dispense Mapping	Dispense Simulation
	Source Plate
Plate1	
WellPlateInform	ation File :

9) Select the well plate information file. (Refer to 4.1 for the creation of a well plate information file.)



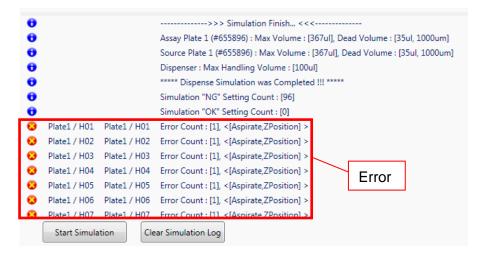
10) Click Start Simulation.



If the simulation is successful, the following message appears.

0			>>> Simulation Finish <<<
0			Assay Plate 1 (#655896) : Max Volume : [367ul], Dead Volume : [35ul, 1000um]
0			Source Plate 1 (#655896) : Max Volume : [367ul], Dead Volume : [35ul, 1000um]
0			Dispenser : Max Handling Volume : [100ul]
0			***** Dispense Simulation was Completed !!! *****
θ			Simulation "NG" Setting Count : [0]
0			Simulation "OK" Setting Count : [96]
0	Plate1 / H01	Plate1 / H01	Error Count : [0]
θ	Plate1 / H02	Plate1 / H02	Error Count : [0]
0	Plate1 / H03	Plate1 / H03	Error Count : [0]
θ	Plate1 / H04	Plate1 / H04	Error Count : [0]
0	Plate1 / H05	Plate1 / H05	Error Count : [0]
0	Plate1 / H06	Plate1 / H06	Error Count : [0]
0	Plate1 / H07	Plate1 / H07	Error Count : 101
	Start Simula	tion	ear Simulation Log

If an error occurs with the simulation, an error message appears. Change the error setting, and continue the simulation until an error message does not appear.



To confirm error in detail, double-click each of error items.

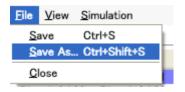
SimulationLog - Cell Voyager	
Source Plate : Plate1 / H01 Assay Plate : Plate1 / H01	
SimulationErrorLog Detail	
 Aspirate [Volume] (The Least Reagent Volume[20 ul]) < (Dead Volume[35 ul]) The Least reagent Volume on this liquid handling is less than dead volume. Please change either "Aspirate Volume" or "Reagent Volume". Setting Data : Reagent Volume(Start of Aspirate)[100 ul], Aspirate Volume[80 ul], Unavailable Volume[35 ul] 	
	ОК

•Example of a simulation error

SimulationLog - Cell Voyager	
Source Plate : Plate1 / H01 Assay Plate : Plate1 / H01	
SimulationErrorLog Detail	
* Aspirate [Volume] (The Least Reagent Volume[20 ul]) < (Dead Volume[35 ul]) The least reagent volume on this liquid handling is less than dead volume. Please change either "Aspirate Volume" or "Reagent Volume". Setting Data : Reagent Volume(Start of Aspirate)[100 ul], Aspirate Volume[80 ul], Unavailable Volume[35 ul]	
This error indicates that liquid amount of Source plate after aspirating is less than Dead Volume. You need to make initial amount for Source plate larger or to make aspirating amount lower.	ОК

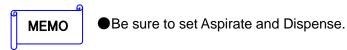
11) Save the dispensing setting file.

In the menu, click File -> Save As.



5.13. Setting the Dispensing Parameters

Basic Dispense Setting



Liquid Surface

This detects the liquid surface. It lowers the tip end in the well on the source plate so far that it comes in contact with the liquid surface to detect the liquid surface position in the well.

To set the Liquid Surface function, select the check box.

📙 🗹 Liquid Surface

Prewet

This aspirates and drops the solution in the well on the source plate to make the inside of the tip wet with the solution.

To set the Prewet function, select the check box. Enter the value of "Volume" (amount of reagent suctioned).

(96-tip rack model: 0≦Volume≦100) (384-tip rack model: 0≦Volume≦20)



AspirateStir

This stirs the solution in the well on the source plate. The solution is aspirated and discharged the number of times specified.

To set the AspirateStir function, select the check box. Enter the values of "Execution" (number of times solution is stirred) and "Volume" (amount of reagent suctioned). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

```
(1 \leq \text{Execution} \leq 5) (0 \leq \text{PostDelay} \leq 10000)
```

(96-tip rack model: $0 \leq \text{Volume} \leq 100$) (384-tip rack model: $0 \leq \text{Volume} \leq 20$)



Airgap

This delivers air on top of the solution inside the tip. It drops the solution to the last drop.

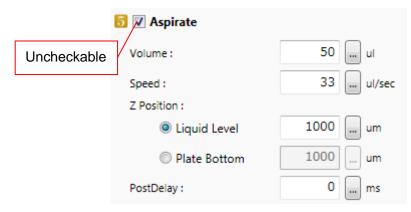
To set the Airgap function, select the check box. Enter the value of "Volume" (air gap).

(96-tip rack model: $0 \leq \text{Volume} \leq 100$) (384-tip rack model: $0 \leq \text{Volume} \leq 20$)



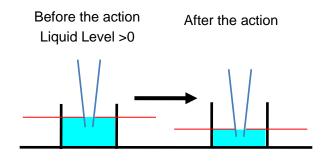
Aspirate

This aspirates the solution from the well on the source plate. Enter the values of "Volume" (amount of reagent filled), "Speed" (filling speed) and "Z Position" (filling position). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled. (96-tip rack model: $0 \le$ Volume ≤ 100) (384-tip rack model: $0 \le$ Volume ≤ 20) (96-tip rack model: $0 \le$ Speed ≤ 34) (384-tip rack model: $0 \le$ Speed ≤ 7) ($0 \le$ PostDelay ≤ 10000) (-50000 \le LiquidLevel, PlateBottom ≤ 100000)

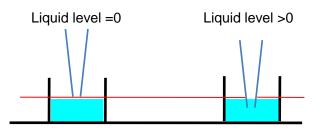


Z Position	Explanation
	Track the drop in liquid level due to filling.
	Input value indicates the distance between the tip
Liquid Level	and liquid level.
	(+ : Below the liquid level, 0 : Around the liquid level,
	- : Above the liquid level)
	Fill the solution by using the bottom face of the plate
Plate Bottom	as a reference.
	(+: Below the bottom face of the plate, 0: Around the
	bottom face of the plate, - : Above the bottom face of
	the plate)

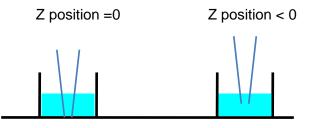
Liquid Level tracks the fall of the liquid level that occurs as the solution is filled and lowers the tip position in accordance with the fall of the liquid level. The relative position of the liquid level and tip is fixed.



Liquid Level indicates the distance from the tip to the liquid surface. The plus indicates the downward tip position and the minus indicates the upward tip position in relation to the liquid level.



With regards to Plate Bottom, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the bottom plane of the well. Enter minus to set.



AspirateTiptouch

This lets the tip come in contact with the well wall on the source plate and drops the droplet adhered to the end.

To set the AspirateTiptouch function, select the check box.

🚹 🗸 AspirateTiptouch

Dispense

This enables you to set how to drop the solution.

Enter the values of "Speed" (dripping speed) and "Z Position" (dripping position). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled. The value of "Volume" (amount of solution dripped into each well) is calculated automatically based on the set parameters.

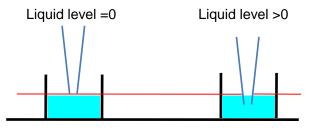
(96-tip rack model: $0 \leq \text{Speed} \leq 34$) (384-tip rack model: $0 \leq \text{Speed} \leq 7$) (-50000 $\leq \text{LiquidLevel}$, PlateTop, PlateBottom ≤ 100000)

(0≦PostDelay≦10000)

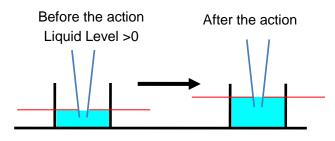
7 闭 Dispense	Unchec	kable		
Volume :		50	ul	
Speed :		33		ul/sec
Z Position :			_	
Liquid	Level	-1000		um
۲	Liquid Level Ti	racking		
End Position				
\odot	Start Position			
O Plate	Тор	-1000		um
Plate	Bottom	-1000		um
PostDelay :		0		ms

Z Position	Explanation
	Distance between the tip and liquid level
Liquid Level	(+ : Below the liquid level, 0 : Around the liquid level,
	- : Above the liquid level)
Liquid Level Tracking	Track the rise in liquid level due to dripping.
End Position	Drip the solution by using the liquid level after dripping as
	a reference.
Start Position	Drip the solution by using the liquid level before dripping
	as a reference.
	Drop by using the top face of the plate as a reference.
Plate Top	(+ : Below the top face of the plate, 0 : Around the top face
	of the plate, - : Above the top face of the plate)
	Drop by using the bottom face of the plate as a refer-
	ence.
Plate Bottom	(+ : Below the bottom face of the plate, 0 : Around the
	bottom face of the plate, - : Above the bottom face of the
	plate)

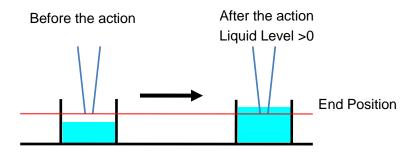
Liquid Level indicates the distance from the tip to the liquid level. The plus indicates the downward tip position and the minus indicates the upward tip position in relation to the liquid level.



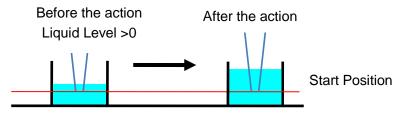
Liquid Level Tracking tracks the rise of the liquid level that occurs as the solution is dropped and raises the tip position in accordance with the rise of the liquid level. The relative position of the liquid level and tip is fixed.



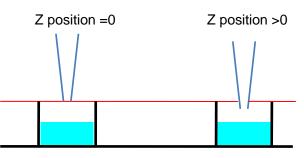
End Position sets the tip to a position that is shifted from the liquid level after the action by the liquid level you enter. The tip position before and after the action is unchanged.



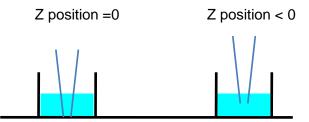
Start Position sets the tip to a position that is shifted from the liquid level before the action by the liquid level you enter. The tip position before and after the action is unchanged.



•With regards to Plate Top, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the top plane of the well.



With regards to Plate Bottom, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the bottom plane of the well. Enter minus to set.



DispenseStir

This stirs the solution in the well after the solution is dropped. The solution in the well is aspirated and discharged the specified number of times.

To set the DispenseStir function, select the check box. Enter the values of "Execution" (number of times solution is stirred) and "Volume" (amount of reagent suctioned). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

 $(1 \leq \text{Execution} \leq 5) (0 \leq \text{PostDelay} \leq 10000)$

(96-tip rack model: $0 \leq \text{Volume} \leq 100$) (384-tip rack model: $0 \leq \text{Volume} \leq 20$)

8 🗸 DispenseStir	
Execution :	1
Volume :	50 ul
PostDelay :	0 ms

DispenseTiptouch

This lets the tip come in contact with the well wall on the assay plate and drops the droplet adhered to the end.

To set the DispenseTiptouch function, select the check box.

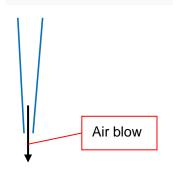
<u> v</u> DispenseTiptouch

AirBlow

This allows to blow air from inside tip to drop droplet adherent on the tip top. To set the AirBlow function, select the check box.

This function can be used only when liquid amount (including Airgap) inside tip is 0ul.

10 🔽 AirBlow



Advanced Dispense Setting



•Be sure to set Aspirate and Dispense.

Liquid Surface

This detects the liquid surface. It lowers the tip end in the well on the source plate so far that it comes in contact with the liquid surface and detects the liquid surface position in the well.

To set the Liquid Surface function, select the check box.

📙 🗸 Liquid Surface

Prewet

This aspirates and drops the solution in the well on the source plate to make the inside of the tip wet with the solution.

To set the Prewet function, select the check box. Enter the value of "Volume" (amount of reagent suctioned).

(96-tip rack model: $0 \leq \text{Volume} \leq 100$) (384-tip rack model: $0 \leq \text{Volume} \leq 20$)

일 🔽 Prewet



AspirateStir

This stirs the solution in the well on the source plate. The solution is aspirated and discharged the specified number of times.

To set the AspirateStir function, select the check box. Enter the values of "Execution" (number of times solution is stirred) and "Volume" (amount of reagent suctioned). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

```
(1 \leq \text{Execution} \leq 5) (0 \leq \text{PostDelay} \leq 10000)
```

(96-tip rack model: 0≦Volume≦100) (384-tip rack model: 0≦Volume≦20)





5-85

Airgap

This delivers air on top of the solution inside the tip to drop the solution to the last drop.

To set the Airgap function, select the check box. Enter the value of "Volume" (air gap).

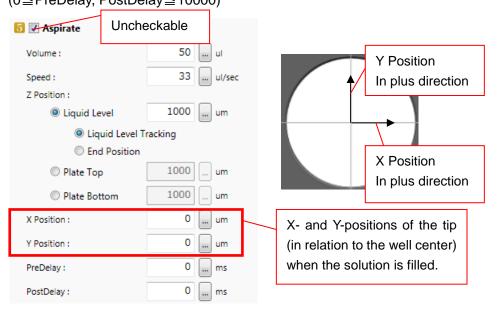
(96-tip rack model: 0≦Volume≦100) (384-tip rack model: 0≦Volume≦20)

🚺 🔽 Airgap		
Volume :	5 ul	Air gap
		Solution

Aspirate

This aspirates the solution from the well on the source plate.

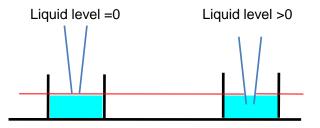
Enter the values of "Volume" (amount of reagent filled), "Speed" (filling speed) and "Z Position" (filling position). Also enter the values of "X Position" and "Y Position" if the tip position at the time of filling is to be specified. Enter the values of "PreDelay" (sleep time before the syringe operation) and "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled. (96-tip rack model: $0 \le$ Volume ≤ 100) (384-tip rack model: $0 \le$ Volume ≤ 20) (96-tip rack model: $0 \le$ Speed ≤ 34) (384-tip rack model: $0 \le$ Speed ≤ 7) (-50000 \le LiquidLevel, PlateTop, PlateBottom ≤ 100000) (-50000 \le X Position, Y Position ≤ 50000) ($0 \le$ PreDelay, PostDelay ≤ 10000)



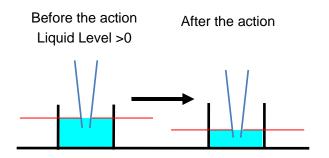
Z Position	Explanation
	Distance between the tip and liquid level
Liquid Level	(+ : Below the liquid level, 0 : Around the liquid level,
	- : Above the liquid level)
Liquid Level Tracking	Track the drop in liquid level due to filling.

End Position	Fill the solution by using the liquid level after filling as a
	reference.
	Fill the solution by using the top face of the plate as a
Diata Tan	reference.
Plate Top	(+ : Below the top face of the plate, 0 : Around the top
	face of the plate, - : Above the top face of the plate)
	Fill the solution by using the bottom face of the plate
	as a reference.
Plate Bottom	(+ : Below the bottom face of the plate, 0 : Around the
	bottom face of the plate, - : Above the bottom face of
	the plate)

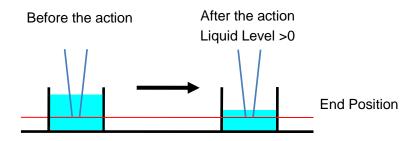
Liquid Level indicates the distance from the tip to the liquid surface. The plus indicates the downward tip position and the minus indicates the upward tip position in relation to the liquid level.



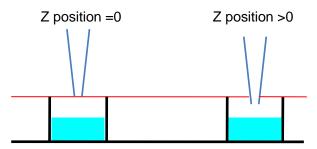
Liquid Level Tracking tracks the fall of the liquid level that occurs as the solution is filled and lowers the tip position in accordance with the fall of the liquid level. The relative position of the liquid level and tip is fixed.



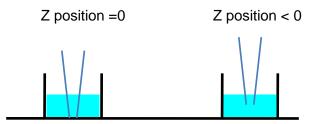
End Position sets the tip to a position that is shifted from the liquid level after the action by the entered liquid level.



•With regards to Plate Top, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the well top plane.



With regards to Plate Bottom, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the bottom plane of the well. Enter minus to set.



AspirateTiptouch

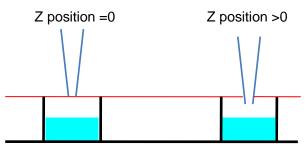
This lets the tip come in contact with the well wall and drops the droplet adhered to the end.

To set the AspirateTiptouch function, select the check box. Enter the value of "Z Position" (position at which the tip touches). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled. $(0 \le PlateTop \le 2000)$ ($0 \le PostDelay \le 10000$)

😚 🔽 AspirateTiptouch	
Z Position :	
CLiquid Level	2000 um
Plate Top	2000 🛄 um
PostDelay :	0 ms

Z Position	Explanation
	Distance between the tip and top face of the plate
Plate Top	(+ : Below the top face of the plate, 0 : Around the top
	face of the plate, - : Above the top face of the plate)

•With regards to Plate Top, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the well top plane.



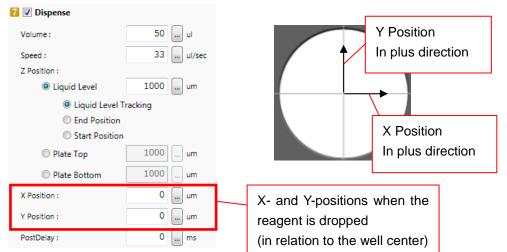
Dispense

This sets how to drop the reagent.

Enter the values of "Volume" (amount of reagent dripped into each well), "Speed" (dripping speed) and "Z Position" (dripping position). Also enter the values of "X Position" and "Y Position" if the tip position at the time of dripping is to be specified. Enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

(96-tip rack model: $0 \le \text{Volume} \le 100$) (384-tip rack model: $0 \le \text{Volume} \le 20$) (96-tip rack model: $0 \le \text{Speed} \le 34$) (384-tip rack model: $0 \le \text{Speed} \le 7$) (-50000 \le LiquidLevel, PlateTop, PlateBottom \le 100000) (50000 \le V Pagitian V Pagitian \le 50000) (0 \le PagetPalay \le 10000)

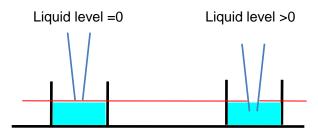
(-50000≦X Position, Y Position≦50000) (0≦PostDelay≦10000)



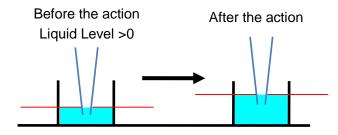
Z Position	Explanation
	Distance between the tip and liquid level
Liquid Level	(+ : Below the liquid level, 0 : Around the liquid level,
	- : Above the liquid level)
Liquid Level Tracking	Track the rise in liquid level due to dripping.
End Position	Drip the solution by using the liquid level after dripping as
	a reference.
Start Position	Drip the solution by using the liquid level before dripping
Start POSILION	as a reference.
	Drop by using the top face of the plate as a reference.
Plate Top	(+: Below the top face of the plate, 0: Around the top face
	of the plate, - : Above the top face of the plate)

	Drop by using the bottom face of the plate as a refer-
	ence.
Plate Bottom	(+ : Below the bottom face of the plate, 0 : Around the
	bottom face of the plate, - : Above the bottom face of the
	plate)

Liquid Level indicates the distance from the tip to the liquid surface. The plus indicates the downward tip position and the minus indicates the upward tip position in relation to the liquid level.

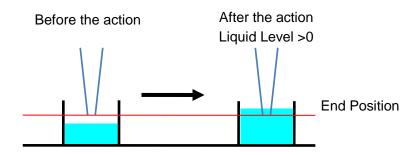


Liquid Level Tracking tracks the fall of the liquid level that occurs as the solution is filled and lowers the tip position in accordance with the fall of the liquid level. The relative position of the liquid level and tip is fixed.



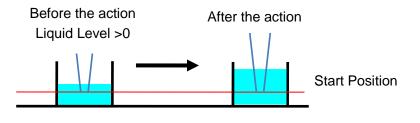
End Position sets the tip to a position that is shifted from the liquid level after the action by the entered liquid level.

The tip position does not change before and after the action.

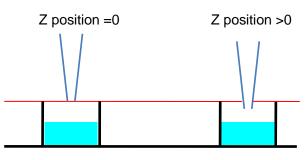


•Start Position sets the tip to a position that is shifted from the liquid level before the action by the liquid level you enter.

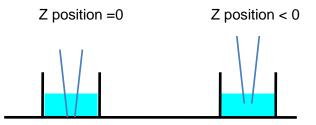
The tip position does not change before and after the action.



•With regards to Plate Top, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the top plane of the well.



With regards to Plate Bottom, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the bottom plane of the well. Enter minus to set.



DispenseStir

This stirs the dropped solution in the well. The solution in the well is aspirated and discharged the specified number of times.

To set the DispenseStir function, select the check box. Enter the values of "Execution" (number of times solution is stirred) and "Volume" (amount of reagent suctioned). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

 $(1 \leq \text{Execution} \leq 5) (0 \leq \text{PostDelay} \leq 10000)$

(96-tip rack model: 0≦Volume≦100) (384-tip rack model: 0≦Volume≦20)



DispenseTiptouch

This lets the tip come in contact with the well wall to drop the solution adhered to the end.

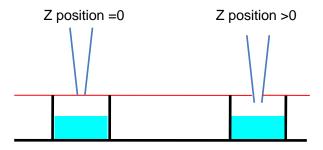
To set the DispenseTiptouch function, select the check box. Enter the value of "Z Position" (position at which the tip touches). Also enter the value of "PostDelay" (sleep time after the syringe operation) if any high-viscosity solution is handled.

 $(0 \leq PlateTop \leq 2000)$ $(0 \leq PostDelay \leq 10000)$

᠑ 📝 DispenseTiptouch	
Z Position :	
C Liquid Level	2000 um
Plate Top	2000 um
PostDelay :	0 ms

Z Position	Explanation
	Distance between the tip and top face of the plate
Plate Top	(+ : Below the top face of the plate, 0 : Around the top
	face of the plate, - : Above the top face of the plate)

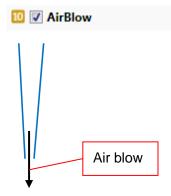
•With regards to Plate Top, the plus indicates the downward tip position and the minus indicates the upward tip position in relation to the well top plane.



AirBlow

This allows to blow air from inside tip to drop droplet adherent on the tip top. To set the AirBlow function, select the check box.

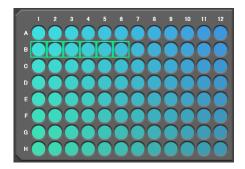
This function can be used only when liquid amount (including Airgap) inside tip is 0ul.



Auxiliary Input Function

This function is useful when entering the dispensing parameters for the wells in the specified range by changing their values at an equal distance, etc.

1) Specify a range of wells.



2) Click the button next to the entry box to open the auxiliary input screen.

일 🗸 Prewet	
Volume :	

ار ... 50

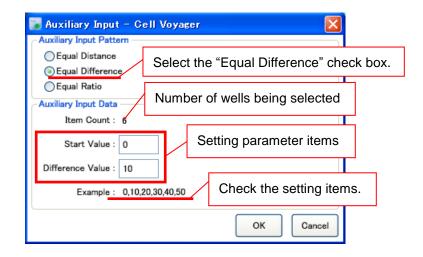
Equal Distance

Values are entered for the wells in the specified range at an equal distance. Enter the values of "Start Value" (value entered for the first well) and "End Value" (value entered for the last well).

🐻 Auxiliary Input – Cell Voyager 🛛 🔀
Auxiliary Input Pattern - Select the "Equal Distance" check box.
O Equal Difference
Cauciliary Input Data
Item Count : 6
Start Value : 5
End Value : 30
Example : 5,10,15,20,25,30 Check the setting items.
OK Cancel

Equal Difference

Values are entered for the wells in the specified range at an equal difference. Enter the values of "Start Value" (value entered for the first well) and "Difference Value" (value of equal difference).



Equal Ratio

Values are entered for the wells in the specified range at an equal ratio. Enter the values of "Start Value" (value entered for the first well) and "Ratio Value" (value of equal ratio).

🖥 Auxiliary Input – Cell Voyager 🛛 🔀
Auxiliary Input Pattern Cequal Distance Equal Distance Select the "Equal Ratio" check box.
Auxiliary Input Data
Item Count : 6
Start Value : 1 Setting parameter items
Ratio Value : 2 Example : 1.2.4.8.16.22 Check the setting items.
Example : 1,2,4,8,16,32
OK Cancel

5.14. Setting the Water Immersion Lenses

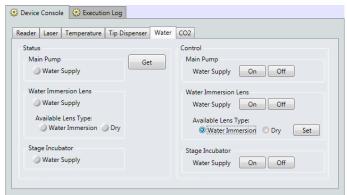
This setting is necessary to use water immersion lenses. (Water immersion lens model only)

1) Prepare the water supply bottle and the drainage bottle. Put pure water into the water supply bottle. (Pour off the water in the bottles regularly.)



VARNING WARNING

- In the case of replacing water in the bottles, make sure that the MAIN POWER breaker surely turns OFF to shut down CV7000. (Refer to 3.5) Be careful not to turn the breaker ON by oversight while at work.
- 2) Open the "Water" tab within the "Device Console" tab (refer to 6.4) on the "Reader Control" screen.



3) To supply the water in the main pump, click "On" of "Water Supply" in the "Main Pump" area. Wait for about ten minutes after clicking.

leader Laser Temperature Tip Disp	enser Water CO2
Status	Control
Main Pump	Get Main Pump
Water Supply	Water Supple On Off
Water Immersion Lens	Water Immersion Lens
Water Supply	Water Supply On Off
Available Lens Type:	Available Lens Type:
Water Immersion Ory	Water Immersion Dry Set
Stage Incubator	Stage Incubator
Water Supply	Water Supply On Off

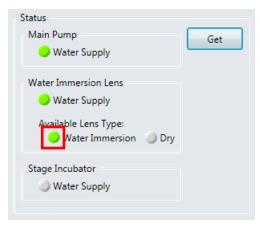
4) Click "On" of "Water Supply" in the "Water Immersion Lens" area.

😳 Device Console 🔅 Execution Log	
Device Console Consol	CO2 Control Main Pump Water Supply On Off Water Immersion Loss Water Supply On Off Available Lens Type: © Water Immersion Ory Set
Stage Incubator Water Supply	Stage Incubator Water Supply On Off

5) Select "Water Immersion" and click "Set" If you use the water immersion lenses. Select "Dry" and click "Set" if you use the dry lenses.

eader Laser Temperature Tip Di	spenser Water	CO2
Status		Control
Main Pump	Get	Main Pump
Water Supply		Water Supply On Off
Water Immersion Lens		Water Immersion Lens
Water Supply		Water Supply On Off
Available Lens Type:		Available Lens Type:
Water Immersion Ory		Water Immersion Ory Set
Stage Incubator		Stage Incubator
Water Supply		Water Supply On Off

6) Click "Get" and check the current status information. Confirm the illuminating green icon of "Water Immersion".



■You can use the dry lenses in the examples below.				
Status	Status			
Main Pump	Main Pump Get			
Vater Supply	Water Supply			
Water Immersion Lens	Water Immersion Lens			
Vater Supply	Water Supply			
Available Lens Type:	Available Lens Type:			
Water Immersion Ory	Water Immersion Ory			
Stage Incubator	Stage Incubator			
Water Supply	Water Supply Green			

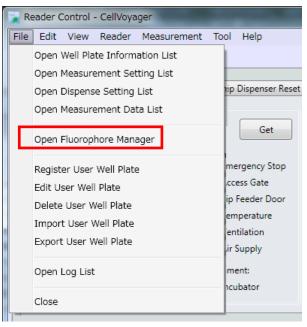
7) Click "Off" of "Water Supply" in the "Main Pump" area before the equipment is turned off. The water is drained away from the main pump.

ader Laser Temperature Tip Dispense	
Status	Control
Main Pump	Set Main Pump
Water Supply	Water Supply Off Off
Water Immersion Lens	Water Immersion Lens
Water Supply	Water Supply On Off
Available Lens Type:	Available Lens Type:
Water Immersion Ory	Water Immersion Dry Set
Stage Incubator	Stage Incubator
Water Supply	Water Supply On Off

5.15. Registering Fluorophore

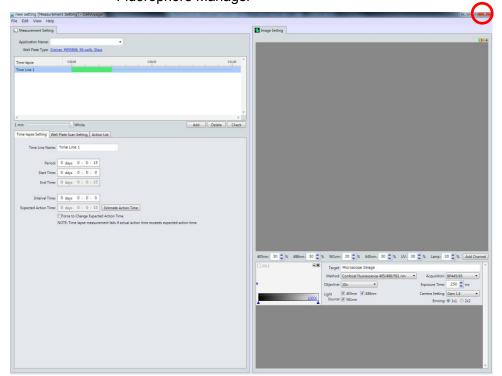
Register spectral information of fluorophore which is used to perform crosstalk correction by Image Correction Software

1) Select "File" -> "Open Fluorophore Manager" from menu of "Reader Control" window.





 "Open Fluorophore Manager" cannot be selected when "Measurement Setting" window is open.
 Close "Measurement Setting" window and select "Open Fluorophore Manager"



2) Click "Add Fluorophore"

Fluorophore Manager - CellVoyager	
Fluorophores Hoechist33342 mKusabira Orange2 Azami Green Alexa Fluor 488 MitoTracker Deep Red DAPI FusionRed_spectra	Add Fluorophore Delete Fluorophore
	OK Cancel

3) Load fluorophore spectral data file.



- Fluorephore spectral data file must meet following condition.
 - CSV format
 - It contains wavelength and emission spectral data.
 - Spectral data is normalized maximum value to be 100.
 - Data is arranged in the column direction as following. V

Wavelength	Emission	
300	0	
301	0	
302	0	
•	•	
•	•	
•	•	

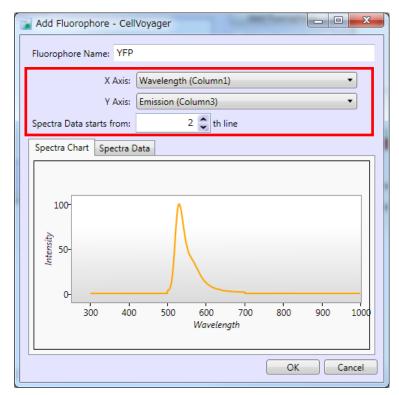
•Fluorephore spectral data file may be divided to excitation and emission. Load emission spectral data file.

4) Fluorophore spectral data file name is input in "Fluorophore Name" automatically. Change it as needed.

Add Fluorophore - Cell			
Fluorophore Name: YFP			
X Axis:	Wavelength (Column1)		
Y Axis: Emission (Column3)			
Spectra Data starts from: 2 🗘 th line			
Spectra Chart Spectra	Data		
100- 100- 100- 100- 100- 100- 100- 100-	0 500 600 700 800 900 1000 Wavelength		
	OK Cancel		

5) Select wavelength data column as "X Axis" and emission spectral data column as "Y Axis".

Input "Spectra Data starts from * th line".



Add Fluorophore - CellVoyager
Fluorophore Name: YFP
X Axis: Wavelength (Column1) Y Axis: Emission (Column3) Spectra Data starts from: 2 th line Spectra Chart Spectra Data
100- Aj 50-
0
Wavelength
OK Cancel

6) Confirm if there are no problems in emission spectral data and click "OK".

МЕМО

Following dialog is appeared if "Fluorophore Name" is same as that of already registered. Click "Yes" to overwrite emission spectral data. Click "No" to return to "Add Fluorophore" window.

CellVoyager Measurement System			x
YFP already exists. Do you want to overwrite it ?			
<u>Y</u> es	<u>N</u> o		

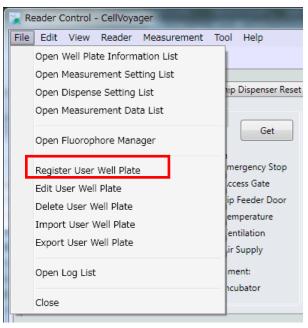
7) New fluorophore is added in "Fluorophores" list. Click "OK".

Fluorophore Manager - CellVoyager	
Fluorophores	Add Fluorophore
Hoechist33342	
mKusabira Orange2	Delete Fluorophore
Azami Green	
Alexa Fluor 488	
MitoTracker Deep Red	
DAPI	
EusionRed_spectra	
YFP	
	OK Cancel
	Cancer
And the second states and th	

5.16. Registering Well Plate

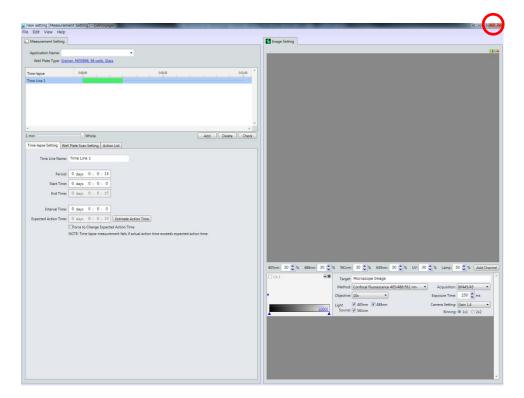
Register User Well Plate

1) Select "File" -> "Register User Well Plate" from menu of "Reader Control" window.



мемо

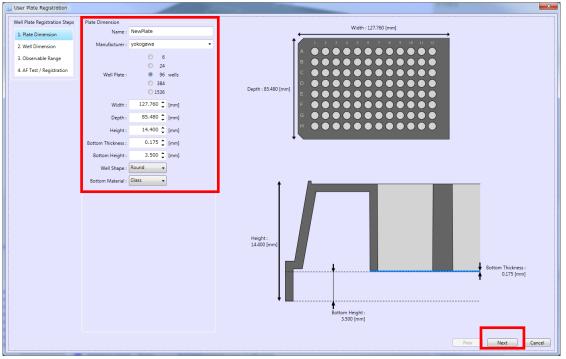
 "Register User Well Plate" cannot be selected when "Measurement Setting" window is open.
 Close "Measurement Setting" window and select "Register User Well Plate"



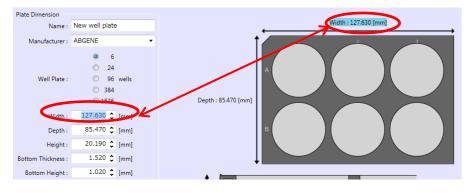


If water immersion objective lens is mounted, perform water supply for lens by referring page 6-43 before performing this procedure.

2) Input name, manufacturer, number of well, outer dimension, well shape and bottom material of well plate, then click "Next".

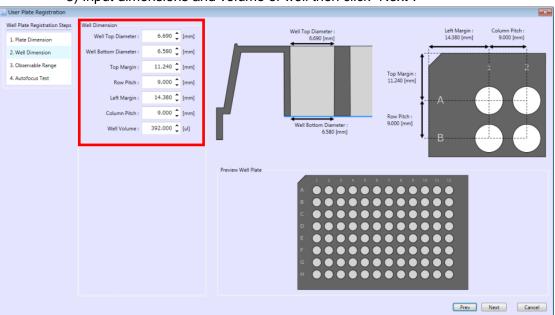


Correlated part is highlighted when dimension is input.



If incorrect value is input, red frame is shown and it cannot to go to next step.

Width :	150 🛟 [mm]
Depth :	85.470 🛟 [mm]
Height :	20.190 🛟 [mm]
Bottom Thickness :	1.520 🛟 [mm]
Bottom Height :	1.020 🛟 [mm]

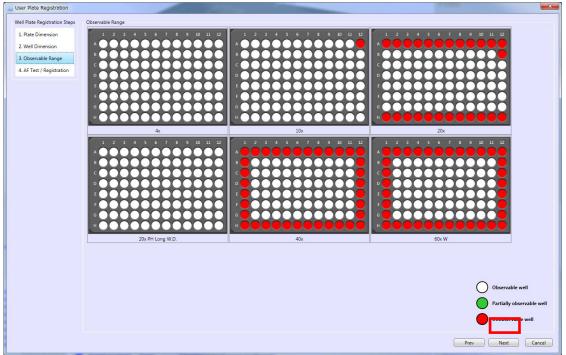


3) Input dimensions and volume of well then click "Next".

Correlated part is highlighted when dimension is input.



4) Observable range of using each objective lens is shown then click "Next".



Well Plate Registration Step	User Plate Registration		-			×
	Well Plate Registration Steps 1. Plate Dimension 2. Well Dimension 3. Observable Range	Name : Manufacturer : Wellplate : Weilt : Depth : Bottom Height : Bottom Height : Bottom Height : Well Sottom Diameter : Well Top Diameter : Top Margin : Row Pitch : Left Margin : Column Pitch :	yokogawa 96 wells 127.760 [mm] 85.480 [mm] 14.400 [mm] 0.175 [mm] 3.500 [mm] 6.580 [mm] 11.240 [mm] 9.000 [mm]	Plate Bottom : ① Detect upperside of the plate bottom. If not, detect downside of the plate bottom. ③ Detect only downside of the plate bottom.	Result	

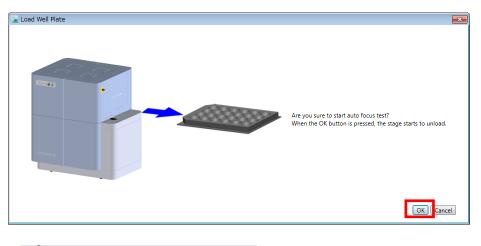
5) Confirm input well plate information.

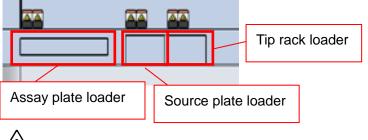
6) Test whether autofocus is available or not by using registered well plate. Normally, check "Detect upperside of the plate bottom. If not, detect down side of the plate bottom". In following case, check "Detect only downside of the plate bottom".

- Cell culture surface is treated specially. Click "Run AF Test".

AF Test	
Plate Bottom :	
 Detect upperside of the plate bottom. If not, detect downside of the plate bottom. Detect only downside of the plate bottom. 	
Run AF Test	

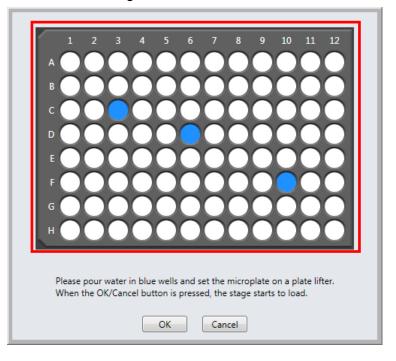
7) Click "OK" to move stage to the loader exit area.

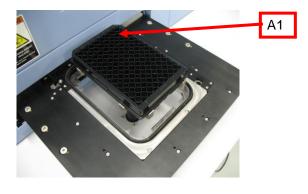




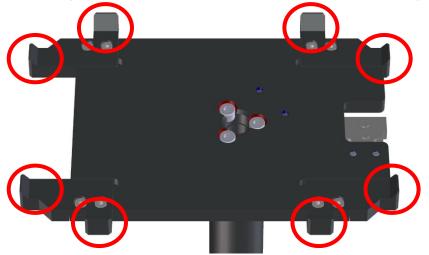
●DO NOT touch loader exit area while moving the loaders.

8) Prepare clean well plate, pour distillated water in designated well. Set the plate so that the well "A1" on the well plate comes to the top left-hand corner of the stage.

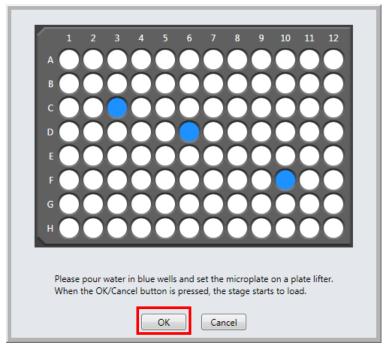




Set the assay plate so that its eight positions align with the eight positions of the stage. Failure to do so may cause measurement unit damage.



9) Click "OK" to start AF test.



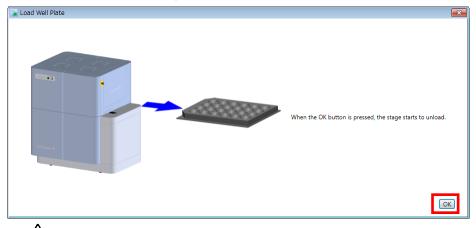
10) After AF test finishes, following window is shown. Click "OK".



It becomes error if AF test fails in every time or there is gap between measurement value and user input value of "Bottom height" or "Bottom thickness". In this case, the plate cannot be registered.

In this case, confirm that plate is clear and dimensions of plate are correct.

11) Click "OK" to move stage to the loader exit area.



/ WARNING

MEMO

Set the assay plate so that its eight positions align with the eight positions of the stage. Failure to do so may cause measurement unit damage.

12) If water immersion objective lens is mounted, bottom of well plate may be getting wet with water. If so, please wipe it.Click "OK" to house stage into instrument.



Il Plate Registration Steps	Well Plate Format		AF Test	Result	
Plate Dimension	Name :	NewPlate	Plate Bottom :	**************************************	A
	Manufacturer :	yokogawa	O Detect upperside of the plate bottom.	Stage unloaded. Stage loaded.	
Well Dimension)j	If not, detect downside of the plate bottom.	Use 20x PH Long W.D. for measuring Bottom Height and Bottom Thickness.	
Observable Range	Wellplate :	96 wells	Detect only downside of the plate bottom.	Measuring Bottom Height Success rate of Bottom Height measuring : 6 / 6	
AF Test / Registration	Width :	127.760 [mm]		Measuring Bottom Thickness	
Ar rest/ negistration	Depth :	85.480 [mm]	Run AF Test	Success rate of Bottom Thickness measuring : 6 / 6	
				Runing AF test using 4x AF Succeeded at : C03 (X= 0[um], Y= 0[um])	
	Height :	14.400 [mm]		AF Succeeded at : C03 (X=-1645[um], Y= 1645[um])	
	Bottom Thickness :	0.175 [mm]		AF Succeeded at : D06 (X= 0[um], Y= 0[um]) AF Succeeded at : D06 (X=-1645[um], Y= 1645[um])	
	Bottom Height :	3.500 [mm]		AF Succeeded at : D06 (A=-104-5[um], Y= 104-5[um]) AF Succeeded at : F10 (X= 0[um], Y= 0[um])	
	Bottom Material :	Plastic		AF Succeeded at : F10 (X=-1645[um], Y= 1645[um])	
	Well Shape :	Round		Runing AF test using 10x AF Succeeded at : C03 (X= 0[um], Y= 0[um])	
				AF Succeeded at : C03 (X=-1645[um], Y= 1645[um])	
	Well Top Diameter :	6.690 [mm]		AF Succeeded at : D06 (X= 0[um], Y= 0[um]) AF Succeeded at : D06 (X=-1645[um], Y= 1645[um])	
	Well Bottom Diameter :	6.580 [mm]		AF Succeeded at : D0b (X=-1645[um], Y= 1645[um]) AF Succeeded at : F10 (X= 0[um], Y= 0[um])	
	Top Margin :	11.240 [mm]		AF Succeeded at : F10 (X=-1645[um], Y= 1645[um])	1
	Row Pitch :	9.000 [mm]		Runing AF test using 20x AF Succeeded at : C03 (X= 0[um], Y= 0[um])	
	Left Margin :	14.380 [mm]		AF succeeded at : CO3 (X= 0[0In], Y= 0[0In]) AF Succeeded at : CO3 (X=-1645[um], Y= 1645[um])	
	Column Pitch :	9.000 [mm]		AF Succeeded at : D06 (X= 0[um], Y= 0[um])	
	Well Volume :			AF Succeeded at : D06 (X=-1645[um], Y= 1645[um]) AF Succeeded at : F10 (X= 0[um], Y= 0[um])	
	Well Volume :	392.0 [ul]		AF Succeeded at : 10 (X=-1645[um], Y= 1645[um])	
				Runing AF test using 20x PH Long W.D.	
				AF Succeeded at : C03 (X= 0[um], Y= 0[um]) AF Succeeded at : C03 (X=-1645[um], Y= 1645[um])	
				AF Succeeded at : D06 (X= 0[um], Y= 0[um])	
				AF Succeeded at : D06 (X=-1645[um], Y= 1645[um])	
				AF Succeeded at : F10 (X= 0[um], Y= 0[um]) AF Succeeded at : F10 (X=-1645[um], Y= 1645[um])	
				Runing AF test using 40x	
				AF Succeeded at : C03 (X= 0[um], Y= 0[um])	
				AF Succeeded at : C03 (X=-1645[um], Y= 1645[um]) AF Succeeded at : D06 (X= 0[um], Y= 0[um])	
				AF succeeded at : D06 (X= 0[um], Y= 0[um]) AF Succeeded at : D06 (X=-1645[um], Y= 1645[um])	
				AF Succeeded at : F10 (X = 0[um], Y = 0[um])	
				AF Succeeded at : F10 (X=-1645[um], Y= 1645[um])	
				Runing AF test using 60x W	
				AF Succeeded at : C03 (X= 0[um], Y= 0[um]) AF Succeeded at : C03 (X=-1645[um], Y= 1645[um])	
				AF Failed at : D06 (X= 0[um], Y= 0[um])	
				AE Esilad at - DOE IV _ 1645(-m1 V = 1645(-m1)	

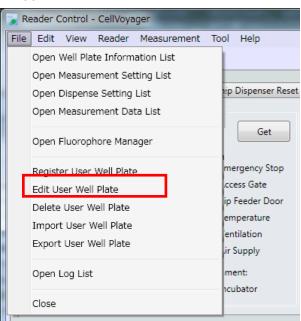
13) Click "Register".

14) Confirmation window for well plate registration is shown. If you should forbid editing or deleting information file of the registered well plate, check "Protect this well plate file". Click "OK" to finish registering.

Confirmation	
Are you sure to register this well plate ?	
Protect this well plate file.	
OK Cancel	

Edit User Well Plate

1) Select "File" -> "Edit User Well Plate" from menu of "Reader Control" window.





 "Edit User Well Plate" cannot be selected when "Measurement Setting" window is open.

Close "Measurement Setting" window and select "Edit User Well Plate"

leasurement Setting		Image Setting
pplication Name:		
Well Plate Type: Gre	einer, #655896, 96 wells, Glass	
e-lapse e Line 1	00000	
	Whole Add Delete	·
	ell Plate Scan Setting Action List	
Time Line Name:	s Time Line 1	
Period	t 0 days 0 : 0 : 15	
Start Time:	e 0 days 0 : 0 : 0	
End Time	e 0 days 0: 0:15	
	e: 0 days 0 : 0 : 0	
	e: 0 days 0 : 0 : 15 Estimate Action Time	
	Force to Change Expected Action Time NOTE: Time lapse measurement fails if actual action time exceeds expected action time.	
		405mm 30 C % 488mm 30 C % 561mm 30 C % 640mm 30 C % UV 30 C % Lampi 30 C % A
		Ch 1 Target: Microscope Image Method: Confocal Fluorescence 405/488/561 nm Acquisition: BP445/45
		Objective 10x Objective 10x Exposure Time: 250 © m
		Light 📝 405nm 📝 488nm Camera Setting: Gain 1.4
		10000 Source () 561nm Binning: @ 1x1 O 2

MEMO

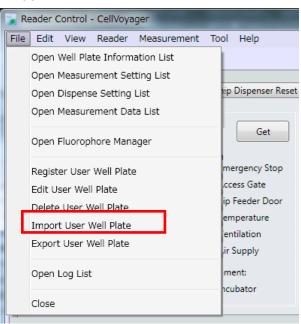
If water immersion objective lens is mounted, perform water supply for lens by referring page 6-43 before performing this procedure. 2) Select well plate information to edit. Edit and register well plate information by referring procedure from page 5-101 to page 5-108.



Name, Manufacturer and Well number cannot be changed in this procedure. If protected well plate information is edited, the well plate information is registered as another plate that Name, Manufacturer and Well number are same.

Import User Well Plate

 Select "File" -> "Import User Well Plate" from menu of "Reader Control" window.



МЕМО

 "Import User Well Plate" cannot be selected when "Measurement Setting" window is open.
 Close "Measurement Setting" window and select "Import User Well Plate"

idit View Help	
asurement Setting	Image Setting
plication Name:	
Well Plate Type: Greiner. #655896.96 wells. Glass	
-lapse 0.0000 000,00 *	
Line 1	
Whole Add Delete Check	
lapse Setting Weil Plate Scan Setting Action List	
Time Line Name: Time Line 1	
Period: 0 days 0: 0: 15	
Start Time: 0 days 0 : 0 : 0	
End Time 0 days 0 : 0 : 15	
Interval Time: 0 days 0 : 0 : 0	
sected Action Time: 0 days 0 : 0 : 15 Estimate Action Time	
Force to Change Expected Action Time	
NOTE: Time lapse measurement fails if actual action time exceeds expected action time.	
	405nm: 30 C % 488nm: 30 C % 561nm: 30 C % 640nm: 30 C % UV: 30 C % Lamp: 30 C % Ad
	Ch 1 Target: Microscope Image
	Method: Confocal Fluorescence 405/488/561 nm Acquisition: BP445/45
	Objective: 10x • Exposure Time: 250 🗘 ms
	Linke V 405nm V 488nm Camera Setting Gain 14
	10000 Source V 561nm Binning: @ 1x1 O 2x2

MEMO

If water immersion objective lens is mounted, perform water supply for lens by referring page 6-43 before performing this procedure. File selecting dialog is shown. Select to load well plate information file (.wpp). Register well plate information by referring procedure from page 5-101 to page 5-108.

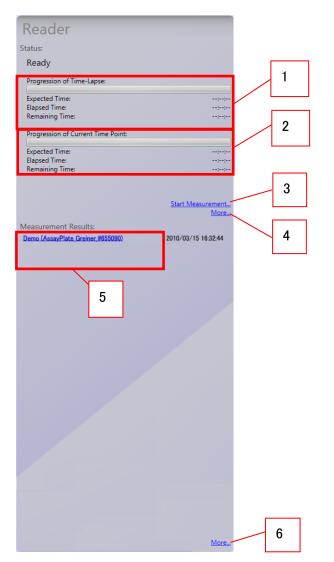


Name, Manufacturer and Well number cannot be changed in this procedure.

6. Explanation of Measurement Software Screens

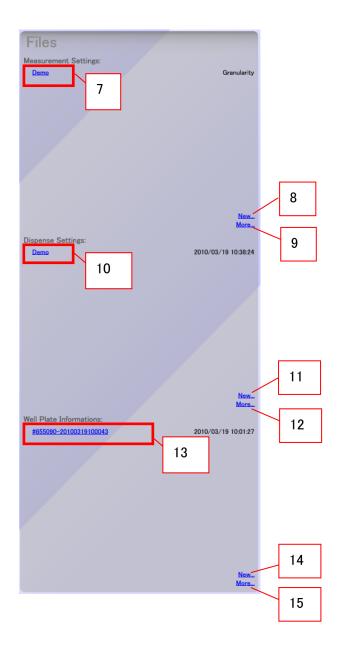
6.1. Main Screen

Reader area



- Progress status display of time-lapse measurement Expected Time : Time at which all measurement will end Elapsed Time : Elapsed time after start of measurement Remaining Time : Remaining time to end of measurement
- 2) Progress status display of processing at one time pointExpected Time : Time at which measurement at the time point will endElapsed Time : Elapsed time after start of measurement at the time pointRemaining Time : Remaining time to end of measurement at the time point
- 3) Open the screen to start measurement.
- 4) Open the Reader Control screen.
- 5) Recently measured or referenced data (Clicking here displays measured data.)
- 6) Browse measured data.

Files area



- Recently accessed measurement settings (Clicking here opens the edit screen.)
- 8) Create new measurement settings.
- 9) Edit measurement settings.
- 10) Recently accessed dispensing settings (Clicking here opens the edit screen.)
- 11) Create new dispensing settings.
- 12) Edit dispensing settings.
- 13) Recently accessed well plate information (Clicking here opens the edit screen.)
- 14) Create new well plate information.
- 15) Edit well plate information.

6.2. Measurement Setting File Screen

Image Setting 1 5 Add Delete Check 6 2 3 4 0 : 15 Estimate Action Time Force to Change Expected Action Time NOTE: Time lapse measurement fails if act 00 C 0 2 t Confocal Fluorescen Exposure Time: 250 📚 ms 7

Main Measurement Setting File Screen

1) Menus

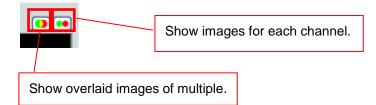
) МСПИЗ	
File menu	Explanation
Save	Save the measurement setting file under the same name.
Save As	Save the measurement setting file under a new name.
Close	Close the measurement setting file.

Edit menu	Explanation
Undo	Undo the last operation.
Redo	Redo the last operation.
Cut	Cut the selected item.
Сору	Copy the selected item.
Paste	Paste the selected item.

View menu	Explanation
Time Line Setting	Open the Time-lapse Setting tab.
Plate Scan Setting	Open the Well Plate Scan Setting tab.
Action List Setting	Open the Action List tab.
Show Overlay Images	Open the preview screen showing overlaid
Show Overlay Images	images of multiple channels.
Show Tilo Imagaa	Open the preview screen showing images
Show Tile Images	for each channel.

Help menu	Explanation
About	Show the version information of the measurement software.

- 2) Time-lapse Setting tab
- 3) Well Plate Scan Setting tab
- 4) Action List tab
- 5) Select the display format for channel images.



- 6) Preview screen
- 7) Imaging channel setting screen

Time-lapse Setting Screen

Measurement Setting								12121212
Application Name: Gra	nularity			_				
Well Plate Type: Grein	ner, #65509	0, 96 wells, Pla						
ime-lapse	0.00.00		2	0.00:30			0:01:00	-
ïme Line 1								
		3		4	5	6	7	
8		<u> </u>				۱Ľ ب	۱Ľــِـا	-
min		Vhole			Add	Copy	Delete	heck
								IICLK
					()	() (
	Plate Scan		on List			()(
	Plate Scan	Setting Acti	on List			()(
Time-lapse Setting Well	Plate Scan Time Line	Setting Acti	on List]			
Time-lapse Setting Well	Plate Scan Time Line	Setting Acti	on List					
Time-lapse Setting Well	Plate Scan Time Line 0 days	Setting Acti	on List					
Time-lapse Setting Well Time Line Name: Period:	Plate Scan Time Line 0 days 0 days	Setting Acti 2 1 0 : 0 : 43						
Time-lapse Setting Well Time Line Name: Period: Start Time:	Plate Scan Time Line 0 days 0 days 0 days	Setting Acti 1 0 : 0 : 43 0 : 0 : 0 0 : 0 : 43			9			
Time-lapse Setting Well Time Line Name: Period: Start Time:	Plate Scan Time Line 0 days 0 days 0 days	Setting Acti 2 1 0 : 0 : 43 0 : 0 : 0						
Time-lapse Setting Well Time Line Name: Period: Start Time: End Time:	Plate Scan Time Line 0 days 0 days 0 days 0 days	Setting Acti 1 0 : 0 : 43 0 : 0 : 0 0 : 0 : 43		ion Time)				

- 1) Application name
- 2) Well plate product name

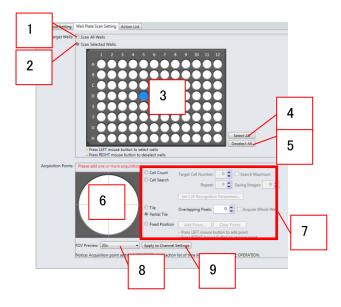
The well plate product can be changed by clicking here.

- 3) Time-line display area
- 4) Add a time line.
- 5) Copy the selected time line and add to the last of time line list.
- 6) Delete the selected time line.
- 7) Automatically and optimally arrange the time lines overlapping along the time axis
- 8) Change the display scale for time line.

9) Time-line setting items

Item	Explanation
Time Line Name	Specify the name of the time line.
Period	Specify the period of the time line, or duration after the
	start time until the time line ends.
Start Time	Specify the start time of the time line as, or the
	duration after the start of measurement until the first
	time point starts.
End Time	The end time of the time line is shown.
Interval Time	Set the duration after a time point ends until the next
	time point starts.
Expected Action Time	The expected processing time defined on the Action
	List tab is shown.
Estimate Action Time	The predicted processing time defined on the Action
	List tab is calculated when this button is clicked.
Force to Change	The Expected Action Time field can be entered.
Expected Action Time	

Well Plate Scan Setting Screen



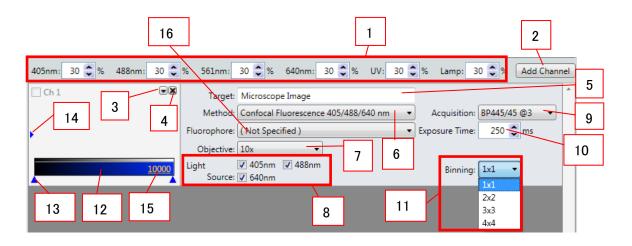
- 1) Measure all the wells.
- 2) Select wells to be measured.
- 3) Display area of wells to be measured
- 4) Select all the wells.
- 5) Unselect all the wells.
- 6) Imaging view field display area
- 7) Set the imaging view field.

Item	Explanation
Cell Count	An image is captured repeatedly while moving through the acquisition points, until the cell count entered in "Cell Number" is reached. Once the number of repetitions reaches the value in "Repeat," the system moves to the next well.
Cell Search	Move to the next acquisition point repeatedly until the value in "Repeat" is reached, to find the acquisition point associated with the largest cell count.
Target Cell Number	Specify the cell count.
Repeat	Specify the number of images to be captured for the same well.
Search Maximum	Save the images one by one, starting from the image associated with the largest cell count.
Saving Images	Specify the number of images to be saved.
Set Cell Recognition Parameters	Open the screen for setting the cell recognition algorithm.
Tile	Images are captured in a tiled manner. (whole region of well)
Partial Tile	Images are captured in a tiled manner. (desired region(s) of well)
Overlapping Pixels	Specify the number of overlapping pixels between images for tiled imaging.

Acquire Whole Well	Whole-well imaging is performed.
Fixed Position	Directly specify the positions of acquisition points.
Add Points	Specify the number and pitch of acquisition points, etc.
Clear Points	Clear the acquisition points.

- 8) Show the view of specified lens magnification.
- 9) Apply the lens magnification specified with "FOV Preview" to "Imaging Channel". (Refer to 5.5)

Imaging Channel Setting Screen



- 1) Set the laser power.
- 2) Create a new imaging channel.
- 3) Move the channel list.

Item	Explanation
Cut This Channel	Cut a channel list.
Copy This Channel	Copy a channel list.
Paste Channel Above	Paste the selected channel list above
Faste Channel Above	the one currently selected.
Paste Channel Below	Paste the selected channel list below
Faste Chamiler Delow	the one currently selected.

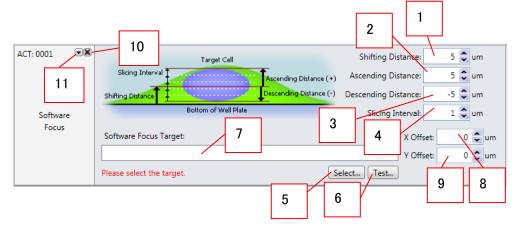
- 4) Delete the imaging channel.
- 5) Imaging target name
- 6) Select the optical system used for imaging.
- 7) Select the magnification factor.
- 8) Select the laser.
- 9) Select the fluorescence filter.
- 10) Camera exposure time
- 11) Camera binning
- 12) Adjust the image color.
- 13) Contrast bar
- 14) Expand the screen for contrast adjustment.
- 15) Select the maximum intensity for contrast adjustment.
- 16) Select fluorophore.

Action List Screen

Time-lapse Setting	Well Plate Scan Setting Action List
Software Focus A 1 Acquisition	Fluorescence Acquisition 2 3D Fluorescence Acquisition 2 3D Fluorescence Acquisition 2 3 3 3 5 5 4 5 5 6 4 5 5 6 4 5 5 6 4 5 5 6 7 6 4 5 5 6 7 6 7 6 7 7 6 7 7 8 7 7 7 8 7
8	-
	irement sequence to shorten time for measurement ace detection on AF
9	

- 1) Add the Action for Software Focus.
- 2) Add the Action for Fluorescence Acquisition.
- 3) Add the Action for 3D Fluorescence Acquisition.
- 4) Add the Action for Bright-field/Phase-contrast Acquisition.
- 5) Add the Action for Z-Stack Bright-field/Phase-contrast Acquisition.
- 6) Add the Action for DPC Acquisition.
- 7) Add the Action for Dispense Operation.
- 8) Optimize measurement sequence. (Refer to 7.3)
 - Checked: Optical switching is optimized to acquire images with the minimum time.
 - Unckecked: Optical switching is performed to acquire images at same time point.
- 9) Set to force undersurface detection on AF.
 - (Set when performing autofocus during high-speed time-lapse imaging)

Software Focus Screen

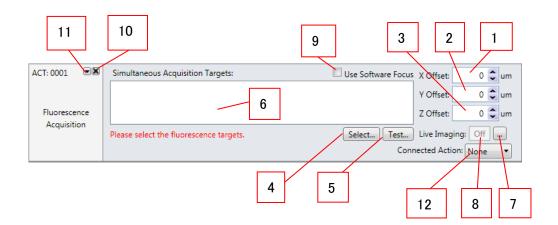


- 1) Reference plane of software focus
- 2) Upper limit of software focus
- 3) Lower limit of software focus
- 4) Z-axis pitch of software focus
- 5) Select the target channel for software focus.
- 6) Open the test preview screen.
- 7) The target channel for software focus is shown.
- 8) Fine-tune the X-axis of image positions.
- 9) Fine-tune the Y-axis of image positions.
- 10) Close the software focus screen.

11) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one
	currently selected.
Paste Action Below	Paste the selected action below the one
FASIE AUIUN DEIUW	currently selected.

Fluorescence Acquisition Screen



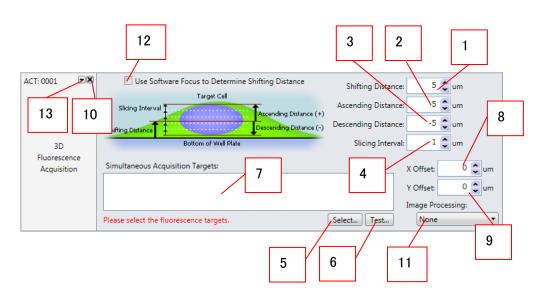
- 1) Fine-tune the X-axis of image positions.
- 2) Fine-tune the Y-axis of image positions.
- 3) Fine-tune the Z-axis of image positions.
- 4) Select the target channel for fluorescence imaging.
- 5) Open the test preview screen.
- 6) Show the target channel for fluorescence imaging.
- 7) Open the screen for setting high-speed time-lapse imaging.
- 8) Use condition of high-speed time-lapse imaging On : Use, Off : Do not use
- 9) Perform fluorescence imaging by using software focus plane as a reference.
- 10) Close the fluorescence imaging screen.

11) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one currently selected.
Paste Action Below	Paste the selected action below the one currently selected.

12) Set the connection to high-speed time-lapse imaging (refer to 7.15).

3D Fluorescence Acquisition Screen



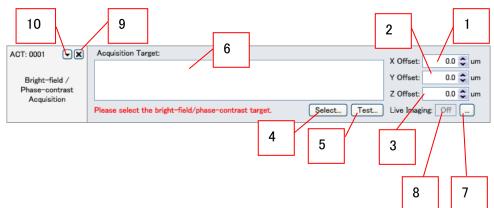
- 1) Reference plane of 3D imaging
- 2) Upper limit of 3D imaging
- 3) Lower limit of 3D imaging
- 4) Z-axis pitch of 3D imaging
- 5) Select the target channel for 3D imaging.
- 6) Open the test preview screen.
- 7) The target channel for 3D imaging is shown.
- 8) Fine-tune the X-axis of image positions.
- 9) Fine-tune the Y-axis of image positions.
- 10) Close the 3D imaging screen.
- 11) Select an output method for Z-stack images.

Item	Explanation
None	Acquired all Z images are saved.
Maximum	MIP images by Z-axis direction are saved.
Minimum	MinIP images by Z-axis direction are saved.
Average	AIP images by Z-axis direction are saved.
Sum	SUM images by Z-axis direction are saved.

12) Perform 3D imaging by using the software focus plane as a reference.13) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one
	currently selected.
Deate Action Balow	Paste the selected action below the one
Paste Action Below	currently selected.

Bright Field/Phase Contrast Acquisition Screen



1) Fine-tune the X-axis of image positions.

2) Fine-tune the Y-axis of image positions.

3) Fine-tune the Z-axis of image positions.

4) Select the target channel for bright field/phase contrast imaging.

5) Open the test preview screen.

6) The target channel for bright field/phase contrast imaging is shown.

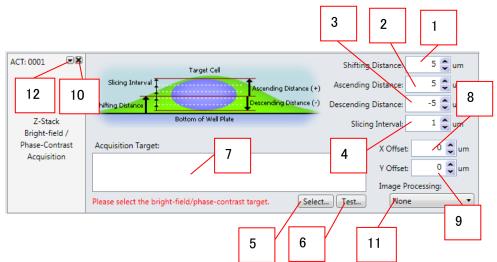
7) Open the screen for setting high-speed time-lapse imaging.

8) Use condition of high-speed time-lapse imaging - On : Use, Off : Do not use

9) Close the Bright field/Phase Contrast Acquisition screen.

10) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one currently selected.
Paste Action Below	Paste the selected action below the one currently selected.



Z-Stack Bright-field/Phase-contrast Acquisition

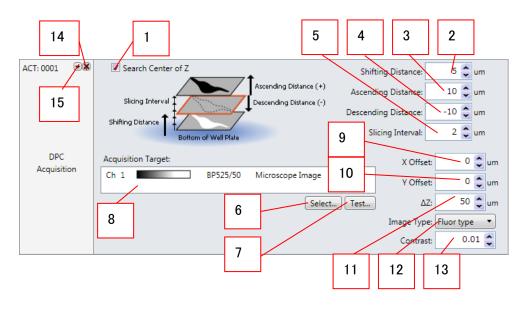
- 1) Reference plane of Z-stack imaging
- 2) Upper limit of Z-stack imaging
- 3) Lower limit of Z-stack imaging
- 4) Z-axis pitch of Z-stack imaging
- 5) Select the target channel for Z-stack imaging.
- 6) Open the test preview screen.
- 7) The target channel for Z-stack imaging is shown.
- 8) Fine-tune the X-axis of image positions.
- 9) Fine-tune the Y-axis of image positions.
- 10) Close the Z-Stack Bright-field/Phase-contrast Acquisition screen.
- 11) Select an output method for Z-stack images.

Item	Explanation
None	Acquired all Z images are saved.
Maximum	MIP images by Z-axis direction are saved.
Minimum	MinIP images by Z-axis direction are saved.
Average	AIP images by Z-axis direction are saved.
Sum	SUM images by Z-axis direction are saved.

12) Move the action list.

Item	Explanation	
Cut This Action	Cut an action	
Copy This Action	Copy an action	
Paste Action Above	Paste the selected action above the one currently selected.	
Paste Action Below	Paste the selected action below the one currently selected.	

DPC Acquisition



- 1) Select performing automatic DPC reference position search
- 2) Reference plane of automatic DPC reference position search

*In case that "Search Center of Z" is unchecked, this means DPC reference position

- 3) Upper limit of automatic DPC reference position search
- 4) Lower limit of automatic DPC reference position search
- 5) Z-axis pitch of automatic DPC reference position search
- 6) Select the target channel for DPC imaging.
- 7) Open the test preview screen.
- 8) The target channel for DPC imaging is shown.
- 9) Fine-tune the X-axis of image positions.
- 10) Fine-tune the Y-axis of image positions.

11) Set ΔZ (Z distance of multiple bright field images which are origins of DPC image)

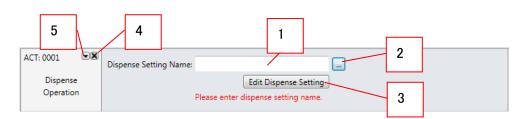
12) Select the type of DPC image

Item	Explanation
Fluor type	Fluorescence like DPC image.
Phase type	Phase contrast like DPC image.

- 13) Set contrast of DPC image to output.
- 14) Delete action of DPC Acquisition.
- 15) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one
	currently selected.
Paste Action Below	Paste the selected action below the one
Fasie Action Delow	currently selected.

Dispense Operation Screen



1) The dispensing setting file is shown.

2) Select an applicable dispensing setting file.

3) Edit the dispensing setting file to have been selected.

4) Close the dispensing screen.

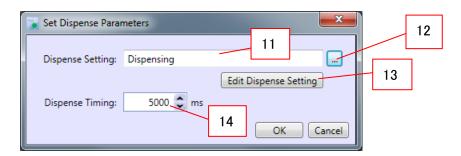
5) Move the action list.

Item	Explanation
Cut This Action	Cut an action
Copy This Action	Copy an action
Paste Action Above	Paste the selected action above the one
	currently selected.
Paste Action Below	Paste the selected action below the one
Faste Action Below	currently selected.

High-speed Time-lapse Setting Screen

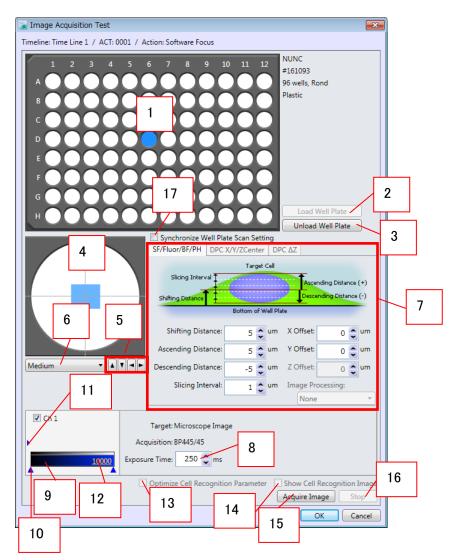
Set Live Imaging Pr	arameters 1		X
Ose Live Imaging:		2	
Period: Interval:	30000 🗘 ms 400 🗘 ms	`	4 5
Dispense Setting:	Dispense Setting Name Dispensing	Timing \$000 ms	Add 6 Change
Dispense Setting:			Delete 7
	Perform AF during live imaging	1	8
Test Interval Value			OK Cancel
	9 10]	

- 1) Selection of whether to use (Use Live Imaging) or not use (No Live Imaging) high-speed time-lapse imaging.
- 2) Period of high-speed time-lapse imaging
- 3) Interval of high-speed time-lapse imaging
- 4) Dispensing setting for high-speed time lapse imaging
- 5) Specify dispensing setting file (Set this item if dispensing is performed.)
- 6) Change dispensing setting file to have been registered.
- 7) Delete dispensing setting file to have been registered.
- 8) Display the Liquid Volume screen. (Refer to 8.4.)
- 9) Test to calculate "Interval" from the setting parameters such as exposure time, etc.
- 10) Selection of whether to perform autofocus during high-speed time-lapse imaging.



- 11) Dispensing setting file
- 12) Select dispensing setting file.
- 13) Edit the dispensing setting file to have been selected.
- 14) Timing at which to drip reagent after the start of high-speed time-lapse imaging (Set this item if dispensing is performed.)

Image Acquisition Test Setting Screen



1) Select the well used in the image-acquisition test.

2) Load the well plate.

3) Remove the well plate.

4) Select the view field for image-acquisition test.

5) Move the view field for image-acquisition test.

6) Select the moving speed of the view field for image-acquisition test.

7) Set imaging conditions. (Detail is shown in next page)

8) Camera exposure time

9) Adjust the image color.

10) Contrast bar

11) Expand the screen for contrast adjustment.

12) Select the maximum intensity for contrast adjustment.

13) Optimize the set values of recognition algorithm.

14) Show binary images.

15) Start the image-acquisition test.

16) Stop the image-acquisition test.

17) Reflect the wells observed on the preview screen, and the corresponding view field, in the Acquisition Points settings.

Image Acquisition Setting Screen (Acquisition Test Setting)

<Acquisition position setting

(Software Focus, Fluorescence/ BF/ PH Acquisition)>

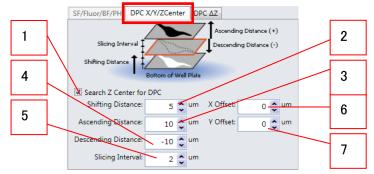


- 1) Reference plane of Z-stack imaging
- 2) Upper limit of Z-stack imaging
- 3) Lower limit of Z-stack imaging
- 4) Z-axis pitch of Z-stack imaging
- 5) Fine-tune the X-axis of image positions.
- 6) Fine-tune the Y-axis of image positions.
- 7) Fine-tune the Z-axis of image positions.

8) Select an output method for Z-stack images.

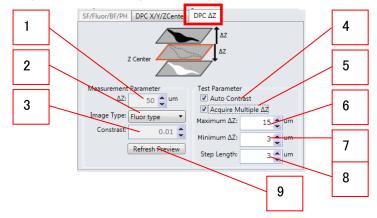
Item	Explanation
None	Acquired all Z images are saved.
Maximum	MIP images by Z-axis direction are saved.
Minimum	MinIP images by Z-axis direction are saved.
Average	AIP images by Z-axis direction are saved.
Sum	SUM images by Z-axis direction are saved.

<Acquisition position setting (DPC Acquisition)>



- 1) Select performing automatic DPC reference position search
- 2) Reference plane of automatic DPC reference position search
 - *In case that "Search Center of Z" is unchecked, this means DPC reference position
- 3) Upper limit of automatic DPC reference position search
- 4) Lower limit of automatic DPC reference position search
- 5) Z-axis pitch of automatic DPC reference position search
- 6) Fine-tune the X-axis of image positions.
- 7) Fine-tune the Y-axis of image positions.

<Image property setting (DPC Acquisition)>



1) Set ΔZ (Z distance of multiple bright field images which are origins of DPC image)

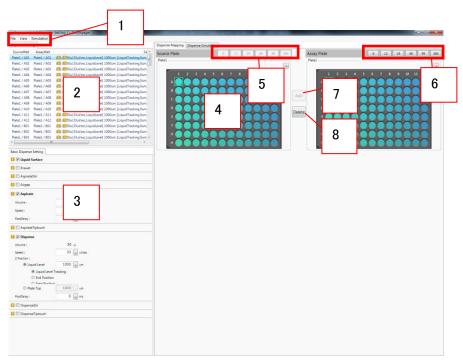
2) Select the type of DPC image

Item	Explanation
Fluor type	Fluorescence like DPC image.
Phase type	Phase contrast like DPC image.

- 3) Set contrast of DPC image to output.
- 4) Set auto contrast.
- 5) Select performing multiple ΔZ test acquisition.
- 6) Maximum ΔZ of multiple ΔZ test acquisition.
- 7) Minimum ΔZ of multiple ΔZ test acquisition.
- 8) ΔZ pitch of multiple ΔZ test acquisition.
- 9) Apply new contrast value to test images.

6.3. Dispensing Setting File Screen

Overview of Dispensing Setting File Screen



1) Menus

1) Мениз	
File	Explanation
Save	Save the dispensing setting file under the same name.
Save As	Save the dispensing setting file under a new name.
Close	Close the dispensing setting file.

View	Explanation
Dispense Mapping	Open the Dispense Mapping screen.
Dispense Simulation	Open the Dispense Simulation screen.
Basic Dispense Setting	Open the Basic Dispense Setting screen.
Advanced Dispense Setting	Open the Advanced Dispense Setting screen.

Simulation	Explanation
Start Simulation	Start simulation based on the dispensing settings.
Clear Simulation Log	Clear the simulation log.

- 2) Show the list of dispensing settings.
- 3) Dispensing settings for each well
- 4) Plate view screen
- 5) Add a well plate to the source plate view screen.
- 6) Add a well plate to the assay plate view screen.
- 7) Associate the source and assay plates.
- 8) Delete the association of source and assay plates.

Basic Setting Screen

1			
📘 🗹 Liquid Surface			
2 Rrewet	_	3	
Volume :	50 ul		
83		5	
Execution :	1	6	
Volume :	50 ul	0	
PostDelay :	0	7	
🚺 🛃 Airgap 8			
Volume :	5 ut	9	
5 V Aspirate			7
Volume :	50 ul	10	
Speed :	33 ul/sec	<u> </u>	1
Z Position :		11	
Liquid Level	1000 um		
Plate Bottom	1000 um	12	
PostDelay :	0 ms		
🜀 🗷 AspirateTiptouch		13	
14			

- 1) Liquid Surface function ON/OFF
- 2) Prewet function ON/OFF
- 3) Amount of solution suctioned in the Prewet mode
- 4) AspirateStir function ON/OFF
- 5) Number of times solution is stirred in the AspirateStir mode
- 6) Amount of solution suctioned in the AspirateStir mode
- 7) Sleep time after the syringe operation in the AspirateStir mode
- 8) Airgap function ON/OFF
- 9) Airgap volume
- 10) Amount of solution filled in the Aspirate mode
- 11) Filling speed in the Aspirate mode
- 12) Tip position at which solution is aspirated in the Aspirate mode Liquid Level: Track the drop in liquid level due to filling
 - The value to be entered means distance between the tip and

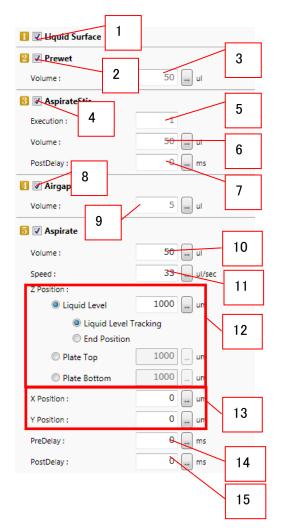
liquid level.

- Plate Bottom: Use the bottom face of the plate as a reference
- 13) Sleep time after the syringe operation in the Aspirate mode
- 14) AspirateTiptouch function ON/OFF

7 📝 Dispense	
Volume :	50 of 15
Speed :	33 ul/se 16
Z Position :	
Liquid Level	1000 um
Liquid Level T	racking
End Position	17
Start Position	
Plate Top	1000 um
Plate Bottom	1000 um
PostDelay :	0 ms 18
8 DispenseStir	
-	0 ms 18
B DispenseStir	0 ms 18
Execution :	0 ms 18 1 20 50 ul 21
Execution : 19	0 ms 18
DispenseStir Execution : 19 Volume : PostDelay :	0 ms 18 1 20 50 ul 21

- 15) Amount of solution dripped in the Dispense mode
- 16) Dripping speed in the Dispense mode
- 17) Tip position at which solution is dripped in the Dispense mode
 Liquid Level : Distance between the tip and liquid level
 Liquid Level Tracking : Track the rise in liquid level due to dripping
 End Position : Drip the solution from the liquid level after dripping
 Start Position : Drip the solution from the liquid level before dripping
 Plate Top : Use the top face of the plate as a reference
 Plate Bottom: Use the bottom face of the plate as a reference
- 18) Sleep time after the syringe operation in the Dispense mode
- 19) DispenseStir function ON/OFF
- 20) Number of times solution is stirred in the DispenseStir mode
- 21) Amount of solution suctioned in the DispenseStir mode
- 22) Sleep time after the syringe operation in the DispenseStir mode
- 23) DispenseTiptouch function ON/OFF
- 24) AirBlow function ON/OFF

Advanced Setting Screen



- 1) Liquid Surface function ON/OFF
- 2) Prewet function ON/OFF
- 3) Amount of solution suctioned in the Prewet mode
- 4) AspirateStir function ON/OFF
- 5) Number of times solution is stirred in the AspirateStir mode
- 6) Amount of solution suctioned in the AspirateStir mode
- 7) Sleep time after the syringe operation in the AspirateStir mode
- 8) Airgap function ON/OFF
- 9) Airgap volume
- 10) Amount of solution filled in the Aspirate mode
- 11) Filling speed in the Aspirate mode
- 12) Z position of the tip when filling in the Aspirate mode
 Liquid Level : Distance between the tip and liquid level
 Liquid Level Tracking : Track the drop in liquid level due to filling
 End Position : Fill the solution from the liquid level after filling
 Plate Top : Use the top face of the plate as a reference
 Plate Bottom: Use the bottom face of the plate as a reference
- 13) XY positions of the tip when filling in the Aspirate mode
- 14) Sleep time before the syringe operation in the Aspirate mode
- 15) Sleep time after the syringe operation in the Aspirate mode

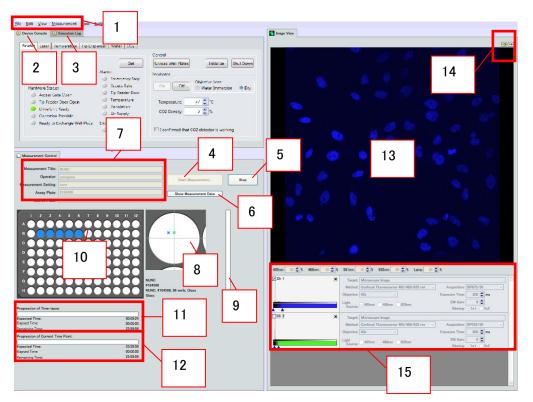
16		
6 🗹 AspirateTiptouch		
Z Position :		
O Liquid Level	2000 um	
Plate Top	2000 um	17
PostDelay :	0 ms	18
7 🗷 Dispense		19
Volume :	55 ul	13
Speed :	33 ul/sec	20
Z Position :		
Liquid Level	1000 um	
Liquid Level T	racking	
End Position		21
Start Position		21
Plate Top	1000 um	
Plate Bottom	1000 um	
X Position :	0 um	22
Y Position :	0 um	
PostDelay :	0 ms	23
8 DispenseStir		
Execution :	1-	25
Volume :	50 ul	26
PostDelay : 28	0 ms	27
IJ 🕑 DispenseTiptouch		
Z Position :		
Cliquid Level	2000 um	
Plate Top	2000 um	29
PostDelay :	0 ms	30
10 ⊮ AirBlow 31		

- 16) AspirateTiptouch function ON/OFF
- 17) Z position at which AspirateTiptouch is performed
 - Plate Top : Use the top face of the plate as a reference
- 18) Sleep time after the syringe operation in the AspirateTiptouch mode
- 19) Amount of solution dripped in the Dispense mode
- 20) Dripping speed in the Dispense mode
- 21) Tip position at which solution is dripped in the Dispense modeLiquid Level : Distance between the tip and liquid levelLiquid Level Tracking : Track the rise in liquid level due to dripping

End Position : Drip the solution from the liquid level after dripping Start Position : Drip the solution from the liquid level before dripping Plate Top : Use the top face of the plate as a reference

- Plate Bottom: Use the bottom face of the plate as a reference
- 22) XY positions of the tip when dripping in the Dispense mode
- 23) Sleep time after the syringe operation in the Dispense mode
- 24) DispenseStir function ON/OFF
- 25) Number of times solution is stirred in the DispenseStir mode
- 26) Amount of solution suctioned in the DispenseStir mode
- 27) Sleep time after the syringe operation in the DispenseStir mode
- 28) DispenseTiptouch function ON/OFF
- 29) Z position at which DispenseTiptouch is performed Plate Top : Use the top face of the plate as a reference
- 30) Sleep time after the syringe operation in the DispenseTiptouch mode
- 31) AirBlow function ON/OFF

6.4. Reader Control Screen



1) Menus

File menu	Explanation
Open Well Plate Information List	Open the screen for selecting a well plate
Open Well Plate Information List	information file.
Open Measurement Setting List	Open the screen for selecting a measurement
Open measurement Setting List	setting file.
Open Dispense Setting List	Open the screen for selecting a dispensing
Open Dispense Setting List	setting file.
Open Measurement Data List	Open the screen for selecting measured
	results.
Register User Well Plate	Register new well plate information.
Edit User Well Plate	Edit well plate information registered by user.
Delete User Well Plate	Delete well plate information registered by user.
Import User Well Plate	Import well plate information.
Export User Well Plate	Export well plate information registered by user.
Open Fluorophore Manager	Open the screen for registering fluorophore.
Open Log List	Open Log List screen for temperature and CO2.
Close	Close the Reader Control screen.

Edit menu	Explanation		
Undo	Undo the last operation.		
Redo	Redo the last operation.		
Cut	Cut the selected item.		
Сору	Copy the selected item.		
Paste	Paste the selected item.		

View menu	Explanation
Device Console	Open the Device Console tab.
Execution Log	Open the Execution Log tab.
Show Overlay Images	Show overlaid images of multiple channels.
Show Tile Images	Show images for each channel.

Measurement menu	Explanation		
Start Measurement	Start measurement.		
Stop Measurement	Stop measurement.		

Tool menu Explanation		
Default Image Correction	Show image correction in external control	
Setting for Automation	mode setting window	

Help menu	Explanation		
About	Show the version information of the measurement		
JUUGA	software.		

2) Open the screen for setting the laser, heater, etc.

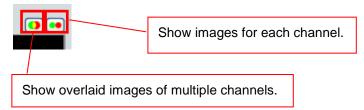
- 3) Open the log screen.
- 4) Start measurement.
- 5) Stop measurement.
- 6) Show measured results.

7) Information items entered at the time of measurement

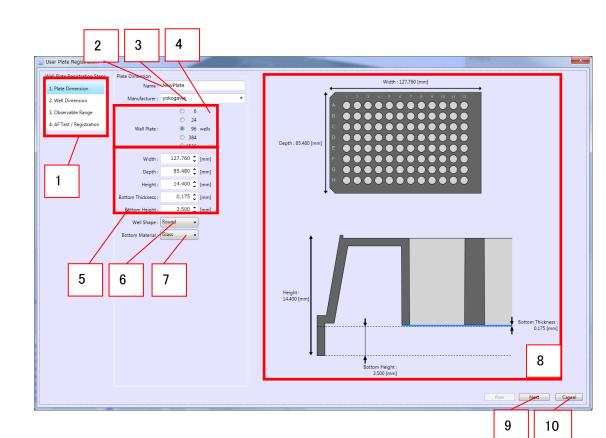
Item	Explanation		
Measurement Title	Title of measurement		
Operator	Name of the person who performed measurement		
Measurement Setting	Measurement setting file		
Assay Plate	Assay Plate name		
Source Plate	Source Plate name		

- 8) View field of measurement
- 9) Progress bar for measurement in Z-axis direction
- 10) The progress of measurement is indicated by well plates.
- 11) Progress status display of time-lapse measurementExpected Time : Expected time at which all measurement will endCurrent Time : Elapsed time after start of measurementRemaining Time : Remaining time to end of measurement
- 12) Progress status display of processing at one time pointExpected Time : Time at which measurement at the time point will endCurrent Time : Elapsed time after start of measurement at the time pointRemaining Time : Remaining time to end of measurement at the time point
- 13) Image currently being captured

14) Select the display format for channel images.



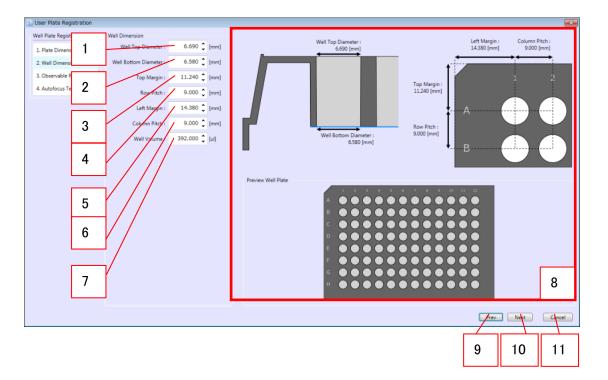
15) The channel settings and laser output are shown.



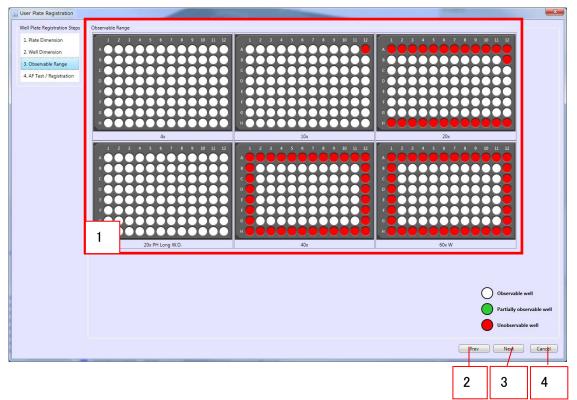
User Plate Registration Screen (Plate Dimension)

- 1) Step of registration of well plate information.
- 2) Name of well plate.
- 3) Manufacturer of well plate.
- 4) Number of wells.
- 5) Dimension of well plate.
- 6) Shape of well.
- 7) Bottom material of well plate.
- 8) Showing dimension and shape of well plate
- 9) Go to next step with saving changes.
- 10) Close the window without saving changes.





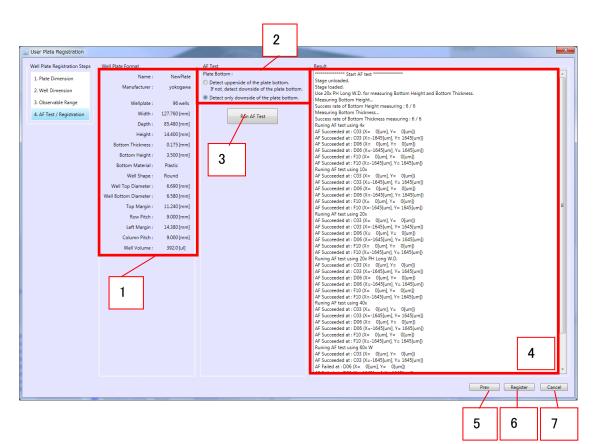
- 1) Diameter (round well)/ side length (rectangle well) of well top.
- 2) Diameter (round well)/ side length (rectangle well) of well bottom.
- 3) Distance between plate upper end and center of the first row well.
- 4) Row pitch of wells
- 5) Distance between plate left end and center of the first column well.
- 6) Column pitch of wells.
- 7) Volume of well.
- 8) Showing dimension and shape of well.
- 9) Return to previous step with saving changes.
- 10) Go to next step with saving changes.
- 11) Close the window without saving changes.



User Plate Registration Screen (Observable Range)

- Showing observable range of each objective lens. White: Whole well is observable Green: Partial of well is observable Red: Whole well is unobservable
- 2) Return to previous step.
- 3) Go to next step.
- 4) Close the window without saving changes.

User Plate Registration Screen (Autofocus Test)



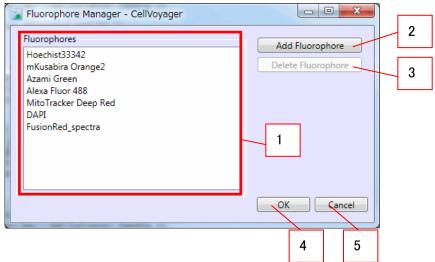
1) Well plate information.

2) Setting of auto focus detecting plane.

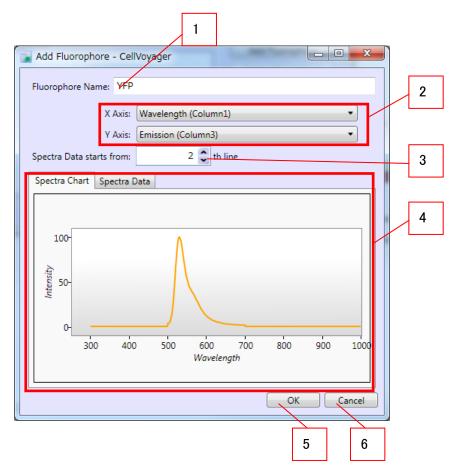
	1
	Explanation
Detect upperside of the plate bottom. If not, detect downside of the plate bottom.	Perform auto focus test on upper side of the plate bottom at first. If succeed, it is set that auto focus is performed on upper side of the plate bottom in this plate. In case of auto focus on upper side of the plate bottom fails, auto focus test on lower side of the plate bottom is performed. If succeed, it is set that auto focus is performed on lower side of the plate bottom in this plate. In case of auto focus on both sides of the plate bottom fails, information of this plate is not registered.
Detect only downside of the plate bottom.	Perform auto focus test on only lower side of the plate bottom at. If succeed, it is set that auto focus is performed on lower side of the plate bottom in this plate. In case of auto focus on lower side of the plate bottom fails, information of this plate is not registered.

- 3) Start auto focus test.
- 4) Result of auto focus test.
- 5) Return to previous step.
- 6) Close the window with registering plate information.
- 7) Close the window without registering plate information.

Fluorophore Manager Screen



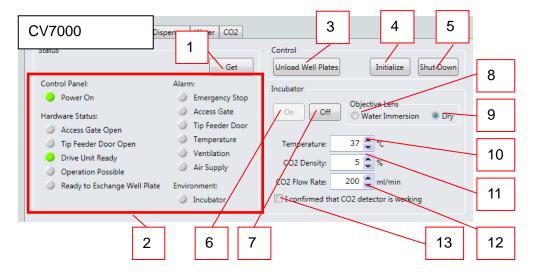
- 1) List of registered fluorophore.
- 2) Add fluorophore
- 3) Delete fluorophore.
- 4) Close the window with saving changes.
- 5) Close the window without saving changes.

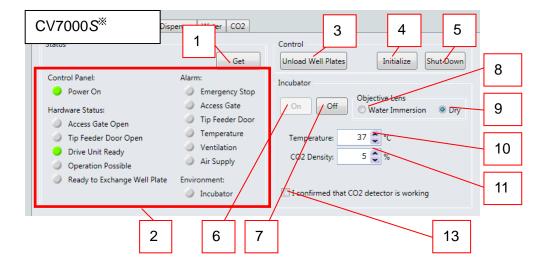


- 1) Fluorophore name.
- 2) Select wavelength data column and emission spectral data column.
- 3) Select line that wavelength data and emission spectral data start.
- 4) Display fluorophore spectral data.
 - Spectra Chart: Display graph Spectra Data: Display numeric data
- 5) Close the window with determining fluorophore spectral data.
- 6) Close the window without determining fluorophore spectral data.

Device Console Screen

Reader tab





Status information		Explanation	
Control Panel	Power On	The power is on.	
Hardware Status	Access Gate Open	The access gate is open.	
	Tip Feeder Door Open	The tip platform gate is open.	
	Drive Unit Ready	The device unit is ready.	
	Operation Possible	Accessible from the measurement	
		software.	
	Ready to Exchange	Well plates can be exchanged.	
	Well Plate	weil plates can be exchanged.	
Alarm	Emergency Stop	An emergency stop has been	
		actuated.	
	Access Gate	An access gate error has	
	Access Oale	occurred.	
	Tip Feeder Door	A tip platform gate error has	
		occurred.	
	Temperature	A temperature error has occurred.	
	Internal Environment	An internal environment fan error	
	Internal Environment	has occurred.	
	Air Supply	An air cylinder error has occurred.	
Environment Incubator		The incubator becomes stable.	

- 3) Move the stage to in front of the access gate on the front side of the measurement section.
- 4) Start/restart the system.
- 5) Shut down the system.
- 6) Turn on the stage incubator.
- 7) Turn off the stage incubator.
- 8) Select the water immersion lenses.
- 9) Select the dry lenses.
- 10) Enter the heater temperature
- 11) Enter the value of CO_2 concentration.
- 12) Enter the value of gas flow rate to the stage incubator.
- 13) Check box for confirmation that CO_2 detector is working. (If unchecked, CO_2 supply doesn't start.)

 ** Gas flow rate can't be set in CV7000S, which is sold after April 2015. (Refer to 3.4.)

Laser Tab

aser	Temperature	Tip Dispenser	Water	CO2				
_			_	Control				
m :				405nm :	Turn On	Turn Off		
m :		Get Status		445nm :	Turn On	Turn Off		
m :		Get Status		488nm :	Turn On	Turn Off		
m :		Get Status		561nm :	Turn On	Turn Off		
v:		Get Status		UV :	Turn On	Turn Off		
-							•	
	3	2			1			
	m : [m : [m : [V : [m : m :	m : Get Status m : Get Status m : Get Status m : Get Status V : Get Status	m : Get Status m : Get Status m : Get Status m : Get Status V : Get Status V : Get Status	m: Get Status 445nm : m: Get Status 445nm : m: Get Status 488nm : m: Get Status 561nm : V: Get Status UV :	m: Get Status 405nm : _Turn On m: Get Status 445nm : _Turn On m: Get Status 488nm : _Turn On m: Get Status 561nm : _Turn On V: Get Status UV : _Turn On	m: Get Status 405nm : Turn On Turn Off m: Get Status 445nm : Turn On Turn Off m: Get Status 488nm : Turn On Turn Off m: Get Status 561nm : Turn On Turn Off V: Get Status UV : Turn On Turn Off	m: Get Status 405nm : Turn On Turn Off m: Get Status 445nm : Turn On Turn Off m: Get Status 488nm : Turn On Turn Off m: Get Status 561nm : Turn On Turn Off V: Get Status UV : Turn On Turn Off

- Start/shut down the laser (Turn On/Turn Off)
 405nm : 405 nm wavelength laser
 488nm : 488 nm wavelength laser
 561nm(or 532nm) : 561 nm(or 532nm) wavelength laser
 640nm : 640 nm wavelength laser
- 2) Acquire laser status information.
- 3) Laser status information is shown.
 - Turn On : Laser is already running.
 - Turn Off : Laser is off.
 - Starting : Laser is starting.

Temperature Tab

Reader Laser Temperature Tip Dispenser Water CO2	1 2
Status Target: 8 7 Get	Control Target: 37 C Set 3
Heater Temperature: Objective 1 °C Top of Assay Plate °C Objective 2 °C Middle of Assay Plate °C Objective 3 °C Bottom of Assay Plate °C Objective 4 °C Source Plate Shuttle °C Objective 5 °C Objective 6 °C	Turn on/off Heater: Set Objective Top of Assay Plate Middle of Assay Plate Bottom of Assay Plate Source Plate Shuttle Select All Deselect All
9	4 5 6

1) Set heater temperature

2) Reflect the set heater temperature.

3) Turn on the heater for the selected targets of temperature control.

4) Select the targets of temperature control.

Temperature setting location	Explanation
Objective	Object lens heater
Top of Assay Plate	Assay plate top stage heater
Middle of Assay Plate	Assay plate center stage heater
Bottom of Assay Plate	Assay plate bottom stage heater
Source Plate Shuttle	Source plate heater

5) Select all as targets of temperature control.

- 6) Unselect all.
- 7) Acquire status information.
- 8) Set temperature
- 9) Temperature conditions

Item	Explanation
Objective 1	The temperature of the first positional object lens
Objective 2	The temperature of the second positional object lens
Objective 3	The temperature of the third positional object lens
Objective 4	The temperature of the fourth positional object lens
Objective 5	The temperature of the fifth positional object lens
Objective 6	The temperature of the sixth positional object lens
Top of Assay Plate	Assay plate top stage temperature
Middle of Assay Plate	Assay plate center stage temperature
Bottom of Assay Plate	Assay plate bottom stage temperature
Source Plate Shuttle	Source plate stage temperature

Tip Dispenser tab

Reader Laser Temperature Tip Dispenser Wa	
Status	
Syringe Usage Count :	
Tip Rack Status: 2	
Tip Status:	
Remaining Tips:	
5 Get Status 4	

1) Number of times that dispenser syringe is used.



After number of times that syringe is used reach 80 % of operating life, caution is displayed on "System Log".

System Log		
02015/09/03 18:04:58	Reader	Reader device monitor has been started.
2015/09/03 18:04:58	Reader	Dispenser syringe usage count has approached the recommended limit : 1801/2000, Usage rate : 90%
02015/09/03 18:04:58	Reader	UV laser getting turned on. Please wait at least 5 minutes.

It is recommended to exchange syringe when this caution appears. Please contact us.

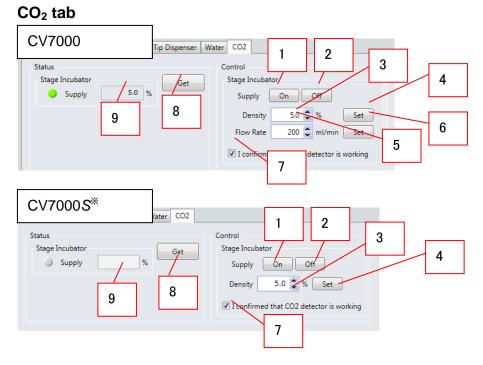
- 2) Whether or not there is a tip rack at the dispenser (Ready/Not Ready/Not Available)
- 3) Tip setting condition at the dispenser (Attached/Not Attached)
- 4) Number of tips remaining on the tip rack at the dispenser (96-tip rack model: 0 to 96) (384-tip rack model: 0 to 384)
- 5) Acquire status information.

10 D2 Reader Laser Temperature Tip Dispenser 2 1 Statu Control Main Pump Main Pump Get Water Supply Off Water Supply On / 3 Water Immersion Lens Water Immersion Lens Water Supply Water Supply On Off 4 5 Available Lens Type: Available Lens Type: 🔵 Water Immersion 🔵 Dry Water Immersion Opry Set 7 Stage Incubator Stage Incubator Water Supply Off Water Supply On 6 9 11 8

Water tab

- 1) The water is supplied to the main pump.
- 2) The water is discharged from the main pump.
- 3) The water is supplied to the water immersion lenses.
- 4) The water is not supplied to the water immersion lenses.
- 5) Select the water immersion lenses.
- 6) Select the dry lenses.
- 7) Set the selected lens type.
- 8) The water is supplied to the stage incubator.
- 9) The water is not supplied to the stage incubator.
- 10) Acquire status information.
- 11) Status information is shown.

Status information			Explanation	
Main Pump		Water Supply	Water supply to the main pump has	
			been executed.	
v	Vater Immersion Lens	Water Supply	Water supply to the water immersion	
v		Water Supply	lenses has been executed.	
	Available Lens Type	Water Immersion	Water Immersion lenses can be	
			used.	
		Dry	Dry lenses can be used.	
Stage Incubator		Water Supply	Water supply to the stage incubator	
			has been executed.	



- 1) CO₂ is supplied to the stage incubator.
- 2) CO_2 is not supplied to the stage incubator.
- 3) Enter the value of CO₂ concentration.
- 4) Set the entered CO₂ concentration.
- 5) Enter the value of gas flow rate to the stage incubator.
- 6) Set the value of gas flow rate.
- Check box for confirmation that CO₂ detector is working. (If unchecked, CO₂ supply doesn't start.)
- 8) Acquire status information.
- 9) Status information for CO₂ concentration to be supplied.
- * Gas flow rate can't be set in CV7000S, which is sold after April 2015. (Refer to 3.4.)

7. Setting Examples of Measurement Setting Files

7.1. Imaging by Auto-focus

Images are captured without using the software focus.

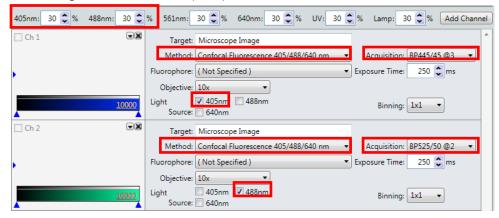
Confocal Imaging by Auto-focus

Confocal imaging is performed on the auto-focus plane at two wavelengths (405 nm, 488 nm).

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

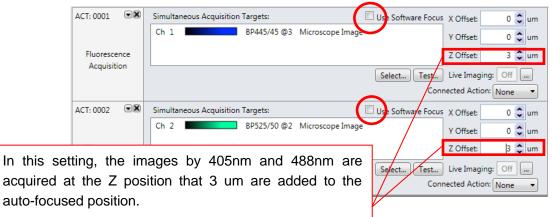
Application Name:	Granularity		
Well Plate Type:	Greiner, #655896, 96 wells, Glass		

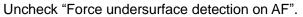
- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45
 - Light Source: 405nm (30%)
 - Ch2 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP525/50
 - Light Source: 488nm (30%)

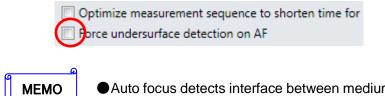


4) Set the items on the Action List tab. (Refer to 5.7)

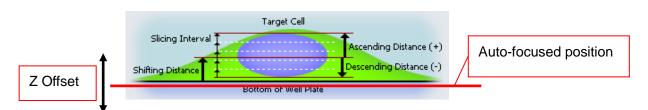
Set Ch1 and Ch2 for the two Fluorescence Acquisition tasks, respectively. Unselect the "Use Software Focus" check box.



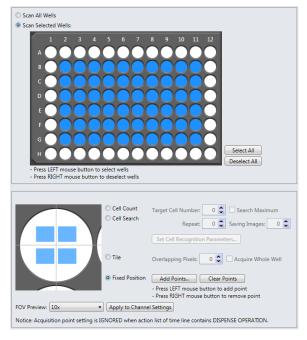




Auto focus detects interface between medium and glass by the auto focus unit.



5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)



6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Per	iod: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Expected Action Ti	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

Bright Field Imaging by Auto-focus

Bright field imaging is performed on the auto-focus plane.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



3) Set the imaging channels. (Refer to 5.5)Ch1 Method: BrightfieldLight Source: Lamp (5%)

405nm: 30 🔷 % 488nm: 30 🗘	% 561nm: 30 🔷 % 640nm: 30 🔷 %	UV: 30 🔷 % Lamp: 5 🔷 % Add Channel
Ch 1	Target: Microscope Image	A
	Method: Brightfield	✓ Acquisition: BP445/45 @3 ▼
•		Exposure Time: 250 🗢 ms
	Objective: 10x 🔹	
10000	Light Source: 🔽 Lamp	Binning: 1x1 🗸

4) Set the items on the Action List tab. (Refer to 5.7) Set Ch1 for the Bright-field/Phase-contrast Acquisition task.

ACT: 0001	Acquisition Target:	X Offset:	0 🛟 um
Bright-field /	Ch 1 BP525/50 Microscope Image	Y Offset:	0 🗘 um
Phase-contrast		Z Offset:	0 🛟 um
Acquisition	Select	Live Imag	ing: Off

Uncheck "Force undersurface detection on AF".

Optimize measurement sequence to shorten time for porce undersurface detection on AF

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6.)

6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Per	iod: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Interval II	lo days o : o : o
Expected Action Ti	me: 0 days 0 : 0 : 04 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

Z-Stack Imaging by Bright Field

Z-stack imaging by bright field is performed around the auto-focus plane.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



3) Set the imaging channels. (Refer to 5.5) Ch1 Method: Brightfield Light Source: Lamp (5%)

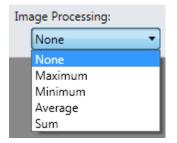
405nm: 30 🔷 % 488nm: 30 🗘	% 561nm: 30 🔷 % 640nm: 30 🔷 %	UV: 30 🔷 % Lamp: 5 🔷 % Add Channel
Ch 1	Target: Microscope Image	*
	Method: Brightfield	✓ Acquisition: BP445/45 @3 ▼
•		Exposure Time: 250 🖨 ms
	Objective: 10x 🔹	
10000	Light Source: 🔽 Lamp	Binning: 1x1 -

4) Set the items on the Action List tab. (Refer to 5.7)

Set Ch1 to Z-Stack Bright-field/Phase-Contrast Acquisition

ACT: 0001	Target Cell Shifting Distance:	5 🛟 um
	Slicing Interval Ascending Distance (+) Ascending Distance:	5 🛟 um
	Shifting Distance Descending Distance (-) Descending Distance:	-5 🛟 um
Z-Stack Bright-field /	Bottom of Well Plate Slicing Interval:	1 🛟 um
Phase-Contrast	Acquisition Target: X Offset:	0 🛟 um
Acquisition	Ch 1 BP525/50 Microscope Image Y Offset:	0 🔷 um
	Image Proces	ssing:
	Select Test None	

Select an output method for Z images from "Image Processing." (Refer to 5.1 and 6.2.)



Uncheck "Force undersurface detection on AF".

Optimize measurement sequence to shorten time for brce undersurface detection on AF

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6.)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List	
Time Line Nam	: Time Line 1	
Perio	t: 0 days 0 : 0 : 10	
Start Tim	: 0 days 0: 0: 0	
End Tim	e: 0 days 0 : 0 : 10	
Interval Tim	e: 0 days 0: 0: 0	
Expected Action Tim	e: 0 days 0 : 0 : 11 Estimate Action Time	>

3D Imaging from the Auto-focus Plane

3D imaging is performed at two wavelengths (405 nm, 488 nm) by using the auto-focus plane as the reference.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



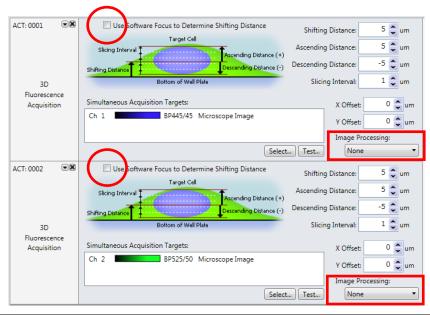
- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%)
 - Ch2 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP525/50 Light Source: 488nm (30%)

405nm: 30 🗢 % 488nm: 30 🗢	% 561nm: 30 🗢 % 640nm: 30 🗢 % UV: 30 🗢 % Lamp: 30 🗢 % Add Channel
Ch 1	Target: Microscope Image
	Method: Confocal Fluorescence 405/488/640 nm 🔹 Acquisition: BP445/45 @3 🔹
•	Fluorophore: (Not Specified) Exposure Time: 250 ms
	Objective: 10x
<u>10000</u>	Light V 405nn 488nm Binning: 1x1 V Source: 640nm
🗌 Ch 2	Target: Microscope Image
	Method: Confocal Fluorescence 405/488/640 nm 🔹 Acquisition: BP525/50 @2 🔹
•	Fluorophore: (Not Specified) Exposure Time: 250 ms
	Objective: 10x 🔻
10000	Light 🔲 405nm 📝 488nm Binning: 1x1 👻

4) Set the items on the Action List tab. (Refer to 5.7)

Set Ch1 and Ch2 for the two 3D Fluorescence Acquisition tasks, respectively.

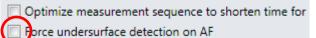
Unselect the "Use Software Focus to Determine Shifting Distance" check box.



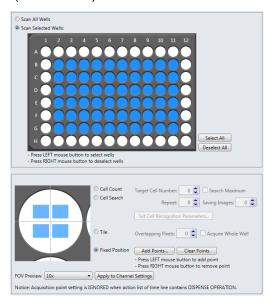
Select an output method for Z images from "Image Processing." (Refer to 5.1 and 6.2.)

Ima	age Processing:
	None 🔻
	None
	Maximum
	Minimum
	Average
	Sum

Uncheck "Force undersurface detection on AF".



5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)



6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	ime: Time Line 1
Per	iod: 0 days 0 : 0 : 24
Start Ti	ime: 0 days 0 : 0 : 0
otore in	
End Ti	ime: 0 days 0 : 0 : 24
Interval Ti	ime: 0 days 0 : 0 : 0
Expected Action Ti	ime: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

Epifluorescence Imaging by Auto-focus

Epifluorescence imaging is performed on the auto-focus plane.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



3) Set the imaging channels. (Refer to 5.5) Ch1 Method: Epifluorescence UV Lamp (UV model only) Light Source: UV (30%)

405nm: 30 🗘 % 488nm: 30 🗘	% 561nm: 30 🔷 % 640nm: 30 🗘 %	UV: 30 🗘 % Lamp: 5 🗘 % Add Channel
Ch 1	Target: Microscope Image	*
	Method: Epifluorescence UV Lamp	✓ Acquisition: BP445/45 @3 ▼
•	Fluorophore: (Not Specified)	 Exposure Time: 250 Sms
	Objective: 10x 🔹	
10000	Light Source: VV	Binning: 1x1 -

4) Set the items on the Action List tab. (Refer to 5.7) Set Ch1 for the Fluorescence Acquisition task.

ACT: 0001	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🗘 um
	Ch 1 BP445/45 @3	Microscope Image Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 🗘 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None

Uncheck "Force undersurface detection on AF".

Optimize measurement sequence to shorten time for porce undersurface detection on AF

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)

6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Peri	od: 0 days 0 : 0 : 24
Start Tir	me: 0 days 0 : 0 : 0
End Tir	me: 0 days 0 : 0 : 24
Interval Tir	
Expected Action Tir	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.2. Imaging by Software Focus

Imaging is performed on the software focus plane.

Confocal Imaging by Software Focus

Confocal imaging is performed on the software focus plane at two wavelengths (405 nm, 488 nm).

¥

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

Application Name: Granularity Well Plate Type: Greiner, #655896, 96 wells, Glass

- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%)

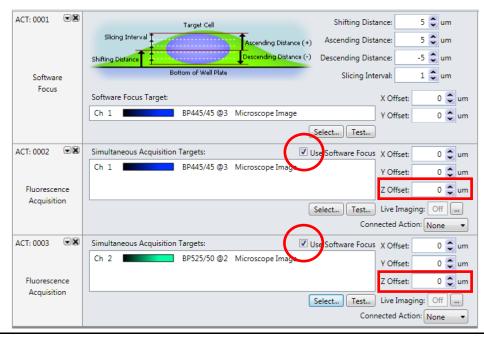
Ch2 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP525/50 Light Source: 488nm (30%)

405nm: 30 🗘 % 488nm: 30 🗘	% 561nm: 30 🗘 % 640nm: 30 🗘 % UV: 30 🗘	% Lamp: 30 🗢 % Add Channel
Ch 1	Target: Microscope Image	*
	Method: Confocal Fluorescence 405/488/640 nm 🔹	Acquisition: BP445/45 @3 🔹
•	Fluorophore: (Not Specified)	Exposure Time: 250 🗢 ms
10000	Objective: 10x Light 405nm 488nm Source: 640nm	Binning: 1x1 🔻
Ch 2	Target: Microscope Image	
	Method: Confocal Fluorescence 405/488/640 nm 🔹	Acquisition: BP525/50 @2 🔹
•	Fluorophore: (Not Specified)	Exposure Time: 250 🗢 ms
10000	Objective: 10x Light 405nm 488nm Source: 640nm	Binning: 1x1 💌

4) Set the items on the Action List tab. (Refer to 5.7)

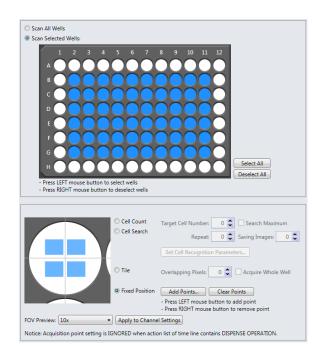
Software focus is applied to Ch1. Images are captured for Ch1 and Ch2 on the software focus plane. To capture images on the software focus plane, set "0" under "Z Offset" for "Fluorescence Acquisition."



Set "Force undersurface detection on AF". Normally, uncheck this box. If high-speed time-lapse imaging with operating autofocus is performed after software focus, check this box.

Optimize measurement sequence to shorten time for Force undersurface detection on AF

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)

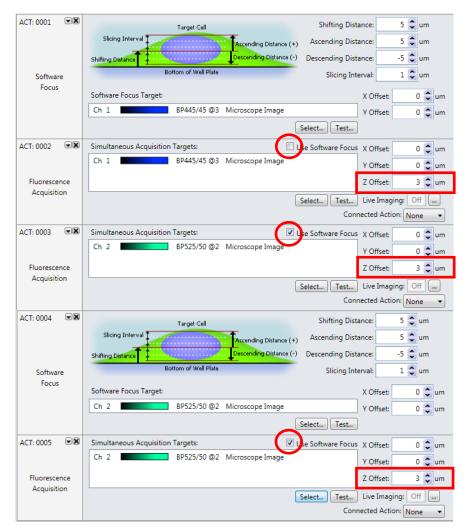


6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	g Wel	I Plate Scan Setti	ing Action List
Time Line	Name:	Time Line 1	
F	Period:	0 days 0 :	0:24
Start	t Time:	0 days 0 :	0 : 0
Start	t nine.	o days o .	
End	d Time:	0 days 0 :	0 : 24
Interva	l Time:	0 days 0 :	0:0
Expected Action	Time	0 days 0 :	0 : 24 Estimate Action Time
Expected Action	r mine.		
		Force to Chan	nge Expected Action Time
		NOTE: Time laps	se measurement fails if actual action time exceeds expected action time.



Images are acquired based on the last software focus setting if multiple software focus settings are set.



- ACT0001: Get the software-focused position scanned in the range by Ch1. ACT0002: Acquire the Ch1 image at the Z position where 3um are added to the auto-focused position.
- ACT0003: Acquire the Ch2 image at the Z position where 3um are added to the software-focused position by Ch1.
- ACT0004: Get the software-focused position scanned in the range by Ch2.
- ACT0005: Acquire the Ch2 image at the Z position where 3um are added to the software-focused position by Ch2.

3D Imaging from the Software Focus Plane

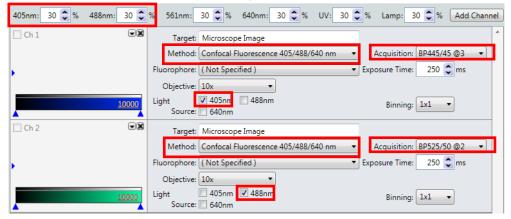
3D imaging is performed at two wavelengths (405 nm, 488 nm) by using the software focus plane as a reference.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%)
 - Ch2 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP525/50 Light Source: 488nm (30%)



4) Set the items on the Action List tab. (Refer to 5.7)

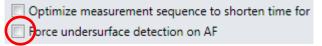
Software focus is applied to Ch1. Images are captured on the software focus plane for Ch1, and 3D imaging is performed for Ch2. Select the "Use Software Focus to Determine Shifting Distance" check box.

ACT: 0001	Target Cell Shifting Distance: 5 um
	Slicing Interval Ascending Distance (+) Ascending Distance: 5 Um Shifting Distance (-) Descending Distance: -5 Um
Software	Bottom of Well Plate Slicing Interval: 1
Focus	Software Focus Target: X Enter "0" if images
	Ch 1 BP445/45 Microscope Image YC are captured on the
ACT: 0002	Simultaneous Acquisition Targets: Use Software Focus X a Software focus plane.
Fluorescence Acquisition	Ch 1 BP445/45 Microscope Image Y Offset: 0 0 m Z Offset: 0 0 um
requisition	Select Test Live Imaging: Off
ACT: 0003	✓ Use oftware Focus to Determine Shifting Distance Target Cell
	Slicing Interval Ascending Distance (+) Shifting Distance (-) Shifting Distance (-) Descending Distance (-) Descending Distance (-)
3D	Bottom of Well Plate Slicing Interval: 1 🕽 um
Fluorescence Acquisition	Simultaneous Acquisition Targets: X Offset: 0 🗘 um
	Ch 2 BP525/50 Microscope Image Y Offset: 0 um
	Image Processing: Select Test

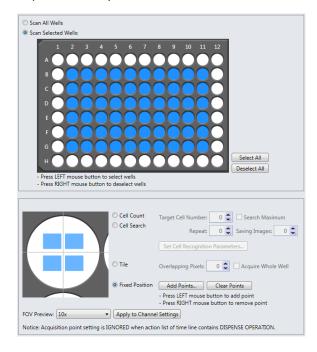
Select an output method for Z images from "Image Processing." (Refer to 5.1 and 6.2.)

Image Processing:	
	None 🔻
	None
	Maximum
	Minimum
	Average
	Sum

Uncheck "Force undersurface detection on AF".



5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)



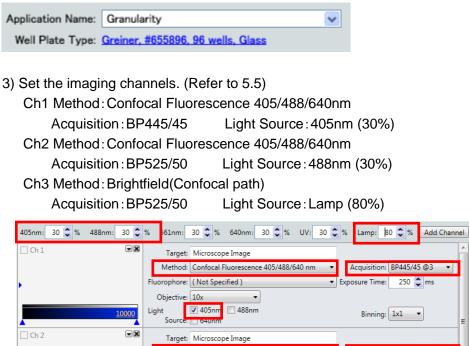
6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting W	/ell Plate Scan Setting Action List
Time Line Nam	e: Time Line 1
Perio	d: 0 days 0 : 0 : 24
Start Tim	e: 0 days 0 : 0 : 0
End Tim	e: 0 days 0 : 0 : 24
Interval Tim	e: 0 days 0 : 0 : 0
Expected Action Tim	e: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.3. Confocal Imaging and Bright-field Imaging of the Same View Field

Confocal imaging is performed at two wavelengths (405 nm, 488 nm) and then imaging of the same view field is performed in the bright-field mode.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



Ch 1	Target: Microscope Image	A
	Method: Confocal Fluorescence 405/488/640 nm	▼ Acquisition: BP445/45 @3 ▼
•	Fluorophore: (Not Specified)	 Exposure Time: 250 Cms
	Objective: 10x 🔹	
10000	Light 🛛 405nm 🔲 488nm Source: 🔄 640nm	Binning: 1x1 -
Ch 2	Target: Microscope Image	
	Method: Confocal Fluorescence 405/488/640 nm	▼ Acquisition: BP525/50 @2 ▼
•	Fluorophore: (Not Specified)	 Exposure Time: 250 ms
	Objective: 10x	
10000	Light 405nm 488nm Source: 640nm	Binning: 1x1
Ch 3	Target: Microscope Image	
	Method: Brightfield(confocal path)	▼ Acquisition: BP445/45 @3 ▼
•		Exposure Time: 250 🗢 ms
	Objective: 10x 🔹	
10000	Light Sourc <mark>: U Lamp</mark>	Binning: 1x1

4) Set the items on the Action List tab. (Refer to 5.7)

Set Ch1 and Ch2 for the Fluorescence Acquisition tasks, respectively. Set Ch3 for the Bright-field/Phase-contrast Acquisition task.

ACT: 0001	•*	Simultaneous Acquisition Targets:	Use Software Focus 🗴 Offset: 0 🗘 um
		Ch 1 BP445/45 @3 Microscope Ima	age Y Offset: 0 🗘 um
Fluoresce	Fluorescence		Z Offset: 0 🗘 um
Acquisit			
			Select Test Live Imaging: Off
			Connected Action: None 🔻
ACT: 0002	×X	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🗘 um
		Ch 2 BP525/50 @2 Microscope Ima	age Y Offset: 0 🗢 um
Fluoresce	ence		Z Offset: 0 🗢 um
Acquisit	tion		Select Test Live Imaging: Off
			Connected Action: None
	T		
ACT: 0003		Acquisition Target:	X Offset: 0 🚭 um
Bright-fie	eld /	Ch 3 BP445/45 @3 Microscope Ima	age Y Offset: 0 😂 um
Phase-cor			Z Offset: 0 🗢 um
Acquisit	tion		Select Test Live Imaging: Off
🔽 Optimize	e measur	ement sequence to shorten time for measurement	
Che	eck h	nere to acquire hole well plate by	v the confocal fluorescence an

Check here to acquire hole well plate by the confocal fluorescence and then by bright field.

If the checkbox is unchecked, each image data is acquired at the same timing in the case that there are the differences of optical method and objective lens settings in the action list with switching optics..

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line N	ame: Time Line 1
Pe	riod: 0 days 0 : 0 : 24
Start T	ime: 0 days 0 : 0 : 0
Start	
End T	ime: 0 days 0 : 0 : 24
Interval T	ïme: 0 days 0 : 0 : 0
Expected Action T	ime: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.4. Imaging by Simultaneous Laser Emissions at Multi

Wavelengths

Images are acquired by emitting laser beams of multi wavelengths simultaneously. (Multi-camera model only)

Confocal Imaging by Simultaneous Laser Emissions at 3 Wavelengths

Confocal imaging is performed by emitting laser beams of three wavelengths (405 nm, 488 nm and 640 nm) simultaneously.

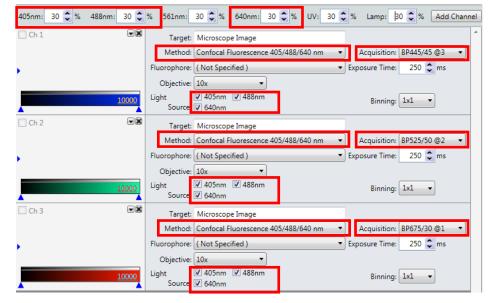
1) Open the measurement setting file edit screen. (Refer to 5.2)

2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm
 - Acquisition: BP445/45 Light Source: 405nm, 488nm, 640nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm
 - Acquisition: BP525/50 Light Source: 405nm, 488nm, 640nm (30%) Ch3 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP675/30 Light Source: 405nm, 488nm, 640nm (30%)





If performing crosstalk correction by Image Correction Software, select used fluorophore.

Fluorophore:	MitoTracker Deep Red 🔹
Objective:	(Not Specified) Hoechist33342
Light	mKusabira Orange2
Source:	Azami Green
	Alexa Fluor 488
	MitoTracker Deep Red
	DAPI
	FusionRed_spectra

Set the items on the Action List tab. (Refer to 5.7)
 Specify three wavelengths for one Fluorescence Acquisition task.

ACT: 0001	Simultaneous Acquisition Targets:	🔽 Use Software Focus	X Offset: 0 🗘 um
	Ch 1 BP445/45 @3	Microscope Image	Y Offset: 0 🗘 um
	Ch 2 BP525/50 @2	Microscope Image	
Fluorescence	Ch 3 BP675/30 @1	Microscope Image	Z Offset: 0 🗘 um
Acquisition		Select Test	Live Imaging: Off
		Conr	ected Action: None 🔻

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Nan	ne: Time Line 1
Perio	od: 0 days 0 : 0 : 24
Start Tin	ne: 0 days 0 : 0 : 0
End Tin	ne: 0 days 0 : 0 : 24
Interval Tin	ne: 0 days 0 : 0 : 0
Expected Action Tin	ne: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

3D Imaging by Simultaneous Laser Emissions at 3 Wavelengths

3D imaging is performed by emitting laser beams of three wavelengths (405 nm, 488 nm, 640 nm) simultaneously.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

Application Name:	Granularity	¥	
Well Plate Type:	Greiner, #655896, 96 wells, Glass		

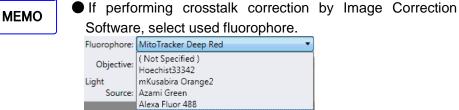
- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP445/45 Light Source: 405nm, 488nm, 640nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm

Light Source: 405nm, 488nm, 640nm (30%) Acquisition: BP525/50 Ch3 Method: Confocal Fluorescence 405/488/640nm

Light Source: 405nm, 488nm, 640nm (30%)





MitoTracker Dee DAPI

FusionRed_spectra

4) Set the items on the Action List tab. (Refer to 5.7)

Specify three wavelengths for one 3D Fluorescence Acquisition task. Unselect the "Use Software Focus to Determine Shifting Distance" check box.

ACT: 0001 💌 🕱	Use Software Focus to Determine Shifting Distance	Shifting Distance: 5 🍨 um
	Target Cell Slicing Interval Ascending Distance (+)	Ascending Distance: 5 🔹 um
	Shifting Distance (+) Descending Distance (-)	Descending Distance: -5 拿 um
3D	Bottom of Well Plate	Slicing Interval: 1 🤤 um
Fluorescence Acquisition	Simultaneous Acquisition Targets:	X Offset: 0 🗘 u
	Ch 1 BP445/45 Microscope Image	A
	Ch 2 BP525/50 Microscope Image	Y Offset: 0 🗘 u
	Ch 3 BP675/30 Microscope Image	 Image Processing:
	Select	

Select an output method for Z images from "Image Processing." (Refer to 5.1 and 6.2.)

Image Processing:		
	None 🔻	
	None	
	Maximum	
	Minimum	
	Average	
	Sum	

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3.)

Time Janco Sotting	Well Plate Scan Setting Action List
Time-tapse setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Peri	iod: 0 days 0 : 0 : 24
Start Tir	me: 0 days 0 : 0 : 0
End Tir	me: 0 days 0 : 0 : 24
T	
Interval Tir	me: 0 days 0 : 0 : 0
Expected Action Tir	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.5. High-speed Time-lapse Imaging per Well

In Case without Performing Autofocus during Imaging



In this case, perform autofocus before the first image acquisition. Focus may shift gradually by thermal expansion of machine.

High-speed time-lapse imaging is performed with a laser beam of 488 nm in wavelength at an imaging interval of 400 ms and imaging time of 30 seconds per well, with dispensing performed five seconds after the start of imaging.

1) Open the measurement setting file edit screen. (Refer to 5.2)

2) Enter the application name. (Refer to 5.4)



3) Set the imaging channels. (Refer to 5.5)

Ch1 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP525/50 Light Source: 488nm (30%) Binning: 2



Since a very large amount of image data will be acquired in high-speed time-lapse imaging, the binning setting more than "2x2" is recommended.

405nm: 30 🗢 % 488nm: 30 🗢	% 561nm: 30 🗘 % 640nm: 30 🗘 % UV: 3	30 🗘 % Lamp: 30 🗘 % Add Channel
Ch 1	Target: Microscope Image	^
	Method: Confocal Fluorescence 405/488/640 nm	▼ Acquisition: BP525/50 @2 ▼
•	Fluorophore: (Not Specified)	 Exposure Time: 100 🗢 ms
	Objective: 10x 🔻	1
<u>10000</u>	Light 0405nm 488nm Source: 640nm	Binning: 2x2 -
^	Source. 🔄 640nm	1x1
		2x2 3x3
		4x4

4) Set the items on the Action List tab. (Refer to 5.7)

To perform fluorescence imaging, set "Fluorescence Acquisition" task. To perform bright field/phase contrast imaging, set a "Bright-field/ Phase-contrast Acquisition" task.

ACT: 0001	Simultaneous Acquisition Targets:	Use Software Focus	X Offset: 0 🗘 um
	Ch 1 BP525/50 @2	Microscope Image	Y Offset: 0 🗘 um
Fluorescence			Z Offset: 0 🗘 um
Acquisition		Select Test	Live Imaging: Off 🛄
		Conr	nected Action: None 🔻

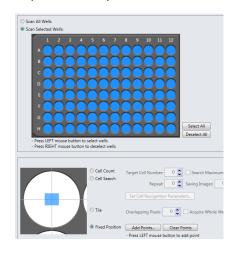
Uncheck "Force undersurface detection on AF".

Optimize measurement sequence to shorten time for brce undersurface detection on AF

5) Set high-speed time-lapse imaging. Uncheck "Performing AF during live imaging". (Refer to 5.8)

🚡 S	et Live Imaging Pa	arameters		X
0	No Live Imaging			
٥ ر	Use Live Imaging:			
	Period:	30000 🗘 ms		
	Interval:	400 🗘 ms		
		Dispense Setting Name	Timing	Add
		Dispensing	5000 ms	Change
	Dispense Setting:			Delete
				Liquid Volume
	(Perform AF during live imaging		
	Test Interval Value]		OK Cancel

6) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)





- •When images are captured by high-speed time-lapse imaging, the view field covers only one point at the center of the well.
- Select same acquisition points to correspond to the well-plate map specified by dispensing setting file (refer to 5.12) for Assay plate.

7) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3.)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Nar	ne: Time Line 1
Peri	od: 0 days 0 : 0 : 24
Start Tir	ne: 0 days 0 : 0 : 0
End Tir	ne: 0 days 0 : 0 : 24
Interval Tir	ne: 0 days 0 : 0 : 0
Expected Action Tir	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

8) Save the measurement setting file. (Refer to 5.11)

In Case with Performing Autofocus during Imaging

High-speed time-lapse imaging is performed with a laser beam of 488 nm in wavelength at an imaging interval of 10 seconds and imaging time of 5 minutes per well, with dispensing performed 20 seconds after the start of imaging.

1) Open the measurement setting file edit screen. (Refer to 5.2)

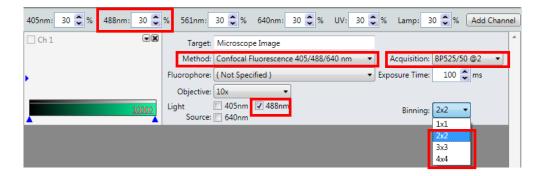
2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP525/50 Light Source: 488nm (30%) Binning: 2



Since a very large amount of image data will be acquired in high-speed time-lapse imaging, the binning setting more than "2x2" is recommended.



4) Set the items on the Action List tab. (Refer to 5.7)

To perform fluorescence imaging, set "Fluorescence Acquisition" task. To perform bright field/phase contrast imaging, set a "Bright-field/ Phase-contrast Acquisition" task.

Software focus is applied to Ch1. Images are captured for Ch1 on the software focus plane. Check "Use Software Focus" and set "0" under "Z Offset" for "Fluorescence Acquisition."

Click "Live Imaging"

ACT: 0001		r.	
ACT: 0001	Target Cell	Shifting Distance:	5 🔷 um
	Slicing Interval Ascending Distance (+)	Ascending Distance:	5 🗘 um
	Shifting Distance Descending Distance (-)	Descending Distance:	-5 🗘 um
Software	Bottom of Well Plate	Slicing Interval:	1 🗘 um
Focus	Software Focus Target:	X Off	iset: 0 😂 un
	Ch 1 BP525/50 @2 Microscope Image	Y Off	iset: 0 🗘 un
		elect Test	
ACT: 0002 🔍	Simultaneous Acquisition Targets:	se Software Focus X Off	iset: 0 😂 un
	Ch 1 BP525/50 @2 Microscope Image	Y Off	iset: 0 🗘 un
Fluorescence		Z Off	íset: 0 🗘 un
Acquisition		Select Test Live I	imaging: Off 🛄
		Connected	Action: None 🔻

Check "Force undersurface detection on AF".

Optimize measurement sequence to shorten time for orce undersurface detection on AF

5) Set high-speed time-lapse imaging. Check "Performing AF during live imaging". (Refer to 5.8)

📜 S	et Live Imaging Pa	arameters		
0	No Live Imaging			
0 (Jse Live Imaging:			
	Period:	300000 🗢 ms		
	Interval:	10000 🗘 ms		
		Dispense Setting Name	Timing	Add
	Dispense Setting:	Dispensing	20000 ms	Change
	Dispense Setting:			Delete
	(Perform AF during live imaging		Liquid Volume
	Test Interval Value			OK Cancel

6) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)

Scan Selected Wells:	
1 2 3 4 5 A 0 0 0 0 0 0 C 0 0 0 0 0 0 C 0 0 0 0 0 0 F 0 0	
	Cell Count Target Cell Number: 0 C Search Maximum Cell Search Repeat: 0 C Saving Images: 0
	Cell Search Repeat 0 Saving Images: Cell Recognition Parameters

MEMO	•When images are captured by high-speed time-lapse
	imaging, the view field covers only one point at the
	center of the well.

- Select same acquisition points to correspond to the well-plate map specified by dispensing setting file (refer to 5.12) for Assay plate.
- 7) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3.)

Time-lapse Setting We	Il Plate Scan Setting Action List
Time Line Name	Time Line 1
Period	
Start Time	0 days 0 : 0 : 0
End Time	: 0 days 0 : 0 : 24
Interval Times	0 days 0 : 0 : 0
Expected Action Times	0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.6. Time-lapse Imaging per Plate without Dispensing

Time-lapse imaging is performed for one assay plate at 15-minute intervals over a period of 50 minutes.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)
 Application Name: Granularity
 Well Plate Type: Greiner, #655896, 96 wells, Glass
- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%)
 - Ch2 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP525/50 Light Source: 488nm (30%)

405nm: 30 🗘 % 488nm: 30 🗘	% 561nm: 30 🗢 % 640nm: 30 🗢 % UV: 30 🗘	% Lamp: 30 🗢 % Add Channel
Ch 1	Target: Microscope Image	
	Method: Confocal Fluorescence 405/488/640 nm 🔹	Acquisition: BP445/45 @3 🔹
•	Fluorophore: (Not Specified) 🔹	Exposure Time: 250 🖨 ms
10000	Objective: 10x Light V 405nn 488nm Source: 640nm	Binning: 1x1 💌
Ch 2	Target: Microscope Image	
	Method: Confocal Fluorescence 405/488/640 nm 🔹	Acquisition: BP525/50 @2 🔻
•	Fluorophore: (Not Specified)	Exposure Time: 250 🗘 ms
10000	Objective: 10x Light 0405nm 0488nm Source: 640nm	Binning: 1x1 💌

4) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001	Simultaneous Acquisition Targets: 🔲 Use Software Focus X Offset: 0 🗘 um
	Ch 1 BP445/45 @3 Microscope Image Y Offset: 0 🗘 um
Fluorescence Acquisition	Z Offset: 0 🗘 um
Acquisition	Select Test Live Imaging: Off
	Connected Action: None
ACT: 0002	Simultaneous Acquisition Targets: 🔲 Use Software Focus X Offset: 0 🗘 um
	Ch 2 BP525/50 @2 Microscope Image Y Offset: 0 🗘 um
Fluorescence	Z Offset: 0 🗘 um
Acquisition	Select Test Live Imaging: Off
	Connected Action: None

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6.)

6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
	T 11 4
Time Line Na	me: Time Line 1
	at 0 to 0 a 0 a
Per	iod: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Expected Action Ti	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7) Enter the values of "Interval Time," "Start Time" and "Period." (Refer to 5.3) (Set the interval time to 15 minutes, the start time to 0 minute and the period to 50 minutes.)

Time-lapse	0:00:00 0:05:00	0:10:00 0:15:00	0:20:00 0:25:00	0.30-00	0:35:00 0:40	00 0:45:00	0:50:00
Fime Line 1				_			
							>
min	Whole				Add	Delete	Check
Time-lapse Setting	Well Plate Scan Setting	Action List					
Time Line Nam	ne: Time Line 1						
Perio	od: 0 days 0:50:	0					
Start Tin	ne: Odays 0:0:	0					
End Tin	ne: 0 days 0 : 50 :	0					
	ne: 0 days 0 : 50 :	0					
		_					

7.7. Time-lapse Imaging per Plate with Dispensing

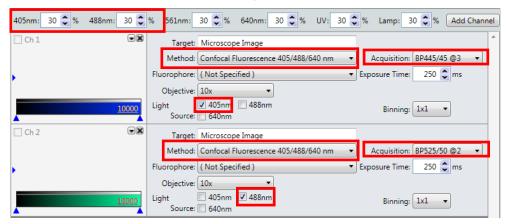
One assay plate is measured once, and 15 minutes thereafter the dispensing is performed from one source plate. After 40 minutes, time-lapse imaging is performed for the assay plate at 15-minute intervals over a period of 50 minutes.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm

Acquisition: BP525/50 Light Source: 488nm (30%)



4) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🗘 um
	Ch 1 BP445/45 @3 Microsco	pe Image Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 📚 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None
ACT: 0002	Simultaneous Acquisition Targets:	🗌 Use Software Focus X Offset: 0 🚭 um
	Ch 2 BP525/50 @2 Microsco	pe Image Y Offset: 0 🗘 um
Fluorescence Acquisition		Z Offset: 0 😂 um
		Select Test Live Imaging: Off
		Connected Action: None

5) Set the imaging wells and view field on the well Plate Scan Setting tab. (Refer to 5.6.)

6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting V	Vell Plate Scan Setting Action List
Time Line Nam	e: Time Line 1
Perio	d: 0 days 0 : 0 : 24
Start Tim	e: 0 days 0 : 0 : 0
End Tim	e: 0 days 0 : 0 : 24
Interval Tim	e: 0 days 0 : 0 : 0
Expected Action Tim	e: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7) Create a dispensing time line. Click "Add" and create a new time line. (Refer to 5.3)

Time-lapse	0:00:00	0:00:30	0:01:00	0:01:30	0:02:00	0:02:30	^
Time Line 1							
Time Line 2							_
							\sim
<							>
1 min	Whole				Add	Delete	Check

8) Set the items on the Action List tab. (Refer to 5.7) Refer to 7.12 for example of dispensing setting

ACT: 0001	Dispense Setting Name:	Dispensing test	
Dispense Operation		Edit Dispense Setting	



 In case of setting "Dispense Operation", assign only "Dispense Operation" in Action List. If other actions like "Fluorescence Acquisition" should be set, assign in other timeline. 9) Set the wells to dispense. (Refer to 5.6)

Solution is dispensed once for each well. The Acquisition Points settings are not reflected.

Scan All Wells Scan Selected Wells: 1 2 3 4 5 6 7 A	8 9 10 11 12
F G G G G G G G G G G G G G G G G G G G	Select All Deselect All
- Press RIGHT mouse button to deselect wells	
Cell Count Cell Search	Target Cell Number: 0 💭 🗆 Search Maximum Repeat: 0 💭 Saving Images: 0
Tile Fixed Position	Set Lell Recognition Parameters Overlapping Pixels: 0 Add Points Clear Points
	Press LEFT mouse button to add point
A	
	elect same acqui

Select same acquisition points to correspond to the well-plate map specified by dispensing setting file (refer to 5.12) for Assay plate.

10) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting We	I Plate Scan Setting Action List
Time Line Name:	Time Line 2
Period:	0 days 0 : 0 : 27
Start Time:	0 days 0 : 0 : 0
End Time:	0 days 0 : 0 : 27
Interval Time:	0 days 0 : 0 : 0
Expected Action Time:	0 days 0 : 0 : 2 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

11) Enter the value of "Start Time" for Time Line 2. (Refer to 5.3)

Time-lapse	0:00:00	0:03:00	0:06:00	0:09:00	0:12:00	0:15:00	0:18:00	0:21:00	^
Time Line 1									
Time Line 2									
Time-lapse Settir	well P	late Scan S	Setting	Action List	1				
Time Line	Name: Time	Line 2							
		CINC 2							
F	Period: 0 d	days 0:	6:22						
Start	Time: 0 d	days 0:	15: 0						
End	Time: 0 a	days 0:	21:22						
Interval	Time: 0 d	days 0:	0:0						
						_			
Expected Action	Time: 0 a	days 0:	6 : 22	Estimate /	Action Time	•			

12) Create a new time-lapse time line. Click "Add" and create a new time line. (Refer to 5.3)

ime-lapse	0:00:00	0:03:00 0:06:	0:09:00	0:12:00	0:15:00 0:1	8:00 0:21	.00
ime Line 1							-
ime Line 2					_		
ime Line 3							
	_						
						· · · · · ·	2
	Whole				Add	Delete	Check

13) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🔷 um
	Ch 1 BP445/45 @3 Microso	ope Image Y Offset: 0 🗘 um
Fluorescence Acquisition		Z Offset: 0 📚 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None
ACT: 0002	Simultaneous Acquisition Targets:	🔲 Use Software Focus X Offset: 0 🗢 um
	Ch 2 BP525/50 @2 Microso	ope Image Y Offset: 0 📚 um
Fluorescence		Z Offset: 0 📚 um
Acquisition		
		Select Test Live Imaging: Off

- 14) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 15) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting Wel	I Plate Scan Setting Action List
Time Line Name:	Time Line 3
Period:	0 days 0 : 0 : 27
Start Time:	0 days 0 : 0 : 0
End Time:	0 days 0 : 0 : 27
Interval Time:	0 days 0 : 0 : 0
Expected Action Time:	0 days 0 : 0 27 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

16) Enter the values of "Start Time" "Interval Time" and "Period" for Time Line 3. (Refer to 5.3)

Time-lapse	0:00:00	0:30:00	1:00:00	 1:30:00
Time Line 1				
Time Line 2				
Time Line 3				_
		~		
Time-lapse Setting	Well Plate S	can Setting Action List	:	
Time Line Nam	e: Time Line	3	1	
		•	_	
Perio	od: 0 days	0:50:0		
Start Tim	ne: 0 days	0:40:0		
End Tim	0 days	1:30:0		
End IIm	e: 0 days	1: 30: 0		
Interval Tim	e: 0 days	0:15:0		
Interval Int	o days	0.10.0		
Expected Action Tim	e: 0 days	0 : 2 : 54 Estimat	e Action Time	

7.8. Capturing Images According to the View Field Observed in the

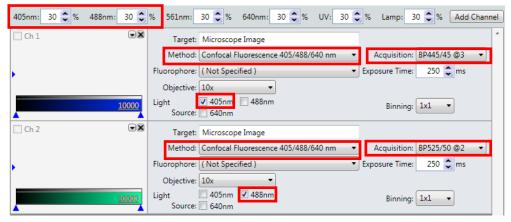
Test Preview

Capture images of the wells observed in the test preview according to the corresponding view field.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

Application Name:				
Well Plate Type:	Greiner, #655896, 96 wells, Glass			

- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm
 - Acquisition: BP525/50 Light Source: 488nm (30%)



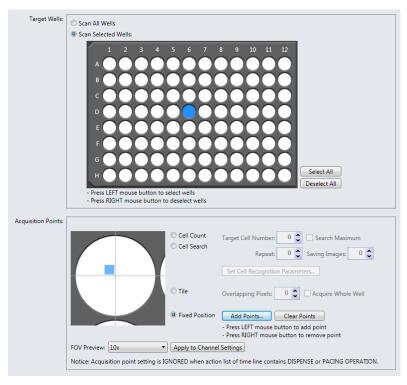
4) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001	Simultaneous Acquisition Targets:	🗆 Use Software Focus 🛛 O 🗲 um
	Ch 1 BP445/45 @3 Micros	scope Image Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 📚 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None
ACT: 0002	Simultaneous Acquisition Targets:	🔲 Use Software Focus 🛛 X Offset: 0 🗢 um
	Ch 2 BP525/50 @2 Micros	scope Image Y Offset: 0 🗢 um
Fluorescence		Z Offset: 0 📚 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None

- 5) Display a test preview. (Refer to 5.9 and 6.2)
 - Before opening the test preview screen, select the "Synchronize Well Plate Scan Setting" check box. Specify the imaging wells and view field.



6) Confirm on the Well Plate Scan Setting tab that the wells and view field specified in the test preview are set. You can also add imaging wells and view fields. (Refer to 5.6)



7) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting V	Vell Plate Scan Setting Action List
Time Line Nam	e: Time Line 1
Perio	d: 0 days 0 : 0 : 24
Start Tim	e: 0 days 0 : 0 : 0
End Tim	e: 0 days 0 : 0 : 24
Interval Tim	ne: 0 days 0 : 0 : 0
Expected Action Tim	e: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

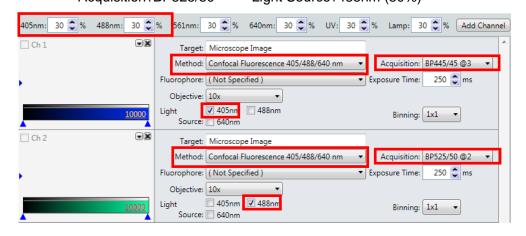
7.9. Imaging by Cell Count

Images are captured repeatedly in the same well while moving through the acquisition points until the specified cell count is reached. Once the number of repetitions reaches the specified value, imaging is stopped and the system moves to the next well.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP525/50 Light Source: 488nm (30%)



4) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001	Simultaneous Acquisition Targets:	🔲 Use Software Focus 🛛 X Offset: 🛛 0 🗢 um
	Ch 1 BP445/45 @3	Microscope Image Y Offset: 0 🗘 um
Fluorescence Acquisition		Z Offset: 0 🗘 um
		Select Test Live Imaging: Off
		Connected Action: None
ACT: 0002	Simultaneous Acquisition Targets:	🔲 Use Software Focus 🛛 X Offset: 🛛 0 🗢 um
	Ch 2 BP525/50 @2	Microscope Image Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 😂 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)

6) Select "Cell Count" on the Acquisition Points screen. (Refer to 5.6)

	Cell Count	Target Cell Number: Repeat:	100 Search Maximum 100 Saving Images: 0
	/	Set Cell Recognition	on Parameters
	OTile	Overlapping Pixels:	0
	Fixed Position	Add Points	Clear Points
		- Press LEFT mouse - Press RIGHT mous	button to add point e button to remove point
FOV Preview: 10x	Apply to Channe	Settings	

7) Set the cell recognition algorithm. (Refer to 5.10)

🐻 Set Ce	Il Recognition Parameters 🛛 🛛 🗙
Algorithm:	Connection
Threshold:	687 🗢
	Remove the fragments of the cell at the edge of the image
Measure:	Mean Cell Diameter: 12 🚭 um
	Minimum Scaling Factor: 0.5 📚 (6.0 um)
	Maximum Scaling Factor: 2.0 🗢 (24.0 um)
	OK Cancel

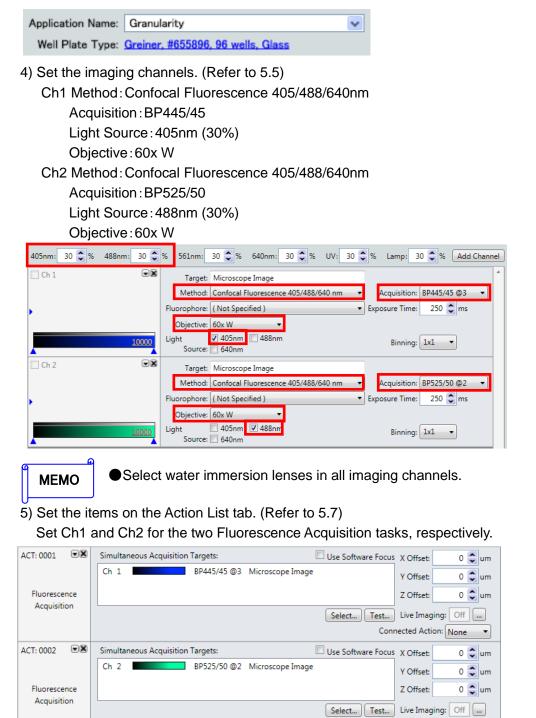
8) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting We	Il Plate Scan Setting Action List
Time Line Name	Time Line 1
Period	0 days 0 : 0 : 24
Start Time:	0 days 0 : 0 : 0
End Time:	0 days 0 : 0 : 24
Interval Time:	0 days 0 : 0 : 0
Expected Action Times	0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.10. Imaging by Water Immersion Lens

Confocal imaging is performed on the auto-focus plane at two wavelengths (405 nm, 488 nm) by water immersion lens. (Water immersion lens model only)

- 1) Supply the water to the water immersion lenses. (Refer to 5.14)
- 2) Open the measurement setting file edit screen. (Refer to 5.2)
- 3) Enter the application name. (Refer to 5.4)



Connected Action: None 🔹

- 6) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 7) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Nar	me: Time Line 1
Peri	od: 0 davs 0 : 0 : 24
Pen	0 days 0 : 0 : 24
Start Tir	me: 0 days 0 : 0 : 0
End Tir	me: 0 days 0 : 0 : 24
Interval Tir	me: 0 days 0 : 0 : 0
Expected Action Tir	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

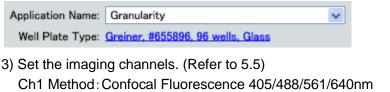
8) Save the measurement setting file. (Refer to 5.11)

•Wipe the water which attached to the bottom of the plate after the measurement by the water immersion lenses.

7.11. Imaging by 4-Laser Scanning (QUAD-DM and Dual filter model only)

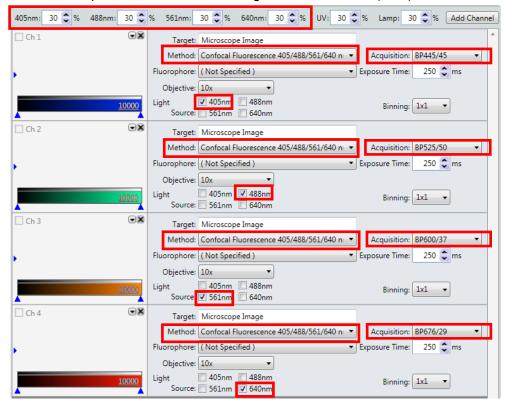
Confocal imaging is performed on the software focus plane at four wavelengths (405 nm, 488 nm, 561nm, 640nm).

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

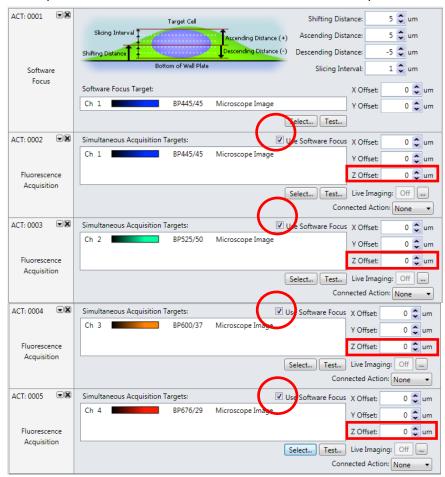


- Acquisition: BP447/50(Dual) Light Source: 405nm (30%) Ch2 Method: Confocal Fluorescence 405/488/561/640nm
- Acquisition : BP522/42(Dual) Light Source : 488nm (30%) Ch3 Method: Confocal Fluorescence 405/488/561/640nm
- Acquisition: BP600/37 Light Source: 561nm (30%) Ch4 Method: Confocal Fluorescence 405/488/561/640nm

Acquisition: BP676/29 Light Source: 640nm (30%)



4) Set the items on the Action List tab. (Refer to 5.7) Software focus is applied to Ch1. Images are captured for Ch1, Ch2, Ch3 and Ch4 on the software focus plane. To capture images on the software focus plane, set "0" under "Z Offset" for "Fluorescence Acquisition."



- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Peri	iod: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Expected Action Ti	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.
	Hore the appended the and a deal action time exceeds expected belon time

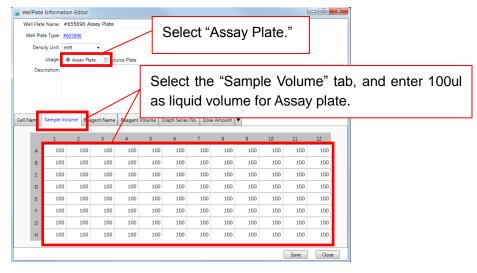
7.12. Example of Dispensing Setting

This section explains about commonly-used setting for well plate information files of Assay plate and Source plate (refer to 4.1), and dispensing setting file (refer to 5.12) to perform measurement using dispensing by 96-well tip rack.

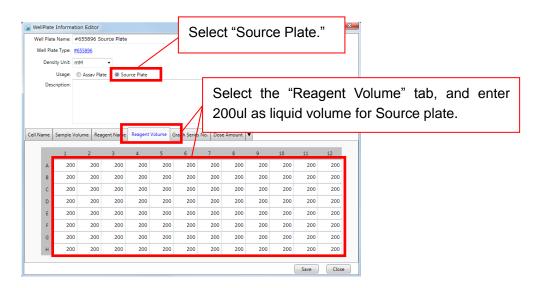
Setting #1 (To dispense below liquid surface of Assay plate)

Dispense 50ul to Assay plate from Source plate in which liquid amount of 200ul is added each well. Dispensing is performed from tip top at the Z position which moved to 1000um below from the liquid surface of Assay plate. This setting is the most common use.

1) Create each of wellplate information files for Assay plate and Source plate. (Refer to 4.1.)



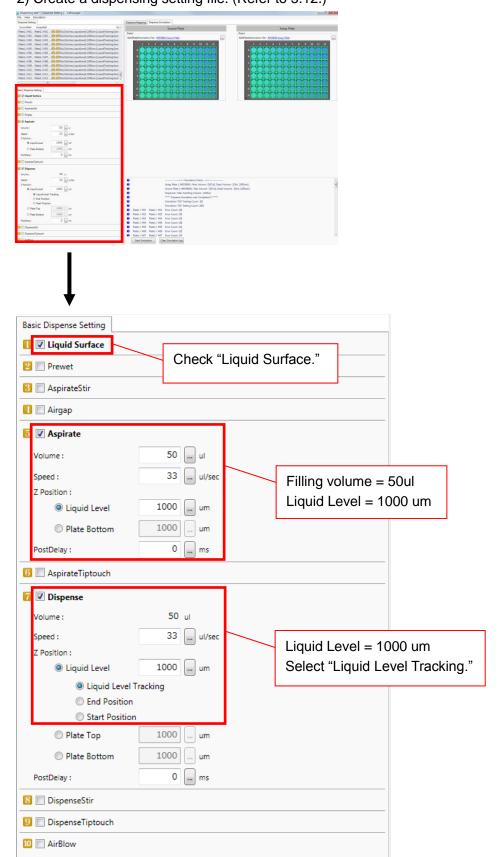
Setting for Assay plate



Setting for Source plate



Indication of reagent volume is 20% - 40% of well volume in Assay Plate and 50% - 80% of well volume in Source Plate.

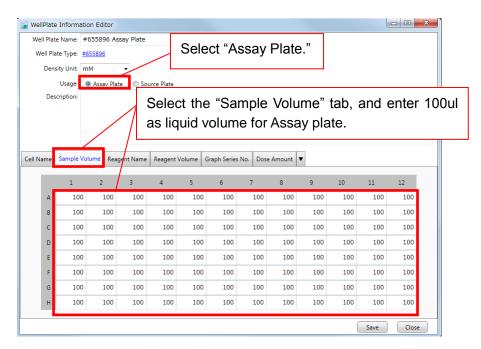


2) Create a dispensing setting file. (Refer to 5.12.)

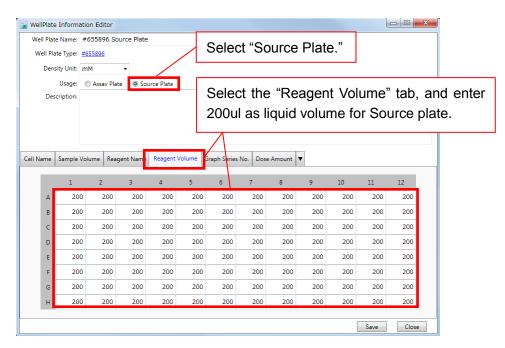
Setting #2 (To dispense above liquid surface of Assay plate)

Dispense 50ul to Assay plate from Source plate in which liquid amount of 200ul is added each well. Dispensing is performed from tip top at the Z position which moved to 2000um above from the liquid surface of Assay plate.

1) Create each of wellplate information files for Assay plate and Source plate. (Refer to 4.1.)

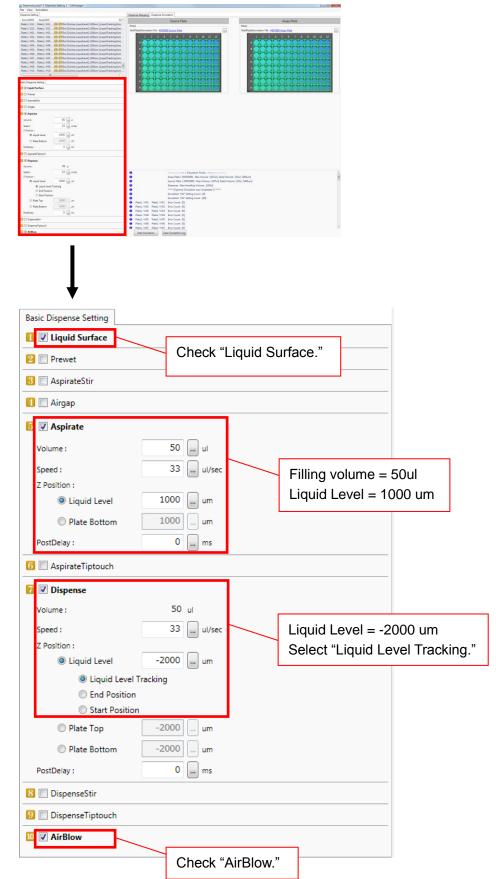


Setting for Assay plate



Setting for Source plate





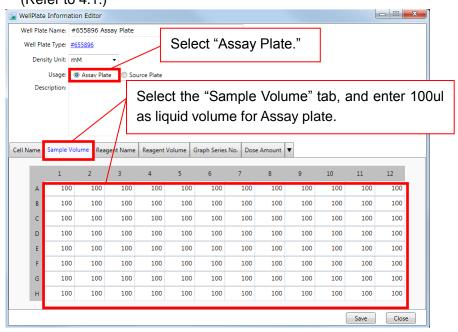
2) Create a dispensing setting file. (Refer to 5.12.)

Setting #3 (To use liquid volume of Source plate until around Dead Volume)

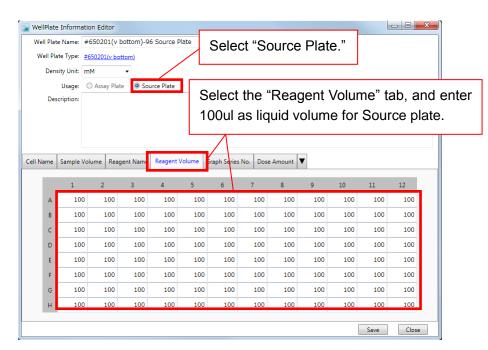
Dispense 80ul to Assay plate from V-bottom Source plate (Dead Volume: 8ul) in which liquid amount of 100ul is added each well.

Dead Volume indicates the minimum volume so as not to make contacts between the tip top and the bottom surface of Source plate.

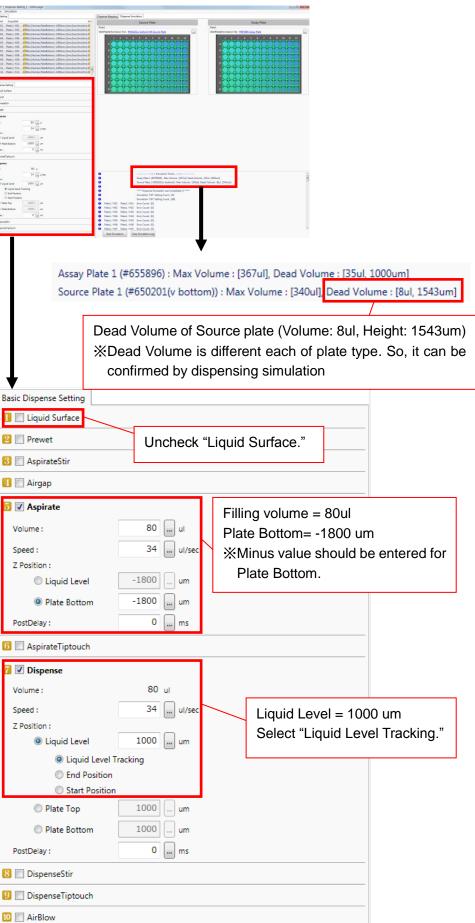
1) Create each of wellplate information files for Assay plate and Source plate. (Refer to 4.1.)



Setting for Assay plate



Setting for Source plate







- To use solution until around Dead Volume, it is recommended to use the plates which has V-shaped bottom.
- Refer to the following table to confirm Dead Volume of each plate type.

Indication for Dead Volume of each plate type

Well number/ Well shape	Well bottom shape	Source plate Dead Volume
96-well / Round-shaped	Flat	Almost 100ul
384-well / Square-shaped	Flat	Almost 25ul
96-well / Round-shaped	V-bottom /	Almost 10ul
90-weil / Round-snaped	Round-shaped	Aimost Tour
384-well / Square-shaped	V-bottom /	Almost 10ul
Squale-shaped	Round-shaped	Aimost Tour
384-well / Square-shaped	V-bottom /	Almost 10ul
Source Shaped	Square-shaped	Aimost Tour

7.13. High-Speed Time-Lapse Imaging with Multi Dispensing

Hitg-speed time-lapse imaging is performed with a laser beam of 488nm in wavelength at an imaging interval of 400ms and imaging time of 70s per well, with dispensing performed 5s after the start of imaging. Furthermore, another compound is dispensed 35s after the start of imaging in a same well.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

Application Name:	Granularity	~	
Well Plate Type:	Greiner, #655896, 96 wells, Glass		

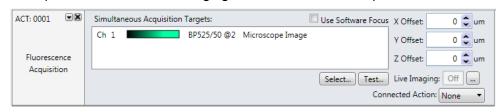
- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP525/50 Light Source: 488nm (30%) Binning: 2



●Since a very large amount of image data will be acquired in high-speed time-lapse imaging, the binning setting more than "2x2" is recommended.

405nm: 30 🗘 % 488nm: 30 🗘	% 561nm: 30 🗢 % 640nm: 30 🗢 % UV: 30	% Lamp: 30 % Add Channel
Ch 1	Target: Microscope Image	^
	Method: Confocal Fluorescence 405/488/640 nm	Acquisition: BP525/50 @2 🔹
•	Fluorophore: (Not Specified)	Exposure Time: 100 🗢 ms
16000	Objective: 10x ▼ Light 0405nm √ 488nm Source: 640nm	Binning: 2x2 •
		2x2 3x3 4x4

4) Set the items on the Action List tab. (Refer to 5.7) To perform fluorescence imaging, set "Fluorescence Acquisition" task.



5) Set high-speed time-lapse imaging. (Refer to 5.8)

Up to 3 dispensing setting files (refer to 5.12, 7.12) can be assigned.

🚡 Set Live	e Imaging Pa	arameters		X		
🔘 No Live	lo Live Imaging					
Ose Live	e Imaging:					
	Period:	70000 🗘 ms				
	Interval:	400 🗘 ms				
		Dispense Setting Name	e Timing	Add		
		First dispensing	5000 ms	Change		
Dispe	ense Setting:	Second dispensing	35000 ms	Delete		
				Liquid Volume		
		Perform AF during liv	/e imaging			
		 ר				
lest	nterval Value			OK Cancel		
	Plane1_(EDR Pil Plane1_(FDR Pil Pilex1_(FDR Pilex1_(FDR	Act (CM) BLDA: Unix capacities (CM) capacities (CM) is lipseful water (CM) Act (CM) BLDA: Unix capacities (CM) AC (CM) BLDA: Unix capacities	Pert	10 10 10		
Servers Servers Servers Servers PARES 155 Rends (201 Rends (201 PARES 156 Rends (201 Rends (201 Rends 156 Rends (201 Rends (201	set. Lagate et (2000 en) Lagathering Burn et al. Lagate et (2000 en) Lagathering Burn et (2000 en) Lagathering	Terrer Torrer James James Handler Barrer Handler Terr				
C Aspiratelle C Aspiratelle C Aspiratelle C Aspiratelle			Malle which come	ound in drippod must be		
0 🖸 Apprete Tytouch 8 💭 Dispense			-	ound is dripped must be		
Doprestite Doprestptsach Die Addise			set as same in all	dispensing setting files.		

The first dispensing is performed from upper left 24wells of Source Plate to center 24wells of Assay Plate 5s after starting imaging.

The second dispensing is performed from lower right 24wells of Source Plate to center 24wells of Assay Plate 35s after starting imaging.

An imaging interval is 400ms. Time-lapse imaging finishes 70s after staring imaging and move to next well.

6) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)

('	(CI	iO	0.0	יי

Target Wells:	C Scan All Wells	Vells:	7 8 9 10 11 12	
	G O			ct All
		mouse button to select wells IT mouse button to deselect w	Dese	
Acquisition Points		T mouse button to deselect w	efs	
Acquisition Points		T mouse button to deselect w	efs	eet All
Acquisition Points:		T mouse button to deselect w	ets Target Cell Number 0 0 0 5e Repett 0 0 0 5ein Set Cell Recognition Parameters.	eet All

- When images are captured by high-speed time-lapse imaging, the view field covers only one point at the center of the well.
- Select the same wells which is assigned in well-plate map of Assay Plate in dispensing setting file (refer to
- 7) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Peri	od: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Expected Action Ti	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.14. Tile Imaging

Image the whole/ partial region(s) of well as tile. Tiled image can be analyzed with displaying whole image. (Only Analysis Software supported model)

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%)
 - Ch2 Method: Confocal Fluorescence 405/488/640nm

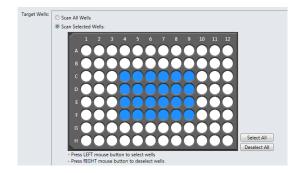
Acquisition: BP525/50 Light Source: 488nm (30%)

405nm: 30 🗢 % 488nm: 30 🜲	% 561nm: 30 🗘 % 640nm: 30 🗘 % UV: 30 🗘 % Lamp: 3	0 🗢 % 🛛 Add Channel
Ch 1	Target: Microscope Image	*
	Method: Confocal Fluorescence 405/488/640 nm	BP445/45 @3 🔻
•	Fluorophore: (Not Specified) Exposure Time:	250 🗘 ms
	Objective: 10x	
10000	Light 2405nm 2488nm Binning: Source: 640nm	1x1 •
Ch 2	Target: Microscope Image	
	Method: Confocal Fluorescence 405/488/640 nm	BP525/50 @2 🔻
•	Fluorophore: (Not Specified) Exposure Time:	250 🗘 ms
10000	Objective: 10x ▼ Light 405nm ▼ 488nm Binning: Source: ■ 640nm	1x1 •

4) Set the items on the Action List tab. (Refer to 5.7)

ACT: 0001 💌 🕱	Simultaneous Acquisition Targets:	🗌 Use Software Focus 🛛 X Offset: 👘 0 🗘 um
	Ch 1 BP445/45 @3	Microscope Image Y Offset: 0 😂 um
Fluorescence		Z Offset: 0 📚 um
Acquisition	<u>.</u>	Select Test Live Imaging: Off
		Connected Action: None 🔻
ACT: 0002	Simultaneous Acquisition Targets:	🔲 Use Software Focus 🛛 X Offset: 🛛 0 🗢 um
	Ch 2 BP525/50 @2	Microscope Image Y Offset: 0 🗘 um
Fluorescence		Z Offset: 0 🗘 um
Acquisition		Select Test Live Imaging: Off
		Connected Action: None 🔻

5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)



6) Select "Tile" or "Partial Tile" in Acquisition Points and assign the value of "Overlapping Pixels". (Refer to 5.6)

Please add one or more acquisition area		
Cell Count	Target Cell Number: 🛛 0 🤤 🗌 Search Maximum	
Cell Search	Repeat: 0 🗘 Saving Images: 0 🗘	
	Set Cell Recognition Parameters	
 Tile Partial Tile 	Overlapping Pixels: 0 🍨 🗆 Acquire Whole Well	
Fixed Position	Add Points Clear Points	
	 Press LEFT mouse button to add point Press RIGHT mouse button to remove point 	
FOV Preview: 20x Apply to Channel	Settings	
Notice: Acquisition point setting is IGNORED when action list of time line contains DISPENSE OPERATION.		

7) Click "Estimete Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting We	ell Plate Scan Setting Action List
Time Line Name	Time Line 1
Period	0 days 0 : 0 : 24
Start Time	0 days 0 : 0 : 0
End Time	0 days 0 : 0 : 24
Interval Time	0 days 0 : 0 : 0
Expected Action Time	: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

7.15. High-speed Time-lapse Imaging working with normal Imaging

After the one time imaging of DAPI stained nuclei by 405 nm laser, High-speed time-lapse imaging is performed by 488 nm laser at an imaging interval of 400 ms and imaging time of 30 seconds per well, with dispensing performed five seconds after the start of imaging.

High-speed Time-lapse Imaging only in the channel which is reactive to dispensing can reduce the amount of image data.

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/561nm

Acquisition: BP445/45	Light Source: 405nm (30%)	Binning: 2
Ch2 Method: Confocal Fluoresc	cence 405/488/561nm	

Acquisition: BP525/50 Light Source: 488nm (30%) Binning: 2



Since a very large amount of image data will be acquired in high-speed time-lapse imaging, the binning setting more than "2x2" is recommended.

Ch 1 Target: Microscope Image Method: Confocal Fluorescence 405/488/561 nm Acquisition: BP445/45 @3 Fluorophore: (Not Specified) Exposure Time: 100 \$ms
Fluorophore: (Not Specified) Exposure Time: 100 ms
Objective: 10x
10000 Light ✓ 405nm □ 488nm Binning: 2x2 ✓ Source: 561nm 561nm <td< th=""></td<>
Ch 2 Target: Microscope Image
Method: Confocal Fluorescence 405/488/561 nm 🔹 Acquisition: BP525/50 @2 🔹
Fluorophore: (Not Specified) Exposure Time: 100 🗘 ms
Objective: 10x
Light □ 405nm
3x3 4x4

4) Set the items on the Action List tab. (Refer to 5.7)

To perform fluorescence imaging, set "Fluorescence Acquisition" task.

Set Live Imaging: "On" only in ACT:0002, where Ch2 is set and set Connected Action: "ACT:02" in ACT:0001, where Ch1 is set. In this setting, imaging in Ch1 runs one time then High-speed time-lapse imaging in Ch2 starts consecutively. These images are linked as a set of data.

ACT: 0001	Simultaneous Acquisition Targets:	Use Software Focus	X Offset: 0 🗘 um
	Ch 1 BP445/45 @3	Microscope Image	Y Offset: 0 🗘 um
Fluorescence Acquisition			Z Offset: 0 🗘 um
Acquisition		Select Test	Live Imaging: Off
		Conr	ected Action ACT: 02 💌
ACT: 0002	Simultaneous Acquisition Targets:	Use Software Focus	X Offset: ACT: 02
	Ch 2 BP525/50 @2	Microscope Image	Y Offset: 0 🗘 um
Fluorescence			Z Offset: 0 🗘 um
Acquisition		Select Test	Live Imaging: On
		Conr	ected Action: None 🔻

• CV7000 Analysis Software can analyze these data.

5) Set high-speed time-lapse imaging. (Refer to 5.8)

MEMO

🍺 S	et Live Imaging Pa	arameters		X	
0	O No Live Imaging				
© ເ	Jse Live Imaging:				
	Period:	30000 🗘 ms			
	Interval:	400 🗘 ms			
		Dispense Setting Name	Timing	Add	
		Dispensing	5000 ms	Change	
	Dispense Setting:			Delete	
				Liquid Volume	
		Perform AF during live imaging			
	Test Interval Value OK Cancel				

7.16. DPC Imaging

In Case of DPC Imaging Only

- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)

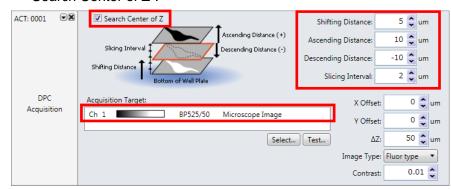


3) Set the imaging channels. (Refer to 5.5)

Ch1 Method: Digital	Phase Contrast	
Acquisition : BP	525/20 Light Source : Lamp (50%))
405nm: 30 🗢 % 445nm: 30 🗘	% 488nm: 30 🗢 % 561nm: 30 🔷 % UV: 30 🗢 % Lamp	50 🛟 Add Channel
Ch 1	Target: Microscope Image	*
	Method: Digital Phase Contrast	ion: BP525/50 🔻
•	Exposure Ti	me: 250 🛟 ms
	Objective: 10x 🔹	
10000	Light Binni Source Binni	ing: 1x1 •

4) Set the item on the Action List tab. (Refer to 5.7) DPC images are captured for Ch1.

In case of performing automatic DPC reference position search, check "Search Center of Z".



In case of without performing automatic DPC reference position search, uncheck "Search Center of Z".

ACT: 0001	Search Center of Z	Shiftin	g Distance:	5 🗘	um
	Ascending Distance (+) Slicing Interval	Ascendin	g Distance:	10 🗘	um
	Slicing Interval	Descendin	g Distance:	-10 🛟	um
	Bottom of Well Plate	Slici	ng Interval:	2 🌲	um
DPC Acquisition	Acquisition Target:		X Offset:		0 🛟 um
Acquisition	Ch 1 BP525/50 Microscope Image		Y Offset:		0 🛟 um
	Select.	Test	ΔZ:	5	0 🛟 um
			Image Type:	Fluor ty	pe 🔻
			Contrast:	(0.01 🗘

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting	Well Plate Scan Setting Action List
Time Line Na	me: Time Line 1
Peri	od: 0 days 0 : 0 : 24
Start Ti	me: 0 days 0 : 0 : 0
End Ti	me: 0 days 0 : 0 : 24
Interval Ti	me: 0 days 0 : 0 : 0
Expected Action Ti	me: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

Confocal Imaging and DPC Imaging of the Same View Field

Confocal imaging is performed at two wavelengths (405 nm, 488 nm) and then imaging of the same view field is performed in the DPC mode.

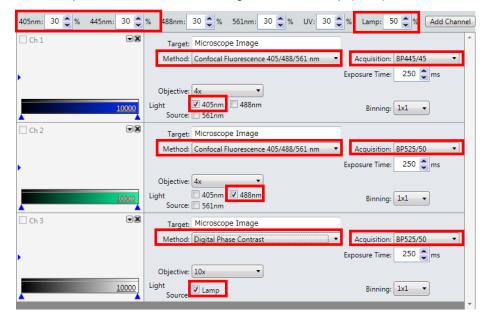
- 1) Open the measurement setting file edit screen. (Refer to 5.2)
- 2) Enter the application name. (Refer to 5.4)



- 3) Set the imaging channels. (Refer to 5.5)
 - Ch1 Method: Confocal Fluorescence 405/488/640nm Acquisition: BP445/45 Light Source: 405nm (30%) Ch2 Method: Confocal Fluorescence 405/488/640nm
 - Acquisition: BP525/50 Light Source: 488nm (30%)

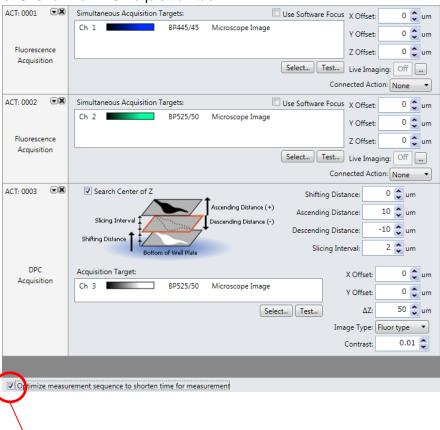
Ch3 Method: Digital Phase Contrast Acquisition: BP525/20 Ligh

Light Source: Lamp (50%)



4) Set the items on the Action List tab. (Refer to 5.7)

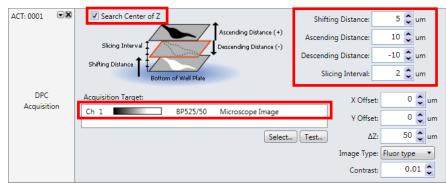
Set Ch1 and Ch2 for the Fluorescence Acquisition tasks, respectively. Set Ch3 for the DPC Acquisition task.



Check here to acquire hole well plate by the confocal fluorescence and then by bright field.

If the checkbox is unchecked, each image data is acquired at the same timing in the case that there are the differences of optical method and objective lens settings in the action list with switching optics..

In case of performing automatic DPC reference position search, check "Search Center of Z".



In case of without performing automatic DPC reference position search, uncheck "Search Center of Z".

ACT: 0001	Search Center of Z	Shifting Distance: 5 🗘 um
	Slicing Interval	Ascending Distance: 10 💭 um Descending Distance: -10 💭 um Slicing Interval: 2 💭 um
DPC Acquisition	Acquisition Target:	X Offset: 0 🗘 um Y Offset: 0 🗘 um
	Select	. Test ΔZ: 50 ℃ um Image Type: Fluor type ▼ Contrast: 0.01 ℃

- 5) Set the imaging wells and view field on the Well Plate Scan Setting tab. (Refer to 5.6)
- 6) Click "Estimate Action Time" on the Time-lapse Setting tab to check the measurement time. (Refer to 5.3)

Time-lapse Setting W	ell Plate Scan Setting Action List
Time Line Name	e: Time Line 1
Period	d: 0 days 0 : 0 : 24
Start Time	e: 0 days 0 : 0 : 0
End Time	0 days 0 : 0 : 24
End Time	: 0 days 0 : 0 : 24
Interval Time	e: 0 days 0 : 0 : 0
Expected Action Time	e: 0 days 0 : 0 : 24 Estimate Action Time
	Force to Change Expected Action Time
	NOTE: Time lapse measurement fails if actual action time exceeds expected action time.

8. Measurement

8.1. Setting Well Plates

●DO NOT set well plate on lifter when stage is inside of CV7000. Well plate or stage can be injured.



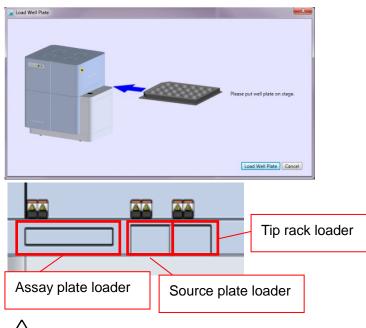
1) Set a well plate on the assay plate loader before displaying a measurement preview or starting measurement.

Click "Load Well Plate" on the Image Acquisition Test screen (refer to 5.9 and 6.2), or click "Start Measurement" on the Start Measurement screen (refer to 8.3 and 8.4).

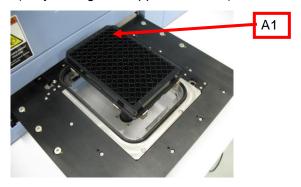
Tradice Time Line 1 / ACT: 0001 / Action Software Food
1 2 3 4 5 7 8 9 10 12 2 11003 4 4 5 6 7 8 9 5 9
E Province Mail Dista Prov Patrice
Protective at the data data from the data data data data data data data dat
Medium Image Processing: Slicing Interval: 1 um Image Processing:
Image: Second Set S/15 10000 Optimize Cell Recognition Parameter Optimize Cell Recognition Parameter Acquire Image Acquire Image

Start Measurement		×
Operator:	Yokogawa 🔻	
Measurement Title:	Measurement -	
Assay Plate:	AssayPlate_Greiner_#655896	
Measurement Setting:	Sample test1	
Assay Plate Information:	<default information="" plate=""></default>	Select
Image Correction Post Process	Dark & Shading Correction	Setting
	 Geometric Correction Channel Registration 	
	Crosstalk Correction	
	Post Process	
	🥥 Auto Analysis	
Turn off lasers when the measurement is finished		

 The stage moves to the loader exit. When the stage has fully moved to the specified position, the Load Well Plate screen opens. (Do not click "Load Well Plate.")

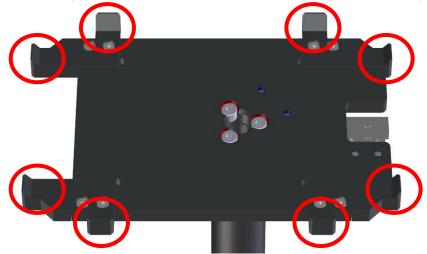


- DO NOT touch loader exit area while moving the loaders.
- 3) Set the assay plate on the stage. Set the plate so that the well "A1" on the well plate comes to the top left-hand corner of the stage.In case of observation by slide glass, please refer to 8.7 (Only slide glass supported model)

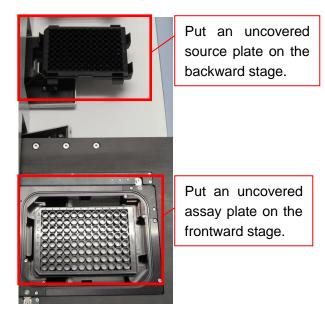




Set the assay plate so that its eight positions align with the eight positions of the stage. Failure to do so may cause measurement unit damage.



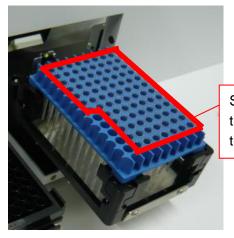
In the case of dispensing measurement, set the source plate on the shuttle at the back of the stage. Set the plate so that the well "A1" on the well plate comes to the top left-hand corner of the stage. In the case of dispensing measurement, be sure to remove the covers of the assay plate and source plate before setting the plates.



- In the case of dispensing measurement, be sure to remove the covers of the assay plate and source plate before setting the plates. Failure to do so may cause dispenser damage.
- The Assay plate and Source plate simulated in the dispensing setting file (refer to 5.12) must be used in dispensing measurement. Failure to do so may cause dispenser damage.
- 4) In dispensing measurement, set a tip rack on the tip rack loader.
 - Dedicated tip rack for CV7000 must be used.
 - (200µl Nested Pipette Tips 105649 from Caliper Life Sciences is used for 96-tip rack model only.)
 - (LT-384-R from Corning is used for 384-tip rack model only.)







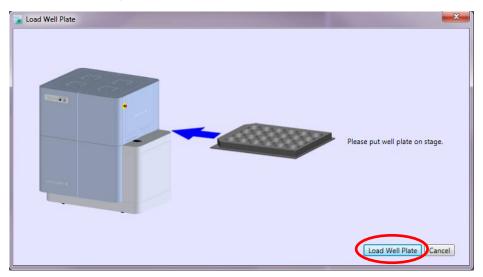
Set a tip rack so that the remaining tip comes to the top-left corner of the tip rack loader.

- Be sure not to use 384-tip rack for 96-tip rack model. Failure to do so may cause dispenser damage.
- Be sure not to use 96-tip rack for 384-tip rack model. Failure to do so may cause dispenser damage.
- Make sure to use dedicated tip rack for each of 96-tip rack model and 384-tip rack model. Failure to do so may cause dispenser damage.

Put the tip disposal box beside CV7000.



5) After setting the plate, click "Load Well Plate" in the "Load Well Plate" screen to move the stage to the reader section.

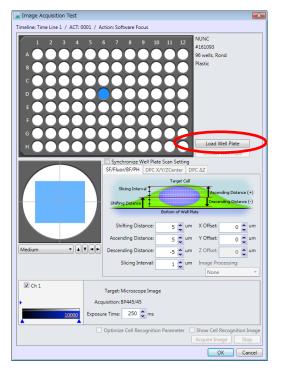


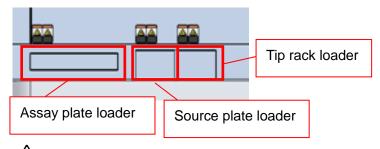


•DO NOT touch loader exit area while moving the loaders.

8.2. Removing the Well Plates

1) When measurement of the assay plate is complete or "Unload Well Plate" on the Image Acquisition Test screen (refer to 5.9 and 6.2) is clicked, the plate is unloaded.

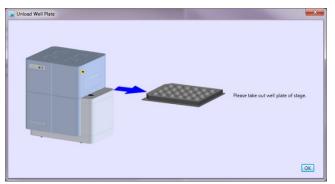




/ WARNING

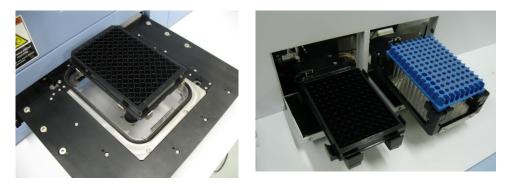
DO NOT touch loader exit area while moving the loaders.

The Unload Well Plate screen opens. Do not click "OK."

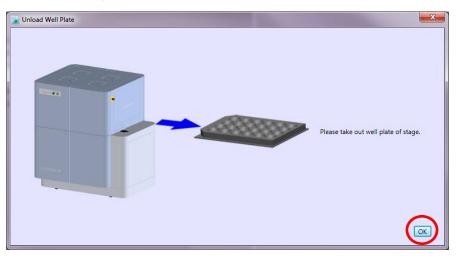


2) Remove the assay plate.

After dispensing measurement, remove also the source plate and tip rack.



3) After removing the plate, click "OK" on the Unload Well Plate screen.





●DO NOT touch loader exit area while moving the loaders.

8.3. Measuring the Assay Plate without Dispensing

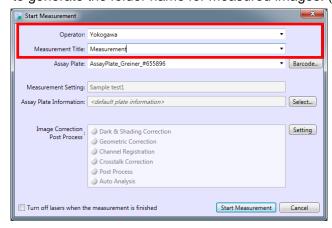
- 1) Create a measurement setting file and save the file. (Refer to Chapter 7)
- Click "Start Measurement" at the bottom of the Status section in the Reader area.

Reader	
Status:	
Ready	
Progression of Time-Lapse:	
Expected Time: Elapsed Time:	
Remaining Time:	
Progression of Current Time Point:	
Expected Time:	::
Elapsed Time: Remaining Time:	::
	Start Measurement More

3) Select the measurement setting file. (Refer to 8.5.)

Filter (OFF)				
easurement Setting	Well Plate Type	Application	Date	
🕒 yokogawa test2	Greiner, #655896, 96 wells, Glass		2012/12/03 15:00:14	
🕒 yokogawa test	Greiner, #655896, 96 wells, Glass		2012/03/15 13:27:40	
🕒 test	BD, #353948, 96 wells, Plastic		2013/04/10 10:37:13	

4) The Start Measurement screen opens. Enter appropriate text in the "Operator" and "Measurement Title" fields. The entered names will be used to generate the folder name for measured images. (Refer to 10.1)



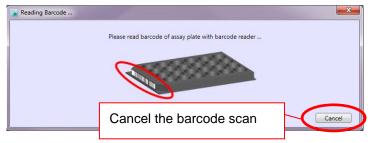
5) Enter appropriate text in the "Assay Plate" field for assay plate name. The entered names will be used to generate the folder names for measured images. (Refer to 10.1)

Operator:	Yokogawa	•	
leasurement Title:	Measurement	•	1
Assay Plate:	AssayPlate_Greiner_#655896	•	Barcoc
Measurement Setting:	Sample test1		
			Select.
Assay Plate Information:	<detault information="" plate=""></detault>		1 Select
Assay Plate Information:	<pre><detault information="" plate=""></detault></pre>		Jelect

Click "Barcode" If there is a barcode on the assay plate. (Barcode reader model only)

🚡 Start Measurement		×
Operator:	Yokogawa 👻	
Measurement Title:	Measurement	
Assay Plate:	AssayPlate_Greiner_#655896	Barcode
Measurement Setting:	Sample test1	
Assay Plate Information:	<default information="" plate=""></default>	Select
Incore Computing		
Image Correction Post Process	Dark & Shading Correction Geometric Correction Channel Registration	Setting
	Crosstalk Correction Post Process	
	Auto Analysis	
Turn off lasers when the	e measurement is finished Start Measurement	Cancel

Barcode reader starts to scan. The scanned barcode data is entered to the "Assay Plate" field.



6) To specify a well plate information file for Assay plate, click "Select" and specify a desired file.

(Measurement can be performed with the default settings.)

🚡 Start Measurement		×
Operator:	Yokogawa 👻	
Measurement Title:	Measurement -	
Assay Plate:	AssayPlate_Greiner_#655896	Barcode
Measurement Setting:	Sample test1	
Assay Plate Information:	<default information="" plate=""></default>	Select
Image Correction Post Process [*]	Dark & Shading Correction Geometric Correction Channel Registration Crosstalk Correction Post Process Auto Analysis	Setting
Turn off lasers when th	e measurement is finished	Cancel

7) Click "Setting" if using post-processing function.

Start Measurement		×
Operator:	Yokogawa 🗸	
Measurement Title:	Measurement -	
Assay Plate:	AssayPlate_Greiner_#655896	Barcode
Measurement Setting:	Sample test1	
Assay Plate Information:	<default information="" plate=""></default>	Select
Image Correction Post Process [:]	Dark & Shading Correction Geometric Correction Channel Registration Crosstalk Correction Post Process Auto Analysis	Setting
Turn off lasers when th	e measurement is finished	Cancel

8) "Measurement Option Settings" window opens.

In case of using post-processing function, check "Use Post Process".

Select "Copy" to automatically copy the measurement data to the designated folder. Select "Move" to automatically move the measurement data to the designated folder.

Measurement Option Set	tings	— X —
Image Correction		
Dark & Shading Correl	ection	
Mode: Dark	: & Shading 🔹	
Use on-the-fly co	rrection	
Geometric Correction		
Interpolation: Near	restNeighbor 🔹	
Use on-the-fly co	rrection	
Channel Registration	(Requires Post Process)	
Mode: Opti	calGroup 🔻	
Crosstalk Correction (Requires Post Process)	
Post Process Use Post Process Copy Move		
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	▼ Browse
Analysis Protocols:	SampleAnalysis	Select
		OK Cancel
		Cancel

9) Designate the folder where to copy or move data from "Select".

Destination:	F:¥BTSData¥MeasurementData •	Browse
Analysis Protocols:		Select

To perform automatic analysis, specify a folder selectable from CV7000 Analysis Software.

(Example: Specify "D:¥BTSData¥MeasurementData" in the WS for analysis as "Destination.")

Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData 🗸	Browse
Analysis Protocols:	SampleAnalysis	Select

To perform automatic analysis, click "Select" of "Analysis Protocol" and then, select analysis protocols to be analyzed.

(About analysis protocol, refer to section 12.6 of IM 80H01C03-01E CV7000 Analysis Software User's Manual)

Click "OK" after selecting.

Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData
Analysis Protocols:	SampleAnalysis Select
Post Process	Setting
Select Analysis Name	Protocols:
Samp Samp	eAnalysis 2 eAnalysis 2 eAnalysis 3 eAnalysis automatic analysis list.
	Select all analysis protocols.
Select All A	nalysis Protocols OK Cancel

MEMO

To display the AnalysisProtocol setting in the Start Measurement screen, launch the CV7000 Analysis Software and set to communicate the network previously. 10) Set image correction setting post imaging (for detail of image correction, please refer to IM 80H01A16-01E Image Correction Software User's Manual). Set "Dark & Shading Correction" and "Geometric Correction". "Dark & Shading" is recommended for "Dark & Shading Correction".

Measurement Option Set	tings	×	
intege Correction			
🕼 Park & Shading Correction			
Mode Dark & Shading			
☑ Use on-the-fly correction			
Geometric Correction			
Interpolation: Near	restNeighbor 🔹		
Use on-the-fly co	rrection		
Channel Registration	(Requires Post Process)		
Mode: OpticalGroup			
Crosstalk Correction (Requires Post Process)		
Post Process			
Use Post Process			
Opy			
Move			
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	 Browse 	
Analysis Protocols:	SampleAnalysis	Select	
		OK Cancel	

11) Check if performing on-the-fly correction.

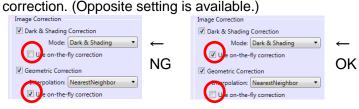
🚡 Measurement Option Set	tings		x
Image Correction			
Dark & Shading Correl	ection		
Mode: Dark	& Shading 🗾 🔻		
🕑 U e on-the-fly co	rrection		
Geometric Correction			
inerpolation: Nea	restNeighbor 🔻		
🗹 U e on-the-fly co	rrection		
Channel Registration	(Requires Post Process)		
Mode: Opti	Mode: OpticalGroup		
Crosstalk Correction	Requires Post Process)		
Post Process			
Use Post Process			
Opy			
Move			
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	 Browse 	-
Analysis Protocols:	SampleAnalysis	Select	
		OK Ca	ncel



 If "Use on-the-fly correction" is checked, image correction is performed before saving image to hard disk.

Processing speed becomes faster than using Post Processor, but raw image data is not saved.

 It cannot to perform "Dark & Shading Correction" without using on-the-fly correction and perform "Geometric Correction" with using on-the-fly correction (Opposite setting is available.)



12) Set "Channel Registration" and "Crosstalk Correction". These 2 items are selectable if "Use Post Process" is checked. Also, they are not correspond to on-the-fly correction.

Measurement Option Set	ings	— ×	
Image Correction			
Dark & Shading Corre	ction		
Mode: Dark	& Shading 🔹		
Use on-the-fly co	rection		
Geometric Correction			
Interpolation: Near	estNeighbor 🔹		
Use on-the-fly co	rection		
Channel Registration	Requires Post Process)		
Mode: Optio			
Ocsstalk Correction (Requires Post Process)			
Use Post Process			
Copy			
Move			
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	Browse	
Analysis Protocols:	SampleAnalysis	Select	
		OK Cance	

MEMO

- •Don't select "Channel Registration" if the images are acquired by high-speed time-lapse mode.
- •To correct fluorescent crosstalk, "Dark & Shading" must be selected in "Dark & Shading Correction", and "Geometric Correction" or "Channel Registration" must be selected.

It is recommended to select both "Geometric Correction" and "Channel Registration".

- "Crosstalk Correction" cannot be performed if "Fluorophore" is not designated in Channel Setting (refer to 5.5)
- •Emission spectrum of fluorophore can be deformed by surrounding environment (pH, temperature, saline concentration). In this case, crosstalk correction cannot be performed normally.

Confirm image after crosstalk correction by Image Correction Software and if the images have abnormality, uncheck "Crosstalk Correction". 13) Click "OK".

Measurement Option Set	tings	 X		
Image Correction		i		
🕼 Dark & Shading Correction				
Mode: Dark	Mode: Dark & Shading			
Use on-the-fly co	rrection			
Geometric Correction				
Interpolation: Near	estNeighbor 🔹			
✓ Use on-the-fly co	rrection			
Channel Registration	(Requires Post Process)			
Mode: Opti	calGroup 🔻			
Crosstalk Correction (Crosstalk Correction (Requires Post Process)			
Post Process				
Use Post Process				
Copy				
Move				
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	Browse		
Analysis Protocols:	SampleAnalysis	select		
		OK Cancel		

14) Click "Start Measurement".

🚡 Start Measurement		— X —
Operator:	Yokogawa 🔻	
Measurement Title:	Measurement -	
Assay Plate:	AssayPlate_Greiner_#655896	Barcode
Measurement Setting:	Sample test1	
Assay Plate Information:	<default information="" plate=""></default>	Select
Image Correction Post Process	Dark & Shading Correction (on-the-fly) Geometric Correction (on-the-fly) Channel Registration Crosstalk Correction Post Process Auto Analysis	Setting
Select this check box if you want to shut down the laser after the measurement.		

- 15) Set well plates on the sample loader. (Refer to 8.1) Measurement will start once the plates are set.
- 16) When the measurement is complete, remove the well plates. (Refer to 8.2)

17) Measured results are displayed. (Refer to Chapter 9)



● To start the laser again, click "Initialize" on the Reader Status tab of the Reader Control screen.

Status	Get	Control Unload Well Plates Initialize Shut Dow
Control Panel: Power On Hardware Status: Access Gate Open Tip Feeder Door Open Drive Unit Ready Operation Possible Ready to Exchange Well Plate	Alarm: Emergency Stop Access Gate Tip Feeder Door Temperature Ventilation Air Supply Environment: Incubator	Incubator On Off Objective Lens O Water Immersion O Dry Temperature: 37 C CO2 Density: 5 %

8.4. Measuring the Assay Plate with Dispensing

- 1) Create a measurement setting file and save the file. (Refer to Chapter 7)
- 2) Click "Start Measurement" at the bottom of the Status section in the Reader area.

Reader	
Status:	
Ready	
Progression of Time-Lapse:	
Expected Time:	
Elapsed Time: Remaining Time:	
Progression of Current Time Point:	
Expected Time:	;;
Elapsed Time: Remaining Time:	::
	Start Measurement More

3) Select the measurement setting file. (Refer to 8.5.)

Measurement Setting List - CellVoyager				
elect a measurement setting :				
Filter (OFF)				
leasurement Setting	Well Plate Type	Application	Date	
🕒 yokogawa test2	Greiner, #655896, 96 wells, Glass		2012/12/03 15:00:14	
O yokogawa test	Greiner, #655896, 96 wells, Glass		2012/03/15 13:27:40	
© test	BD, #353948, 96 wells, Plastic		2013/04/10 10:37:13	
			OK	Cancel

4) The Start Measurement screen opens. Enter appropriate text in the "Operator" and "Measurement Title" fields. The entered names will be used to generate the folder name for measured images. (Refer to 10.1)

operation	Yokogawa 🔹	
Measurement Title:	Measurement 🔹	
Assay Plate:	AssayPlate_Greiner_#655896	Barcode.
Measurement Setting:	Sample test	
Assay Plate Information:		Select
Dispense Setting:		id Volume
	dispense from 1 source plate to 1 assay plate	
Source Plate List:	Source Plate Source Plate Informaion	Change
	1	
Assay Plate Mapping:	Assay Plate 1	
Image Correction		Setting
Post Process	Dark & Shading Correction Geometric Correction	setting
	Channel Registration	
	Crosstalk Correction	
	Post Process	
	Auto Analysis	

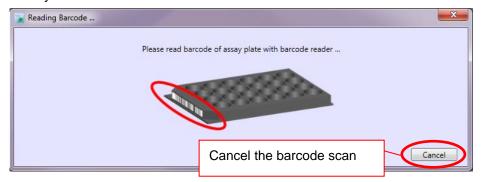
5) Enter appropriate text in the "Assay Plate" field for assay plate name. The entered names will be used to generate the folder name for measured images. (Refer to 10.1)

Start Measurement		— X
Operator:	Yokogawa 👻	
Measurement Title:	Measurement 🔹	_
Assay Plate:	AssayPlate_Greiner_#655896 🗸	Barcode
Measurement Setting:	Sample test	
Assay Plate Information:		Select
Dispense Setting:	SampleTest Liqu	id Volume
	dispense from 1 source plate to 1 assay plate	
Source Plate List:	Source Plate Source Plate Informaion	Change
	1	
Assay Plate Mapping:	Assay Plate 1 🔹	
Image Correction Post Process	Dark & Shading Correction Geometric Correction Channel Registration Crosstalk Correction Post Process Auto Analysis	Setting
Turn off lasers when th	e measurement is finished	Cancel

Click "Barcode" If there is a barcode on the assay plate. (Barcode reader model only)

🚡 Start Measurement			x
Operator:	Yokogawa	•	
Measurement Title:	Measurement	•	
Assay Plate:	AssayPlate_Greiner_#655896		Barcode
Measurement Setting:	Sample test		
Assay Plate Information:			Select
Dispense Setting:	SampleTest	Liqu	id Volume
	dispense from 1 source plate to		
Source Plate List:	Source Plate	Source Plate Informaion	Change
	1		
Assay Plate Mapping:	Arrest Dista 1		
Assay Plate Mapping:	Assay Plate 1		
Image Correction Post Process	Dark & Shading Correction		Setting
Post Process	Geometric Correction		
	Channel Registration		
	Crosstalk Correction		
	Post Process		
	Auto Analysis		
Turn off lasers when the	e measurement is finished	Start Measurement	Cancel

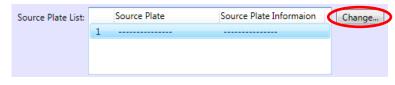
Barcode reader starts to scan. The scanned barcode data is entered to the "Assay Plate" field.



6) Click "Select" and select a desired well plate information file for the assay plate. (Refer to 4.1)

🚡 Start Measurement			×
Operator:	Yokogawa	•	
Measurement Title:	Measurement	•	
Assay Plate:	AssayPlate_Greiner_#655896	•	Barcode
Measurement Setting:	Sample test		
Assay Plate Information:			Select
Dispense Setting:	SampleTest	Liqu	id Volume
	dispense from 1 source plate to	1 assay plate	
Source Plate List:	Source Plate	Source Plate Informaion	Change
	1		
Assay Plate Mapping:	Assay Plate 1 🔹		
Image Correction	Dark & Shading Correction		Setting
Post Process	Geometric Correction		
	Channel Registration		
	Crosstalk Correction		
	Ø Post Process		
	Auto Analysis		
Turn off lasers when th	e measurement is finished	Start Measurement	Cancel

7) Click "Change" and select a well plate information file for the source plate. (Refer to 4.1)



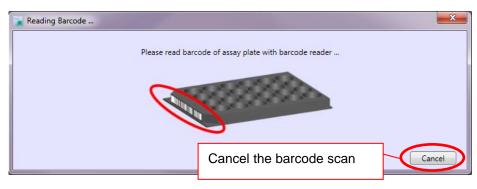
8) Enter appropriate text in the "Source Plate" field for source plate name.

ſ	Source Plate Setting				
	Enter the information of source plate 1:				
İ		Source Plate:	Source Plate #1	•	Barcode
	Source Pla	te Information:			Select
				ОК	Cancel

Click "Barcode" If there is a barcode on the Source plate. (Barcode reader model only)

ſ	Source Plate Setting		×
	Enter the information of source	e plate 1:	
İ	Source Plate:	Source Plate #1	• Barcode
	Source Plate Information:		Select
			OK Cancel

Barcode reader starts to scan. The scanned barcode data is entered to the "Source Plate" field.



9) Click "Select" and select a well plate information file for the source plate. (Refer to 4.1)

Source Plate Setting			×
Enter the information of s	ource plate 1:		
Source Pl	late: Source Plate #1	•	Barcode
Source Plate Informat	ion: #655896 Source Plate		Select
		ОК	Cancel
Source Plate List:	Source Plate Source Plate #1	Source Plate Informaion #655896 Source Plate	Change
Assay Plate Mapping:	Assay Plate 1 🔹		

10) Select "Assay Plate Mapping." If multiple assay plates were set in the dispensing setting file (refer to 5.12), solution is dispensed only to the wells in the assay plate selected in "Assay Plate Mapping."

Assay Plate Mapping:	Assay Plate 1 🛛 💊	•	
	A 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2		Assay plate 1 Assay plate 2
	ultiple assay plate	s cannot be measu	red.

11) Click "Setting" if using post-processing function.

Start Measurement			×
Operator:	Yokogawa	•	
Measurement Title:	Measurement	•	
Assay Plate:	AssayPlate_Greiner_#655896	•	Barcode
Measurement Setting:	Sample test		
Assay Plate Information:			Select
Dispense Setting:	SampleTest	Liqu	id Volume
	dispense from 1 source plate to	1 assay plate	
Source Plate List:	Source Plate	Source Plate Informaion	Change
	1		
Assay Plate Mapping:	Assay Plate 1 🔹	(
Image Correction	Dark & Shading Correction		Setting
Post Process	Geometric Correction		
	Ohannel Registration		
	Crosstalk Correction		
	Post Process		
	Auto Analysis		
Turn off lasers when th	e measurement is finished	Start Measurement	Cancel

12) "Measurement Option Settings" window opens.

In case of using post-processing function, check "Use Post Process".

Select "Copy" to automatically copy the measurement data to the designated folder. Select "Move" to automatically move the measurement data to the designated folder.

🃡 Measurement Option Set	tings	×
Image Correction		
Dark & Shading Correl	ection	
Mode: Dark	& Shading 🔹	
Use on-the-fly co	rrection	
Geometric Correction		
Interpolation: Near	estNeighbor 🔹	
Use on-the-fly co	rrection	
Channel Registration	(Requires Post Process)	
Mode: Opti	calGroup	
Crosstalk Correction (Requires Post Process)	
Post Process Use Post Process Copy Move		
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	▼ Browse
Analysis Protocols:	SampleAnalysis	Select
		OK Cancel

13	Designate	the folder	where	to copy of	or move	data from	"Select"
10,	Designate		WIICIC	to copy t		uata nom	001001

Destination:	F:¥BTSData¥MeasurementData •	Browse
Analysis Protocols:		Select

To perform automatic analysis, specify a folder selectable from CV7000 Analysis Software.

(Example: Specify "D:¥BTSData¥MeasurementData" in the WS for analysis as "Destination.")

Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData 🔹	Browse
Analysis Protocols:	SampleAnalysis	Select

To perform automatic analysis, click "Select" of "Analysis Protocol" and then, select analysis protocols to be analyzed.

(About analysis protocol, refer to section 12.6 of IM 80H01C03-01E CV7000 Analysis Software User's Manual)

Click "OK" after selecting.

Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData • Browse
Analysis Protocols:	SampleAnalysis Select
🕞 Post Process	Setting
Select Analysis	Protocols:
Sampl	eAnalysis 2 eAnalysis 2 eAnalysis 3 Analysis protocols registered in automatic analysis list.
	Select all analysis protocols.
Select All A	nalysis Protocols OK Cancel

МЕМО

To display the AnalysisProtocol setting in the Start Measurement screen, launch the CV7000 Analysis Software and set to communicate the network previously. 14) Set image correction setting post imaging (for detail of image correction, please refer to IM 80H01A16-01E Image Correction Software User's Manual). Set "Dark & Shading Correction" and "Geometric Correction". "Dark & Shading" is recommended for "Dark & Shading Correction".

Intege Correction			
🔽 Fark & Shading Co			
Mode D	ark & Shading		
Use on-the-fly	correction		
eometric Correct	ion		
Interpolation: N	learestNeighbor 🔹		
✓ Use on-the-fly	correction		
Channel Registrati	on (Requires Post Process)		
Mode: C	on (Requires Post Process) opticalGroup		
Mode: C			
Mode: 🖸	pticalGroup		
Mode: C	pticalGroup		
Mode: Q Crosstalk Correction Post Process Use Post Process	pticalGroup		
Mode: Crosstalk Correction Post Process Use Post Process Copy Move	pticalGroup	• Brow	se
Mode: Crosstalk Correction Post Process Use Post Process Copy Move Destination	vpticalGroup v nn (Requires Post Process)	Brown Select	

15) Check if performing on-the-fly correction.

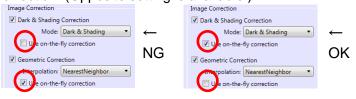
Measurement Option Set	tings		×
Image Correction Dark & Shading Corre Mode: Dark			
Geometric Correction			
Geometric Correction	restNeighbor 🔹		
Channel Registration Mode: Opti			
Crosstalk Correction (Requires Post Process)		
Post Process			
Use Post Process			
 Copy Move 			
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	•	Browse
Analysis Protocols:	SampleAnalysis		Select
			K Cancel

MEMO

If "Use on-the-fly correction" is checked, image correction is performed before saving image to hard disk.

Processing speed becomes faster than using Post Processor, but raw image data is not saved.

 It cannot to perform "Dark & Shading Correction" without using on-the-fly correction and perform "Geometric Correction" with using on-the-fly correction. (Opposite setting is available.)



16) Set "Channel Registration" and "Crosstalk Correction". These 2 items are selectable if "Use Post Process" is checked. Also, they are not correspond to on-the-fly correction.

Measurement Option Set	tings		x
Image Correction			
Dark & Shading Corre	ection		
Mode: Dark	& Shading 🔹		
Use on-the-fly co	rrection		
Geometric Correction			
Interpolation: Near	estNeighbor 🔹		
Use on-the-fly co	rrection		
Channel Registration	(Requires Post Process)		
Mode: Opti	· · · · · · · · · · · · · · · · · · ·		
Cosstalk Correction (Requires Post Process)		
	, ,		
Post Process			
Use Post Process			
Opy			
Move			
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	 Browse. 	
Analysis Protocols:	SampleAnalysis	Select.	
		OK Ca	ance
			anice

MEMO

•Don't select "Channel Registration" if the images are acquired by high-speed time-lapse mode.

•To correct fluorescent crosstalk, "Dark & Shading" must be selected in "Dark & Shading Correction", and "Geometric Correction" or "Channel Registration" must be selected.

It is recommended to select both "Geometric Correction" and "Channel Registration".

- •"Crosstalk Correction" cannot be performed if "Fluorophore" is not designated in Channel Setting (refer to 5.5)
- •Emission spectrum of fluorophore can be deformed by surrounding environment (pH, temperature, saline concentration). In this case, crosstalk correction cannot be performed normally.

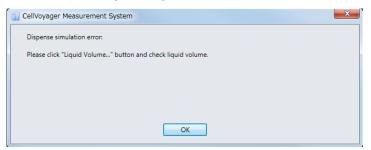
Confirm image after crosstalk correction by Image Correction Software and if the images have abnormality, uncheck "Crosstalk Correction".

Measurement Option Set	tings	— ×
Image Correction		
Dark & Shading Correl	ection	
Mode: Dark	& Shading 🔹	
✓ Use on-the-fly co	rrection	
Geometric Correction		
Interpolation: Nea	estNeighbor 🔹	
☑ Use on-the-fly co	rrection	
Channel Registration	(Requires Post Process)	
Mode: Opti	· · · · · · · · · · · · · · · · · · ·	
Crosstalk Correction (Requires Post Process)	
Post Process		
Use Post Process		
Copy		
Move		
Destination:	¥¥192.168.0.55¥BTSData¥MeasurementData	Browse
Analysis Protocols:	SampleAnalysis	Select
		OK Cancel

18) Click "Start Measurement".

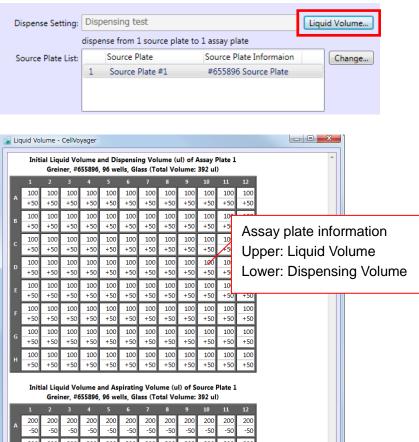
🏹 Start Measurement		×	
Operator:	Yokogawa		
Measurement Title:	Measurement •		
Assay Plate:	AssayPlate_Greiner_#655896	Barcode	
Measurement Setting:	Sample test		
Assay Plate Information:		Select	
Dispense Setting:	SampleTest Liqu	id Volume	
	dispense from 1 source plate to 1 assay plate		
Source Plate List:	Source Plate Source Plate Informaion	Change	
	1		
Assay Plate Mapping:	Assay Plate 1		
Image Correction		Contine	
Post Process			
Channel Registration			
	Crosstalk Correction		
	Post Process Auto Analysis		
\frown	Auto Analysis		
	this check box if you want to shu	it down	
the laser after the measurement.			

Measurement cannot start if any error is contained in the result of simulation in dispensing setting screen. (Refer to 5.12.)





 In the case of error, click "Liquid Volume" to confirm error point. To check error in detail, confirm result obtained by dispensing simulation in dispensing setting screen. (Refer to 5.12.)



200 200 200 200 200 200 200 200 200 200 200 200 -50 -50 -50 -50 -50 -50 -50 -50 -50 -50 -50 200 -50 200 -50 200 200 200 200 -50 200 200 200 200 200 Source plate information -50 -50 -50 -50 -50 -50 -50 -50 Upper: Liquid Volume 200 200 200 200 200 200 200 200 200 20 -50 -50 -50 -50 -50 -50 -50 -50 -50 Lower: Aspirating Volume 200 200 20 200 200 200 200 200 200 200 -50 -50 -50 -50 -50 -5 -5(-5 -5 200 200 200 200 200 200 200 200 200 200 200 200 -50 -50 -50 -50 -50 -5(-5 -50 -50 -50 -50 200 200 200 200 200 200 200 200 200 200 200 200 -50 -50 -50 -50 -50 -50 -50 -50 50 00 Error 200 200 200 200 200 200 200 200 Close

Liquid Volume screen

19) Set well plates on the sample loader. (Refer to 8.1)

Measurement will start once the plates are set.

- In the case of dispensing measurement, be sure to remove the covers of the assay plate and source plate before setting the plates. Failure to do so may cause dispenser damage.
- The Assay plate and Source plate simulated in the dispensing setting file (refer to 5.12) must be used in dispensing measurement. Failure to do so may cause dispenser damage.
- 20) When the measurement is complete, remove the well plates. (Refer to 8.2)
- 21) Measured results are displayed. (Refer to Chapter 9)

МЕМО

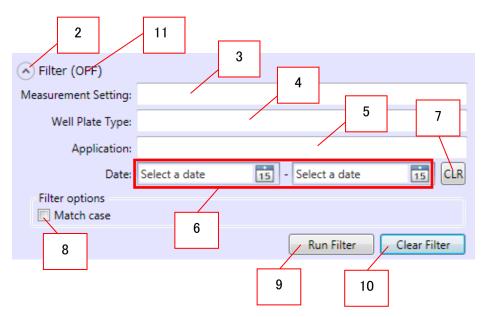
To start the laser again, click "Initialize" on the Reader Status tab of the Reader Control screen.

Reader Laser Temperature Tip Dispenser	Water CO2
Hardware Status: Access Gate Open Tip Feeder Door Open Drive Unit Ready Operation Possible Ready to Exchange Well Plate Enviro	Get Control Incubator Incubator Emergency Stop On Off Objective Lens Access Gate On Off Water Immersion Dry Tip Feeder Door Temperature 37 🔍 *C Ventilation CO2 Density: 5 🗳 % Air Supply I confirmed that CO2 detector is working

8.5. Measurement Setting List Screen

Measurement Setting Well Plate Type Application Date Image: Test 80, #353948, 96 wells, Plastic 2013/04/10 10:37:13 Image: Test Greiner, #655896, 96 wells, Glass 2012/12/03 15:00:14 Image: Test Greiner, #655896, 96 wells, Glass 2012/03/15 13:27:40	Measurement Settin Select a measurem Filter (ON) Measurement Setting: Well Plate Type: Application: Date: Filter options Match case		1		
©yokogawa test2 Greiner, #655896, 96 wells, Glass 2012/12/03 15:00:14	Measurement Setting	Well Plate Type	Application	Date	
	Test 💽	BD, #353948, 96 wells, Plastic		2013/04/10 10:37:13	
©yokogawa test Greiner, #655896, 96 wells, Glass 2012/03/15 13:27:40	🔟 yokogawa test2	Greiner, #655896, 96 wells, Glass		2012/12/03 15:00:14	
	🚾 yokogawa test	Greiner, #655896, 96 wells, Glass		2012/03/15 13:27:40	

1) Screen for search filter setting



- 2) Display the screen for search filter setting.
- 3) Character string of Measurement Setting to perform search filter.
- 4) Character string of Well Plate Type to perform search filter.
- 5) Character string of Application to perform search filter.
- 6) Date range to perform search filter.
- 7) Clear Date range you have specified.
- 8) Perform search filter by case-sensitive.
- 9) Perform search filter.
- 10) Clear result of search filter.
- 11) Search filter ON/OFF

	📕 Sta	art Measurement		1
		Operator:	Yokogawa	
		Measurement Title:	Measurement	2
	4	Assay Plate:	AssayPlate_Greiner_#655896	
	M	leasurement Setting:	Sample test1	3
	Assa	ay Plate Information:	<default information="" plate=""></default>	Select
5]	Image Correction Post Process	 Dark & Shading Correction (on-the-fly) Geometric Correction (on-the-fly) 	Setting
			Channel Registration Crosstalk Correction Post Process	
		urn off lasers when th	Auto Analysis me measurement is finished	Cancel
		7	6 8 9	

- 1) Operator name
- 2) Measurement title
- 3) Assay plate name
- 4) Measurement setting file
- 5) Well plate information file of the Assay plate
- 6) Setting items of postprocessor and image correction.
- (Click "Setting" button to open setting screen described in next page.)
- 7) Automatically turns OFF laser power when the measurement finished.
- 8) Start measurement.
- 9) Cancel measurement.

Measurement Option Settings	
Image Correction	
 ✓ Dark & Shading Correction Mode: Dark & Shading ▼ ✓ Use on-the-fly correction 	1
 Geometric Correction Interpolation: NearestNeighbor Use on-the-fly correction 	2
Channel Registration (Requires Post Process) Mode: OpticalGroup	3
Crosstalk Correction (Requires Post Process)	4
Post Process ✓ Use Post Process O Copy O Copy	5
 Move Destination: ¥¥192.168.0.55¥BTSDa Analysis Protocols: SampleAnalysis 	ta¥MeasurementData • Browse Select
	OK Canc

1) Background/ Shading correction setting

Item		Explanation	
Dark & Shading Correction		Correct the background offset level and shading	
		of measured images.	
Mode	Dark & Chading	Correct the background offset level and shading	
	Dark & Shading	of measured images.	
	Dark	Correct the background offset level of measured	
Dark		images.	
Use on-the-fly correction		Perform on-the-fly correction	

2) Geometric correction setting

Item		Explanation	
Geometric Correction		Correct scaling, rotation and parallel shift effects.	
	NearestNeighbor	Use the value of the nearest pixel to correct	
Interpola	Inealestineighbol	images.	
tion	Bilinear	Decide the pixel value by the distance and	
Dillieal		variation of each of nearby four pixels.	
Use on-the-fly correction		Perform on-the-fly correction	

3) Registration correction setting

Item		Explanation	
Channel Registration		Correct pixel shift between channels.	
		Correct pixel shift by comparing the correlation	
Mode	CompareBased	between the reference channel and each	
		channel.	
	OpticalGroup	Correct pixel shift by comparing the correlation	
		between groups of each XY stage position.	

4) Setting of crosstalk correction

5) Setting items of postprocess

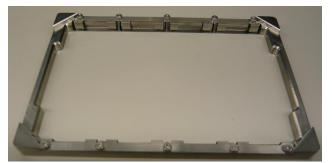
Item	Description		
Use Post	Post processing starts after measurement completed when		
Process	checked.		
Сору	Automatically copy measurement data to designated folder.		
Move	Automatically move measurement data to designated folder.		
	Designate folder to copy or move data by "Select".		
Destination	To perform automatic analysis, specify a folder selectable from		
	the Analysis Support software.		
Analysis	Specify analysis protocols by clicking "Select."		
Protocol	(Analysis software supported model only)		

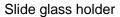
6) Close the screen with saving image correction/ post process setting.

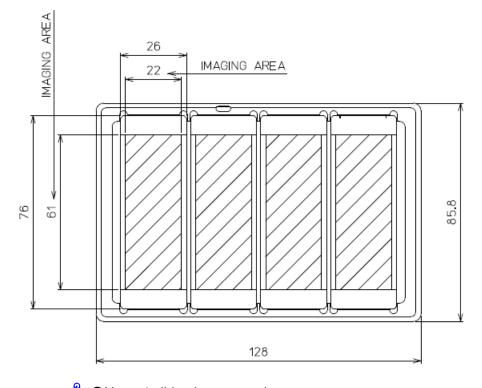
7) Close the screen without saving image correction/ post process setting.

8.7. Set Slide Glasses on Slide Glass Holder

In the case of observation by slide glass, set slides on slide glass holder. (Only slide glass holder supported model.)







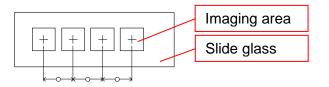
мемо

●Up to 4 slide glasses can be set.

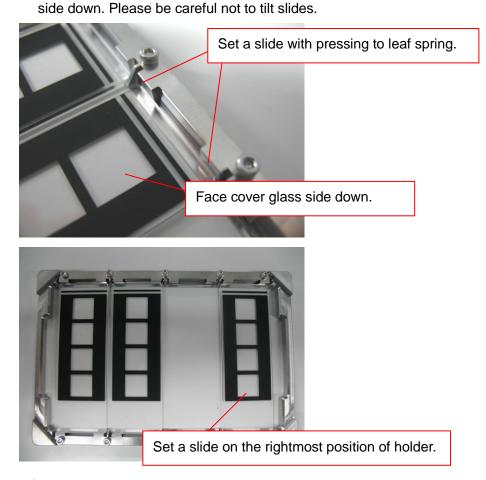
Observable range of slide glass is 61mm X 22mm.

The range can be small when water immersion lens is used.

Slide glass that can be analyzed by CV7000 Analysis Software is what 4 imaging areas are arranged at regular intervals as following diagram.



8-33



•Supported silde glasses are as following which meet ISO 8073/1 (JIS R 3703).

Length [mm]: 76.0 0/-1

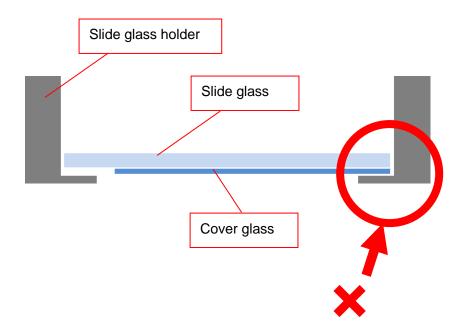
Width [mm]: 26.0 0/-1

Don't use slide glasses not to meet above condition. Slide glasses could be broken if they are set by sheer force. Besides, slide glasses could fall into equipment during measurement and injure the equipment.

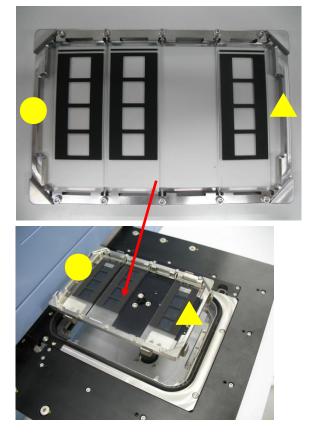
Cover glass must be fixed on slide glass by manicure and others not to fall into machine.

- If the slide glasses not to meet ISO/ JIS, slide glass can't be fixed on slide glass holder and imaging area shift or focus error could occur.
- •Set a slide glass on the rightmost position of holder. Sample isn't needed to set on this slide glass.

•Set slides not to put cover glass on sample holder.



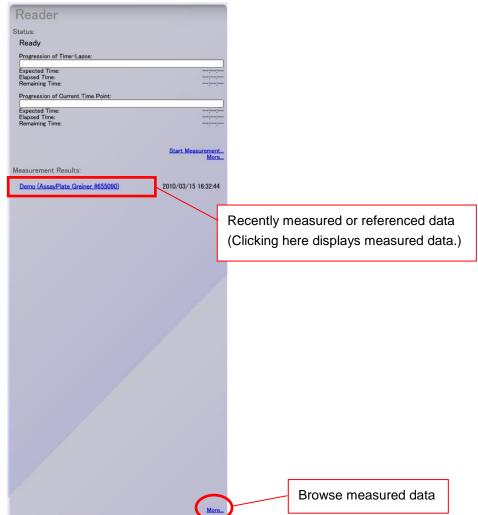
3) Set slide glass holder on the stage. Be careful that direction of holder is correct. For the procedure of setting to the stage, please refer to 8.1.



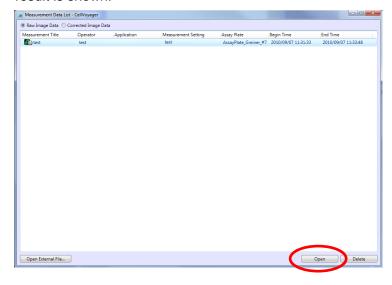
9. Checking Measured Images

9.1. Opening the Measurement Result Screen

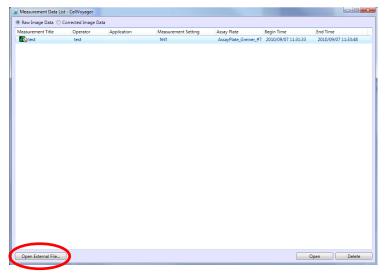
1) Click "More" under "Measurement Results" in the Reader Area.



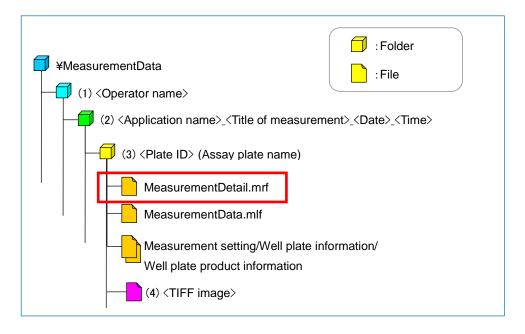
The Measurement Data List screen opens. (Refer to 9.3)
 Select a desired measurement data and then click "Open." The measured result is shown.

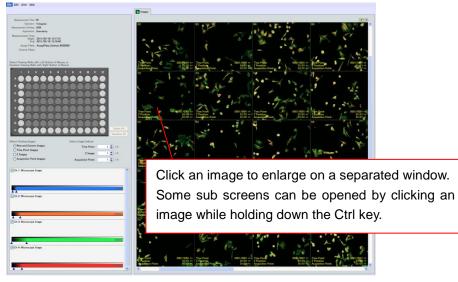


Click "Open External File" if you open the measured result from the external folder you saved.

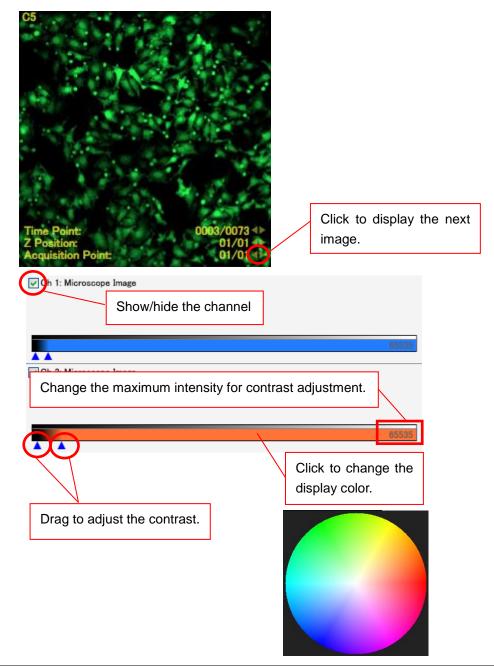


Select the "MeasurementDetail.mrf" file in the measurement folder. The measured result is shown.

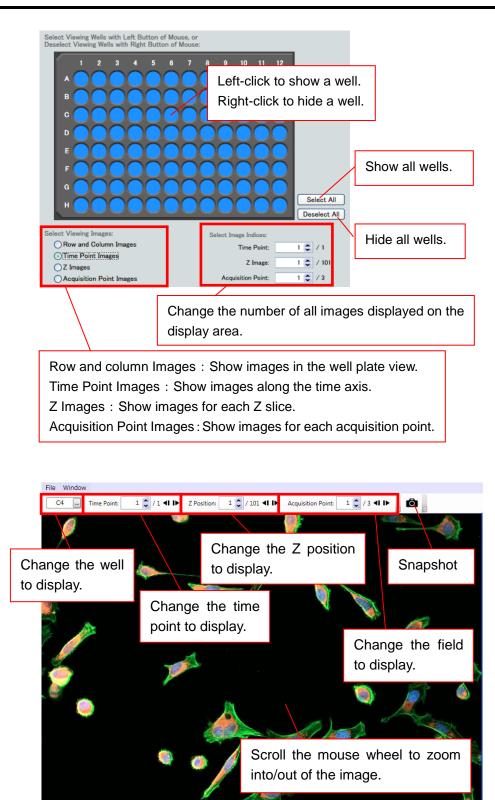




Output screen for measured results (Refer to 9.3)

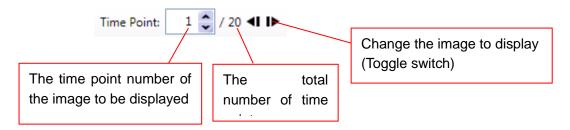


IM 80H01C01-01E



Sub screen for measurement results

Pixel coordinate and each channel of intensity



"Z Position" and "Acquisition Point" is in common with "Time Point"

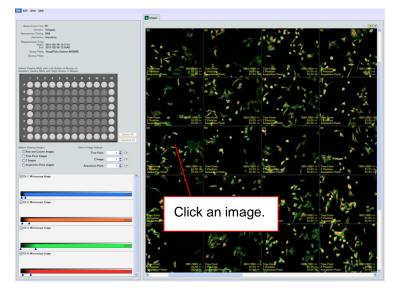
9.2. Creating Movie Files

Movie files can be created from the measurement data acquired by time-lapse imaging.

Creating the Movie File for a Single Field

The movie file about the field displaying on the sub screen is created.

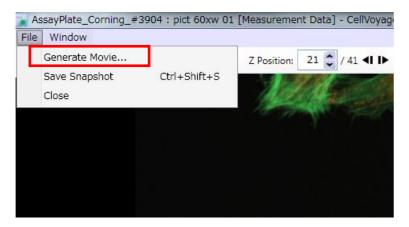
1) An image is displayed to the sub screen by clicking a well image to create movie files. (Refer to 9.3.)



2) From the sub screen, select the field to create movie files.



3) Click "Generate Movie" from the "File" menu of the sub screen.



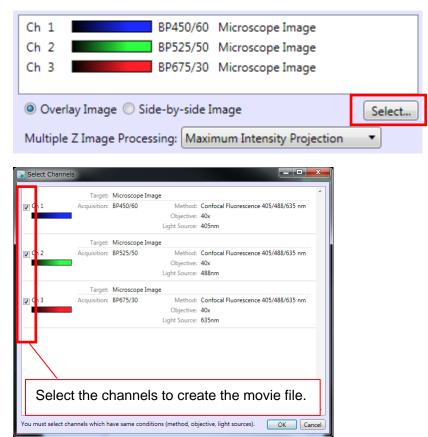
The "Generate Time-lapse Movie" screen is displayed.

Generate Time-lapse Movie
Acquisition Point: No. 1 of H12 well Time Point: from 1 to 51 to
Targets: Ch 1 BP450/60 Microscope Image Ch 2 BP525/50 Microscope Image Ch 3 BP675/30 Microscope Image
Overlay Image Side-by-side Image Select Multiple Z Image Processing: Maximum Intensity Projection
Movie Setting: Video Compressor: Uncompressed Frame Rate: 5 fps
Output Folder: C:¥BTSData¥MeasurementData¥Yokogawa¥test_2010090

4) Specify the time point range to create movie files.

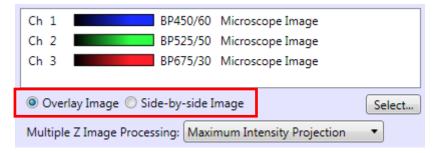
Time Point:			
Time Point:	from	1 🗘 to	51 🌲

5) Select the channels to create the movie file by clicking "Select."

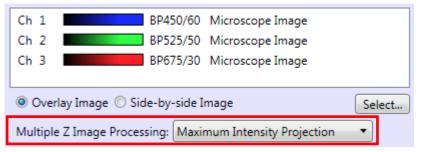


6) If selecting "Overlay Image," the movie file is created with overlaying channel images.

If selecting "Side-by-side Image," the movie file is created with tiling channel images.



7) Select an output item for Z images from "Multiple Z Image Processing," if selecting the channels which have Z-stack images.

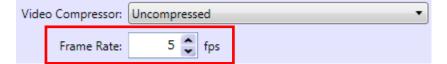


Item	Explanation	
None	No Z-stack image exists.	
Maximum Intensity Projection	Create movie files using the MIP images.	
Maximum intensity Projection	(Refer to 5.1.)	
Minimum Intensity Projection	Create movie files using the MinIP images.	
	(Refer to 5.1.)	
Average Intensity Projection	Create movie files using the AIP images.	
Average Intensity Projection	(Refer to 5.1.)	
	Create movie files using the Z images which have the	
Maximum Intensity Image	highest total intensity in the Z-stack images at each	
	time point.	
Sum Intensity Projection	Create movie files using SUM images. (Refer to 5.1.)	

 Select a compression method for movie files from "Video Compressor." If compressing movie files, "Cinepak Codec" is recommended. If not compressing movie files, select "Uncompressed."

Video Compressor:	Uncompressed 🔹
	Uncompressed
Frame Rate:	MJPEG Compressor
	Cinepak Codec by Radius
	Microsoft Video 1

9) Specify the frame rate for movie replay.



10) Specify a storage folder for movie files.

Output Folder:	C:¥BTSData¥MeasurementData¥Yokogawa¥10x tile_2011			
	Specify a storage folder.			

11) To create movie files is started by clicking "Generate Movie."

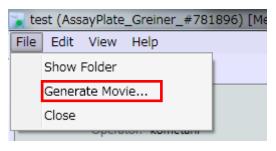
Generate Movie Close	
Curter Tarter Movine Crosse Curter Folder: DABTSDataMeasurementData Progression of All Movie Files: AssayPlate, Greiner #655886 E4 F001 T0001,T0003.avi 1 of 1 File Progression of Current Movie: The Point 2 Control of Current Movie: The Point 2 Control of Current Movie: The Point 2 Control of Current Movie: The Point 2 Control of Current Movie: Control of Curre	Display the output folder.
Canop	Stop creating the movie files.

The screen to display the processing for creating movie files

Creating the Movie Files for All Fields

The movie files for the entire fields in all wells are created.

1) Click "Generate Movie" from the "File" menu in the measurement result screen. (Refer to 9.3.)



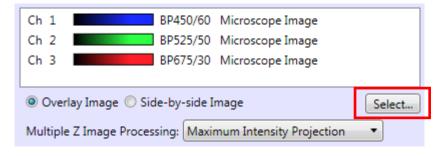
The "Generate Time-lapse Movie" screen is displayed.

🚡 Generate Time-la	apse Movie
Acquisition Point:	All acquisition points of assay plate
Time Point:	from 1 🔪 to 51 💭
Targets:	Ch 1 BP450/60 Microscope Image
	Ch 2 BP525/50 Microscope Image
	Ch 3 BP675/30 Microscope Image
	Overlay Image Side-by-side Image Select Multiple Z Image Processing: Maximum Intensity Projection
Movie Setting:	Video Compressor: Uncompressed
	Frame Rate: 5 🗘 fps
Output Folder:	C:¥BTSData¥MeasurementData¥Yokogawa¥test_2010090
	Generate Movie Close

2) Specify the time point range to create movie files.



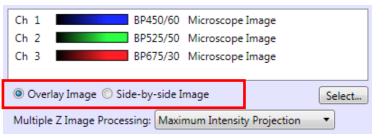
3) Select the channels to create movie files by clicking "Select."



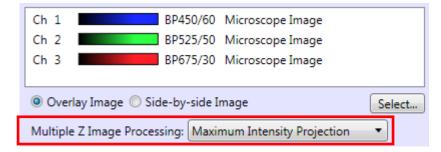
Select Channels			×
Tarriet	Microscope Image		*
Ch1 Acquisition:	1 2	Confocal Fluorescence 405/488/635 nm	- 1
	Objective:		
	Light Source:	405nm	
Target:	Microscope Image		
Ch 2 Acquisition:	BP525/50 Method:	Confocal Fluorescence 405/488/635 nm	
	Objective:	40x	
	Light Source:	488nm	
Target:	Microscope Image		
Ch 3 Acquisition:	BP675/30 Method:	Confocal Fluorescence 405/488/635 nm	
	Objective:	40x	
	Light Source:	635nm	
Select the c	hannels to cre	ate movie files.	
			*
You must select channels which h	ave same conditions (method, ob	jective, light sources).	Cancel

4) If selecting "Overlay Image," movie files are created with overlaying channel images.

If selecting "Side-by-side Image," movie files are created with tiling channel images.



5) Select an output item for Z images from "Multiple Z Image Processing," if selecting the channels which have Z-stack images.



Item	Explanation
None	No Z-stack image exists.
Maximum Intensity Projection	Create movie files using the MIP images.
	(Refer to 5.1.)
Minimum Intensity Projection	Create movie files using the MinIP images.
	(Refer to 5.1.)
Average Intensity Projection	Create movie files using the AIP images.
Average Intensity Projection	(Refer to 5.1.)
	Create movie files using the Z images which
Maximum Intensity Image	have the highest total intensity in the Z-stack
	images at each time point.
Sum Intensity Projection	Create movie files using SUM images.
	(Refer to 5.1.)

 6) Select a compression method for movie files from "Video Compressor." If compressing movie files, "Cinepak Codec" is recommended. If not compressing movie files, select "Uncompressed."

	Video Compressor:	Uncompressed 🔹
		Uncompressed
	Frame Rate:	MJPEG Compressor
l		Cinepak Codec by Radius
		Microsoft Video 1

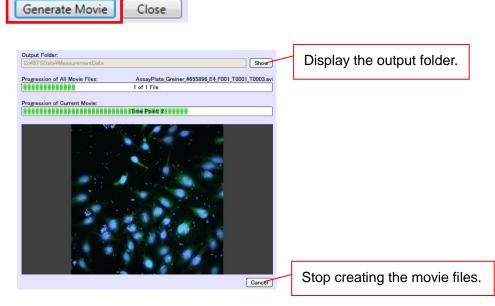
7) Specify the frame rate for movie replay.

Video Compressor:		Uncompressed	•
	Frame Rate:	5 🗘 fps	

8) Specify a storage folder for movie files.

Output Folder:	C:¥BTSData¥MeasurementData¥Yokogawa¥10x tile_2011		
	Specify a storage folder.		

9) To create movie files is started by clicking "Generate Movie."



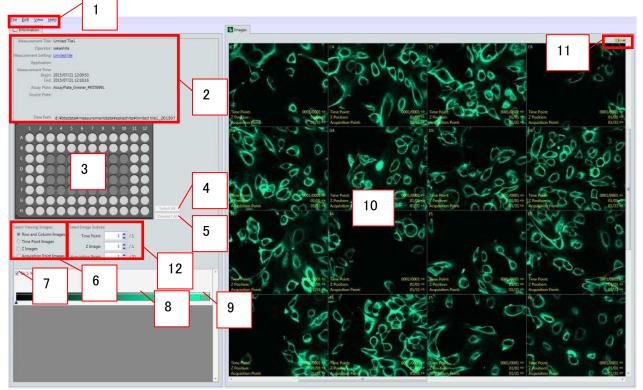
The screen to display the processing for creating movie files

9.3. Explanation of Measurement Result Screens

Measurement Data List Screen

11	12					
🕞 Neasuremer	nt Data List - CellVoyage	r				
Raw Image	Data Corrected Imag	je Data				
Measurement	t Title Operator test	Application	Measurement Setting test	Assay Plate	Begin Time er_#7 2010/09/07 11:31:33	End Time 2010/09/07 11:33:48
1	2	3	4	5	6	7
8					9	10
Open Exter	nal File				d -	Open Delete

- 1) Title of measurement
- 2) Name of the person who performed measurement
- 3) Application name
- 4) Measurement setting file name
- 5) Assay plate name
- 6) Start time of measurement
- 7) End time of measurement
- 8) Select any measurement data.
- 9) Open the selected set of measured data.
- 10) Delete the selected set of measured data.
- 11) Show measured data files.
- 12) Show corrected image data files. (Refer to IM 80H01A16-01E)



Measurement Result Display Screen

1) Menu			
File menu	Explanation		
Show Folder	Display the storage folder of the measurement data.		
Generate Movie	Display the screen to create movie files.		
Close	Close the measurement result display screen.		

Edit menu	Explanation
Undo	Undo the last operation.
Redo	Redo the last operation.
Cut	Cut the selected item.
Сору	Copy the selected item.
Paste	Paste the selected item.

View menu	Explanation	
Row and Column Images	Show images in the well plate view.	
Time Point Images	Show images along the time axis.	
Z Images	Show images for each Z slice.	
Well and Acquisition Point	Show images for each acquisition point.	
Images		
1 Image	Show the display area by 1×1.	
2 Images	Show the display area by 2x2.	
3 Images	Show the display area by 3x3.	
4 Images	Show the display area by 4×4.	

Help menu	Explanation
About	Show the version information of the measurement software.

2) Information items on measured data

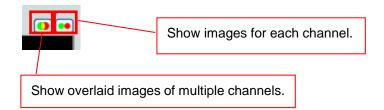
Item	Explanation
Measurement Title	Title of measurement
Operator	Name of the person who performed measurement
Measurement Setting	Measurement setting file name
Application	Application name
Measurement Time Begin	Start time of measurement
Measurement Time End	End time of measurement
Assay Plate	Assay plate name
Source Plate	Source plate name
Data Path	Path of data saved folder

3) Select the well to display the images of.

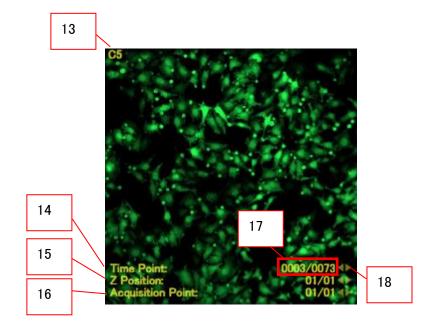
- 4) Select all wells.
- 5) Delete all wells.
- 6) Select the display format of the display area.

Item	Explanation
Row and Column Images	Show images in the well plate view.
Time Point Images	Show images along the time axis.
Z Images	Show images for each Z slice.
Acquisition Point Images	Show images for each acquisition point.

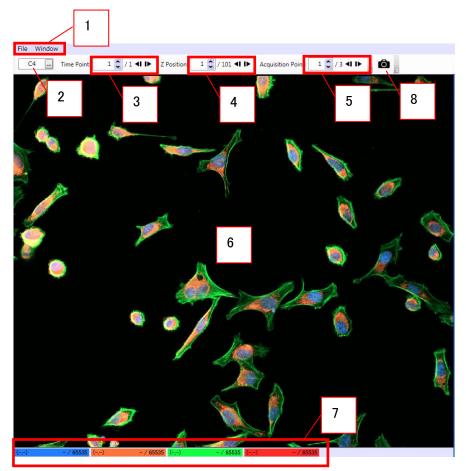
- 7) Show/hide channel images.
- 8) Adjust the contrast/change the display color.
- 9) Setting of the maximum intensity for contrast adjustment
- 10) Display area
- 11) Select the display format for channel images.



12) Change the number of all images displayed on the display area. (Image number of the image being displayed/total number of images)



- 13) Well number
- 14) Time series
- 15) Z position
- 16) View field
- 17) Image number of the image being displayed / total number of images
- 18) Show the next image.



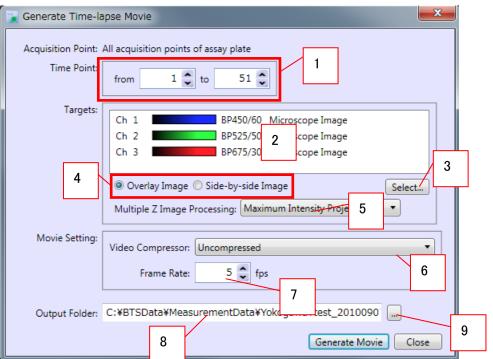
1) Menu

File menu	Explanation
Generate Movie	Display the screen to create movie files.
Save Snapshot	Capture a snapshot of image being displayed.
Close	Close the measurement result display screen.

Window menu	Explanation	
Maximize	Display measurement result sub screens by maximizing.	
Restore	Restore measurement result sub screens.	
Cascade	Display measurement result sub screens by cascade	
	mode.	
Tile	Display measurement result sub screens by tiling.	
Close All Image		
Windows	Close all measurement result sub screens.	

- 2) Change the well to display.
- 3) Change the time point to display.
- 4) Change the Z position to display.
- 5) Change the field to display.
- 6) Display area
- 7) The pixel position and intensity of each channel
- 8) Capture a snapshot of image being displayed.

Generate Time-lapse Movie Screen



- 1) Specify the time point range to create movie files.
- 2) The channels to create movie files
- 3) Select the channels to create movie files.
- 4) Output items for channel images

Item	Explanation
Overlay Image	When creating movie files, channel images are
Overlay image	output by overlaying.
Side by side Image	When creating movie files, channel images are
Side-by-side Image	output by tiling.

5) Select an output method for Z-stack images.

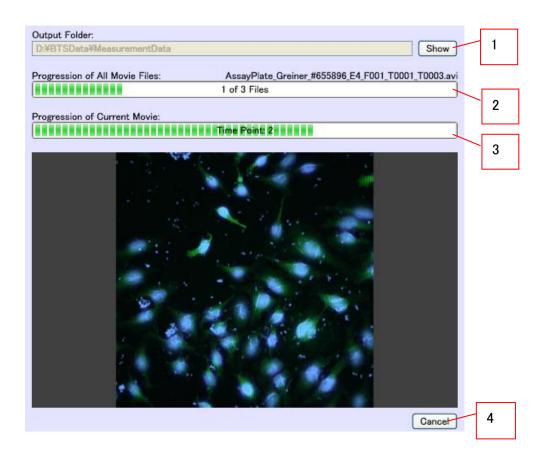
Item	Explanation
None	No Z-stack images exist.
Maximum Intensity Projection	Create movie files by MIP images. (Refer to 5.1.)
Minimum Intensity Projection	Create movie files by MinIP images. (Refer to 5.1.)
Average Intensity Projection	Create movie files by AIP images. (Refer to 5.1.)
Maximum Intensity Image	Create movie files using the Z images which have the highest total intensity in the Z-stack images at each time point.
Sum Intensity Pro- jection	Create movie files by SUM images. (Refer to 5.1.)

6) Select a compression method for creating movie files.

7) Specify the number of images replaying per second.

- 8) Folder path to output movie files
- 9) Select a folder to output movie files.

Generating Time-lapse Movie Screen

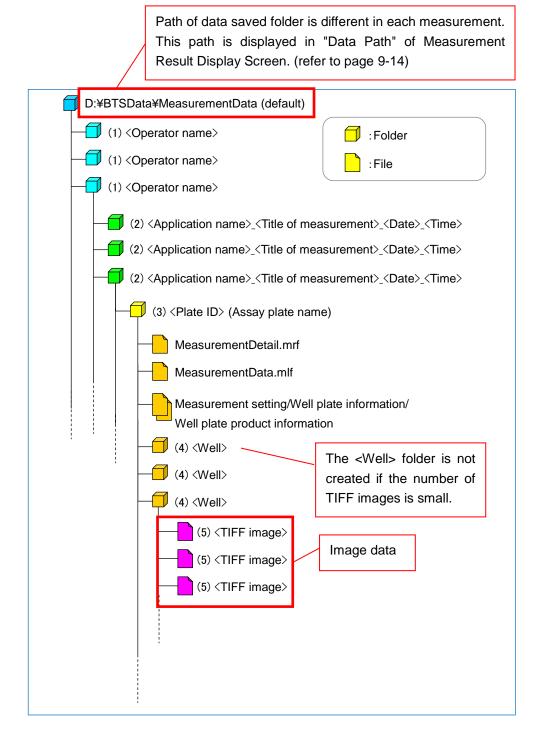


- 1) Display the output folder.
- 2) Processing bar for creating all movie files
- 3) Processing bar for creating current movie file
- 4) Stop creating movie files.

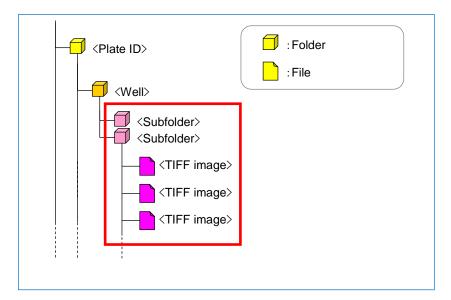
10. Saving Measurement Files

10.1. Save Location of Measurement Files

Folder for measured data has the following hierarchical structure.



If there is a large amount of data, multiple subfolders are created and image files are divided and saved in these sub-folders.



10.2. Image File Name

Image files are saved in the following format:

<Plate ID>_<Well>_T<tpIndex>F<fpIndex>L<tlIndex>A<alIndex>Z<zpIndex>C<chIndex>.tif

Plate ID : Assay plate name Well : A1, H12, etc. tpIndex : Time point number (0001 to 9999) fpIndex : Imaging point number (001 to 999) tlIndex : Time line number (01 to 99) alIndex : Action List number (01 to 99) zpIndex : 3D Z number (01 to 99) chIndex : Imaging channel (01 to 99)

11. Troubleshooting

11.1. Trouble Items

Condition	Check item
The power does not turn	Is the power plug connected to the power outlet?
on.	⇒Check the connection.
	Is 230 VAC power supplied to the power outlet to
	which the main unit is connected?
	⇒Check the breaker installed in the room or other
	applicable location in the building.
	Is the main power switch on the main body turned on?
	⇒Turn on the main power switch at the back of the main unit.
The dispenser does not	Does the source plate contain reagent?
operate or reagent does	⇒Put reagent in the source plate.
not drip.	Isn't the source plate set in a wrong orientation on the shuttle?
	⇒Set the plate so that the well A1 comes to the top
	left-hand corner of the shuttle.
	Have you entered a well plate information file?
	⇒Enter an appropriate value under "Reagent
	Volume" in the well plate information file.
	Are the correct well plate products selected?
	\Rightarrow When entering information in the well plate
	information file, well plate products are selected.
	Select appropriate well plate products. If a wrong
	product is selected, the dispenser may be
	damaged.
	Isn't the tip rack set in a wrong orientation? ⇒The tip rack set on the tip platform should be
	oriented so that remaining tips face the left side
	toward you.
	Have you entered a dispensing setting file?
	⇒Enter appropriate values under "Aspirate" and
	"Dispense." It is recommended that you perform
	simulation once.
	Have you set "Dispense Operation" in the action list
	area of the measurement setting file and specified a
	dispensing setting file?
	⇒Specify the correct dispensing setting file under
	"Dispense Operation."

Condition	Check item
The dispenser does not	Do the acquisition points under "Dispense Operation"
operate or reagent does	match the dispensing points in the dispensing setting
not drip.	file?
not drip.	⇒Dispensing is not performed unless there are
	wells matching the acquisition points specified under
	"Dispense Operation" in the measurement setting file
	and dispensing points specified in the dispensing
	setting file.
Images cannot be	Isn't the bottom of the well plate dirty?
measured.	⇒If the bottom of the well plate is dirty with
	attachment of dust or oil, a focus error may occur.
	Use a cloth, etc., to thoroughly remove the soiling.
	Are the selected well plate products correct?
	⇒When entering information in the well plate
	information file, well plate products are selected.
	Select appropriate well plate products.
	Isn't the assay plate set in a wrong orientation on the
	stage?
	⇒Set the assay plate so that the well A1 comes to
	the top left-hand corner of the stage.
Automatic analysis can	Is the CV7000 Analysis Software launched?
not be performed.	⇒To display the AnalysisProtocol setting in the
	Start Measurement screen, launch the CV7000
	Analysis Software and set to communicate the net-
	work previously.
Images are out of focus.	Have you set software focus?
	⇒When software focus is set, the cell plane to be
	captured becomes stable and the focus accuracy
	improves.
Images are dark or	Isn't the exposure time too short?
otherwise cannot be	⇒Set a longer exposure time.
seen.	Isn't the laser too weak?
	⇒Raise the laser output.
	Isn't the staining time too short?
	⇒Dark samples can be captured brighter by
	setting the binning more than "2x2."
	Isn't the filter set or laser selected incorrectly?
	⇒Set the imaging channels correctly.
The focus shifts in	Is temperature control implemented?
time-lapse imaging.	⇒Control the plate heater temperatures because a
	temperature shift causes the focal plane to deviate. It
	takes about 30 minutes after the plates are set on the
	stage until the temperatures stabilize.

Condition	Check item
-----------	------------

	Are there water drops on bottom of wellplate?
Patterns exist in image	⇒Water drops are attached on bottom of wellplate
	by temperature difference and they can be imaged.
	Wipe the water drops by soft cloth and others.
	Isn't the well plate bottom too dirty?
	⇒Wipe the plate bottom so as to be clean. AF error
	will occur if droplet or dirt is attached onto well plate
	bottom.
	Is the selected well plate product correct?
	⇒When entering information in the well plate
	information file, well plate products are selected.
	Select appropriate well plate products.
	Isn't the well plate inclined?
	⇒Remove well plate from the stage, and reset the
	plate on the stage.
	Don't you observe around well edge?
	⇒Sometimes, It is difficult to auto-focus around well
	edge because of adhesive material.
AF (auto focus) error	It is recommended to observe around well center.
occurred.	Is well filled with liquid solution?
	⇒In the case of drying in well, it may cause
	auto-focus error. Fill well with liquid solution.
	Don't you observe circumference wells when
	preview?
	⇒Depending on the selection of objective lens and
	well plate product, it may cause AF error around well
	area of plate circumference.
	It is recommended to use long working distance
	objective lens such as "20x Long W.D.".
	Don't you select wrong lens type for water immersion
	lens or dry lens?
	⇒Select each available lens type correctly for water
	immersion lens or dry lens. (Refer to 5.14.)

11.2. Message List

Warning Message	Description
Camera <camera> is not</camera>	
stably cooled:	Camera is not enough cooled. Please wait a few minutes.
<temperature> °C</temperature>	
Disk drive free space becomes less than present 50% after measurement.	Please delete unnecessary measurement data in D:¥BTS- Data¥ Measurement folder.
Image acquisition timed out: <cameras></cameras>	The cameras do not respond until timeout. The measurement software automatically retries the image acquisition.
Reader device is being controlled.	Please wait for completion of the device control.
Reader device is locked for	The measurement is running. Please wait for completion of
measurement.	the measurement.

Unexpected System	
Error and Internal	Description
Program Error	
The following error	
messages are left out of	1. Check the cables between the PC and the reader device.
this list.	
- Unexpected System	2. Check the power state of the reader device.
error	2. Check the power state of the reader device.
	3. Check the D:¥BTSData folder. Open this folder with
- Internal program error	Windows Explorer and confirm that the files can be
	accessed.
Please try actions on	
Description.	4. If the error message includes the detail code such as
If you can not solve the	'??_MANU, <id>,<error-code>,<error-message>',</error-message></error-code></id>
problem, please report to	please check the error codes of the reader firmware.
the support engineer.	
	5. Check the installation of runtime libraries.
	(Camera Control Software, DirectX, Visual C++ 2008 SP1)
	6. Check the configuration of the PC.
	(IP address, firewall, local security policy, and so on)

Error Message	Description
"Lock pages in memory"	Please read the installation guide and change local policy
privilege not assigned.	through Windows control panel.
<laser wavelength=""> lasers</laser>	Plagas turn lagare on using 'Device Cancele' > 'Lagar'
are not turned on.	Please turn lasers on using 'Device Console' -> 'Laser'.
<number> camera devices</number>	Please check the cables between the PC and the reader
not detected	device, and check the power state of the reader device.
<wellplate information=""> is not <number> wells.</number></wellplate>	The number of wells of the selected wellplate information is not same as the number of wells of the measurement setting. Please select the same number of wells of wellplate infor- mation as the measurement setting.
Acquisition Point Error at <well>: (position)</well>	The reader device can not move to the specified position of the well. The measurement continues when this error occurs.
AF Error at <well> No. <number> (<assay plate<br="">name>)</assay></number></well>	The auto focus unit can not detect the bottom of the well- plate. Please check that the correct wellplate is loaded. If the bottom of the wellplate is not clear, the auto focus unit may cause an error. The measurement continues when this error occurs.
AF Error occurred.	The auto focus unit can not detect the bottom of the well- plate. Please check that the correct wellplate is loaded. If the bottom of the wellplate is not clear, the auto focus unit may cause an error.
AF parameter not found: <wellplate name=""> (<objective>)</objective></wellplate>	You must obtain the auto focus parameter for the specified wellplate and the specified objective.
Corrected image data directory not found	Please check D:¥BTSData¥CorrectedMeasurementData folder.
Directory access failed, <folder></folder>	Please check the specified folder. It may not exists.
Dispense operation is not available on multiple time points.	Multiple dispensing is not supported. Please delete other 'Dispense Operation' action.
Dispense Timing must be	The value of dispense timing exceeds the period of the live
less than Period.	imaging. Please enter the valid dispense timing.
Elapsed time exceeds the next time point.	The elapsed time of the measurement exceeds the expected measurement time. Please open the measurement setting and save it again.
Exposure Time of Ch <number> is greater than <number> ms.</number></number>	This error occurs when the exposure time is less than <number> ms. (This error message is inappropriate. It will be changed.) Plaese increase the value of Exposure Time of the specified channel greater than <number> ms.</number></number>

Exposure Time of Ch	Please increase the value of Interval of Live Imaging or
<number> is greater than Interval.</number>	decrease the value of Exposure Time of the specified channel.
Failed to allocate camera	This error occurs when the free memory of the process is
buffer.	not enough. Please restart the measurement software.
Failed to allocate the native CameraBuffer object.	This error occurs when the free memory of the process is not enough. Please restart the measurement software.
Failed to aspirate at <well> (source plate name): <detail></detail></well>	Please check the detail of the error code. If it includes '??_MANU', the reader device causes an error.
Failed to count tips.,Please set a new tip rack.	Please load the filled tip rack.
Failed to create directory: <pre><detail></detail></pre>	Please check D:¥BTSData folder. This error occurs when D: drive does not exist.
Failed to dispense at <well> (assay plate name): <detail></detail></well>	Please check the detail of the error code. If it includes '??_MANU', the reader device causes an error.
Failed to execute the measurement., <detail></detail>	Please check the detail of the error code. If it includes '??_MANU', the reader device causes an error.
Failed to get reader device status	Please check the reader device.
Failed to initialize frame grabber.	This error occurs when the free memory of the process is not enough. Please restart the measurement software.
Failed to initialize measurement software.	Please check the detail of the error code. If it includes '??_MANU', the reader device causes an error.
Failed to initialize video compressor: <detail></detail>	The selected video compressor is not supported by the movie generator. Please select another video compressor.
Failed to load <file></file>	The specified file is incorrect. Please report to the support engineer.
Failed to load the measurement data: <file></file>	The specified file is incorrect. Please report to the support engineer.
Failed to load the measurement setting: <file></file>	The specified file is incorrect. Please report to the support engineer.
Failed to load the well plate product information: ID= <number></number>	The <number> .wpp file is incorrect. Please report to the support engineer.</number>
Failed to lock reader device control.	The measurement software is controlling the reader device (such as start-up). Please wait for the completion of the device control.
Failed to prepare measurement. Please try again later.	The measurement software is controlling the reader device (such as start-up). Please wait for the completion of the device control.
Failed to prepare the measurement., <detail></detail>	The reader device is working. Please wait for the completion of the device control.
Failed to read	Please check the barcode of the wellplate. It may not be clear.

	
Failed to save image.	
Image Processing: <image< td=""><td>Please check the disk free space of D: drive.</td></image<>	Please check the disk free space of D: drive.
processing>	
Failed to save the	
measurement setting:	Please check the disk free space of D: drive.
<file></file>	
Failed to scan <folder></folder>	Please check the specified folder. It may not exist.
Failed to shut down	Please check the detail of the error code. If it includes
device, <detail></detail>	'??_MANU', the reader device causes an error.
Failed to start	Please check the detail of the error code. If it includes
device, <detail></detail>	'??_MANU', the reader device causes an error.
Failed to test 2D	Please check the detail of the error code. If it includes
acquisition.	'??_MANU', the reader device causes an error.
Failed to test 3D	Please check the detail of the error code. If it includes
acquisition., <detail></detail>	'??_MANU', the reader device causes an error.
Failed to test acquisition	You can not use the image acquisition test when the
parameters.,Measurement	measurement is running.
is running.	Please wait for the completion of the measurement.
Failed to update data	Please check D:¥BTSData folder. This error occurs when D:
directory: <detail></detail>	drive does not exist.
Image acquisition timed	The camera does not respond until timeout. Please retry
out.	acquire image.
	This error occurs when D:¥BTSData¥MeasurementData
Insufficient disk drive free	folder contains many image files.
space.	Please delete unnecessary measurement data.
Insufficient memory.	This error occurs when the free memory of the process is not enough. Please restart the measurement software.
Interval of live option is less	
than <number> ms.</number>	Please increase the value of Interval of Live Imaging.
	You entered the incorrect application name. For example,
Invalid application name.	the following characters are not available: $\frac{1}{2}$ / : * ? " < >
	You entered the invalid value of the CO2 density. Please
Invalid density.	enter the value from 0.0 to 9.9.
Invalid filter of Ch	You do not select the filter of Acquisition menu of the
<number>.</number>	channel setting. Please select an item of Acquisition menu.
Invalid flow rate.	You entered the invalid value of the CO2 flow rate. Please
	enter the value from 0 to 500.
Invalid measurement	The measurement setting file is incorrect. Please report to
setting. Please check it.	the support engineer.
Invalid number formats of	
operating system. Decimal	The supported decimal symbol is only ".". Please check the
symbol must be ".".	regional formats of Windows:
Please change number	'Start Menu' -> 'Control Panel' -> 'Region and Language' ->
formats of operating system	'Formats'
with Control Panel.	If 'Format' is not 'English', please change the format.
Program will be exited.	in romationot English, please change the format.

Invalid parameter specified.	Please check the input parameter. Invalid parameter field is marked with red color.			
Invalid path of Output Folder.	You entered the invalid path name. Please enter the valid full path name of the output folder.			
Liquid surface detection failed at <well> (<source plate name>)</source </well>	Please check the detail of the error code. If it include '??_MANU', the reader device causes an error.			
Live Imaging is not available on time lapse.	You can not specify the interval time on 'Time-lapse Setting' when 'Live Imaging' of 'Fluorescence Acquisition' or 'Bright-field/Phase-contrast' is turned on.			
Loaded tip rack is not available.	Please load the filled tip rack.			
Local IPv4 network address not found: Local host= <hostname></hostname>	Please check the configuration of IP address.			
Measurement is already running.	Please wait for the completion of the measurement.			
No image data exists.	This error occurs when you generate the movie with the channel which contains no image file. Please deselect such the channel.			
No Image Processing selected.	Please select an item of Image Processing.			
No objective selected.	The objective lens is not selected on the channel setting o the measurement setting. Please select the objective lens.			
No reagent volume for <well> <source plate<br=""/>name></well>	The values of the reagent volume are not specified in the wellplate information. Please enter the reagent volume in the wellplate information.			
No sample volume for <well> <assay name="" plate=""></assay></well>	The values of the sample volume are not specified in the wellplate information. Please enter the sample volume in the wellplate information.			
No target is selected.	Please select targets.			
No Video Compressor selected.	Please select an item of Video Compressor.			
Not enough tips to start measurement. Please set a new tip rack.	Please load the filled tip rack.			
Operator, Measurement Title, or Assay Plate name is too long.	You entered long operator name, long measurement title, o long assay plate name. Please shorten the length of them.			
Output folder must be full path.	You entered the relative path name. Please enter the valid full path name of the output folder.			
Please add a channel.	This error occurs when you delete the channels of the measurement setting. Please add a channel.			
Please add a time line and set its parameters.	This error occurs when no time line exists on the measure- ment setting. Please add a time line.			

Please enter correct	Please check the input parameter. Invalid parameter field is			
parameter.	marked with red color.			
Please enter correct power	You entered the invalid value of the laser/lamp power.			
of <laser>.</laser>	Please enter the value from 0 to 100.			
Please enter dispense	On 'Dispense Operation' action, You do not enter the			
setting name.	dispense setting name.			
Please enter valid file	You entered the incorrect file name. For example, the			
name.	following characters are not available: ¥ / : * ? " < >			
Please enter valid	Please check the input parameter. Invalid parameter field is			
parameters.	marked with red color.			
Please put <assay plate,<="" td=""><td>The assay plate, the source plate, or the tip rack is not</td></assay>	The assay plate, the source plate, or the tip rack is not			
source plate, or tip rack>.	loaded. Please load them.			
Please select a well plate	You do not select a wellplate type on the list view. Please			
type.	select one.			
Please select an item.	You do not select an item on the list view. Please select an item.			
Please select the bright-field/phase-contrast target.	On 'Bright-field/Phase-contrast Acquisition' action and 'Z-Stack Bright-field/Phase-contrast Acquisition', you must select a 'Brightfield' channel or an 'Phase Contrast' channel.			
Please select the different Acquisition of Ch <number>.</number>	You selected the two or more channels which can not be acquired at a time. To acquire the channels at a time, the following parameters must be different from each other: Acquisition.			
Please select the fluorescence targets.	On 'Fluorescence Acquisition' action and '3D Fluorescence Acquisition', you must select a 'Confocal Fluorescence' channel or an 'Epifluorescence' channel.			
Please select the same Method, Objective, Light Source of Ch <number></number>	You selected the two or more channels which can not be acquired at a time. To acquire the channels at a time, the following parameters must be unique: Method, Objective, Light Source.			
Please select the target.	You do not select a channel. Please select.			
Reader is working now.	The measurement software is controlling the reader device (such as start-up). Please wait for the completion of the device control.			
sCMOS camera not found.	Please check the cables between the PC and the reader device, and check the power state of the reader device.			
The action contains invalid	Please check Action List of the measurement setting. Action			
parameters.	shows the reason of the error.			
The dispense operation	You can specify only one dispense operation on the			
already exists in the	measurement setting. Please delete the other dispense			
measurement setting.	operation.			
The number of the				
channels must be up to 99.	This error occurs when you add more than 99 channels to the measurement setting. Please delete unnecessary channels.			

The reader device is working now.	The measurement software is controlling the reader device (such as start-up). Please wait for the completion of the device control.				
The selected target is not bright-field/phase-contrast.	On 'Bright-field/Phase-contrast Acquisition' action and 'Z-Stack Bright-field/Phase-contrast Acquisition', you must select a 'Brightfield' channel or a 'Phase Contrast' channel.				
The selected target is not confocal.	On 'Software Focus' action, you must select a 'Confocal Fluorescence' channel.				
The selected target is not fluorescent.	On 'Fluorescence Acquisition' action and '3D Fluorescence Acquisition', you must select a 'Confocal Fluorescence' channel or an 'Epifluorescence' channel.				
Time Line: <time-line> Action List contains invalid parameter.</time-line>	Action List of the measurement setting contains invalid parameter. Please check Action List. Invalid parameter field is marked with red color.				
Time Line: <time-line> Time-lapse Setting contains invalid parameter.</time-line>	Time-lapse Setting of the measurement setting contains invalid parameter. Please check Time-lapse Setting. Invalid parameter field is marked with red color.				
Time Line: <time-line> Well Plate Scan Setting contains invalid parameter.</time-line>	Well Plate Scan Setting of the measurement setting contains invalid parameter. Please check Well Plate Scan Setting. Invalid parameter field is marked with red color.				
Too many image windows are opened.	This error occurs when you open the new image window without closing other image window. Please close unnecessary image windows.				
Too many targets are selected. Please select up to 4 targets.	You selected more than 4 targets. Please select up to 4 targets.				
You cannot test acquisition parameters when measurement system is working.	You cannot use the image acquisition test when th measurement is running. Please wait for the completion of the measurement.				
You can select either immersion lenses or non-immersion lenses in a measurement setting.	You selected both the immersion lens and the non- immersion lens. Please select either one of them.				

11.3. Error Code List

WP Error code list

Error message	Error code	Measures	Cause of the trouble
WP_IM_WIM_OBJ_CANT_USE	-204713	Select the Water immersion objective mode	Used water immersion objective lens without water.
WP_IM_DRY_OBJ_CANT_USE	-204714	Select the dry objective mode	Used the dry objective with Water immersion objective mode
WP_IM_WA_SPLY_NOT_READY	-204715	Turn on the water supply system.	Used the Water immersion objective without water supply system.

CD Error code list

Error message	Error code	Measures	Cause of the trouble
CD_PLATE_INFO_ERR	-206311	Make sure the source plate	Source plate information is
	-200311	information	incorrect.
		Make sure the source plate	
CD_WELL_POS_PARAM_ERR	-206313	information	Dispensing position is out of
	-200313	Set the dispensing position	range
		at center of the well.	
CD_ARM_XY_NOT_ESC	-206320	Return Z motor to origin	Dispenser is not at the
	-200320	position	evacuation site.
CD_ARM_Z_NOT_ESC	-206321	Return Z motor to origin	Dispenser is not at the
	-200321	position	evacuation site.
CD_CURR_ARM_POS_ERR	-206323	Set the dispenser at the	Dispenser is not at the center
CD_CORK_ARM_FO3_ERK	-200323	center of the well.	of the chip.
		Make sure the plate	
CD_WELL_LIQ_VOL_ERR	-206340	information	Volume setting of filing and
	-206340	Check the filling and	dropping is invalid.
		dropping volume.	
	-206350	Remove the chip from the	Performed chip count/mount
CD_TIP_ALREADY_PICKUP		-206350 dispenser	while the chip already
		uspenser	existing on the dispenser.
		Re-register the chip	
CD_FAIL_TO_TIP_EJECT	206251	mounting position	CV7000 failed to discard the
CD_FAIL_TO_TIP_EJECT	-206351	Make sure that dump has	chip.
		not jammed.	
		Try again by mounting the	
CD_TIP_NOT_PICKUP	-206352	chip	Chip was not mounted on the
CD_TIF_NOT_FICKOF	-200352	Re-register the chip	dispenser.
		mounting position	
CD_FAIL_TO_TIP_PICKUP	-206353	Re-register the chip	Failed to pick up the chips
	-200303	mounting position	י מוופע נט אוטא עם נוופ טוואס

CD_FAIL_TO_TIP_DETECT	-206354	Re-register the chip mounting position	Chip was mounted during the chip detection.
CD_TIP_CNT_UNKNOWN	-206355	Count the chip number.	CV7000 was not sure the number of remaining chips.
CD_TIP_USABLE_CNT_IS_0	-206356	Replace the chip rack	Number of chips at the chip rack was to empty
CD_TIP_RACK_UNUSABLE	-206359	Make sure that the incorporation of the chip rack	Chip rack is not mounted
CD_TIP_CHUTE_DUCT_JAM	-206360	Make sure that dump has not jammed.	Chip may jammed in the dump

MS_IW Error code list

Error message	Error code	Measures	Cause of the trouble
	-201211	Turn ON the power of the	CV7000 power is not turned
MS_IW_POWER_OFF		CV7000	on
	-201214	Turn OFF the emergency	Emergency stop button is
MS_IW_EMERGENCY		stop button.	turned ON
MS_IW_AIR_SUPPLY_ERR	201222	Supply the air to CV7000	Air is not supplied to the
IVIS_IVV_AIR_SUPPLI_ERR	-201223		CV7000

LS Error code List

Error message	Error code	Measures	Cause of the trouble
LS_INTERLOCK_ERR	-207016	Cancel the interlock by	Interlock shutted down the
		using adjustment tools	laser for safety.
LS_ERR_LS_NOT_READY	-207041	Launch the laser.	Manipurated the laser power
			while laser has not ready.

ST Error code list

Error message	Error code	Measures	Cause of the trouble
ST_PLATE_INFO_ERR	-203010	Make sure the plate information	Plate information is incorrect.
ST_TRGT_WELL_NUM_ERR	-203012	Set the stage movement within well radius	Stage movement has variance with the well plate information.
ST_FORBIDDEN_TRGT_WELL	DEN_TRGT_WELL -203014		Stage has moved to Unavailable area
ST_FORBIDDEN_TRGT_POS	-203015	Do not move to the the specified location	Stage has moved to Unavailable area
ST_SET_SAFEGUARD	-203022	Remove the obstacles on the stage	Safety system.
ST_LIFTER_ST_POS_ERR	-203025	Re-register "Loader Handover"position. Change the position of the Sensor enable to operate the lifter	Lifter sensor could not detect the lifter.

11. Troubleshooting

ST_LIFTER_PLATE_EXIST	-203026	Remove the plate on the lifter.	Plate is placed on the lifter
ST_FORBIDDEN_TRGT_AREA	-203028	Do not move at the specified location	Stage has moved to Unavailable area

HT Error code List

Error message	Error code	Measures	Cause of the trouble
HT OVERSCALE ERR	-203521	Check wiring connections	The sensor detects the
HI_OVERSCALE_ERR	-203521	Clean the connector	temperature is too high
	202525	Check wiring connections	Poor Connection
HT_BURNOUT_ERR	-203525	Clean the connector	Wiring is snapped

YP Error code list

Error message	Error code	Measures	Cause of the trouble	
			Motor does not return to the	
			origin.	
YP_MT_NOT_READY	-101001	Motor return to the origin	but If you want to return to	
			origin MTWP01 or MTWP02,	
			please check the valve	
		Cancel the alarm and		
YP_ACK_ERR	-101004	correct the cause of the	Alarm has occurred	
		alarm		
		Cancel the alarm and		
YP_END_ERR	-101005	correct the cause of the	Alarm has occurred	
		alarm		

12. Maintenance and Inspection

- This equipment uses many plastic parts. When cleaning these parts, wipe them with a dry, soft cloth. Do not use benzene, thinner or other chemical substance or detergent to clean plastic parts, as it may cause discoloration, deformation or damage.
- If the equipment malfunctions, contact us without attempting to access the inside or take any other actions to resolve the problem yourself.
- If you perform measurement with the dispensing operation, check that the tip rack disposal box is not full. The tip waste should be disposed of by the waste disposal contractor.

13. Warranty

- 1) The warranty period is one year from the date of installation. Failures that occur within the warranty period will be repaired for free.
- 2) The warranty applies only to this equipment.
- 3) If the following applies, repair will be charged even within the warranty period.
- Failures or damage caused by inappropriate handling or use.
- Failures or damage caused by handling, use, or storage that exceed the tolerance of the design and specifications.
- Failures or damage caused by the repairs and modifications made by the user.
- Failures or damage caused by the transportation, movement, dropping, or the like after purchase.
- Failures or damage caused by a fire, natural disaster (earthquake, storm, or flood disaster), salt damage, gas damage, and abnormal voltage.
- 4) Any other damage not attributable to Yokogawa is outside the scope of the warranty.
- 5) If you need any repairs, please consult with us.

Contact Us

Please contact the dealer inquiries about this product.

Manufacturer International Sales Team Life Science Center, Measurement Business HQ Yokogawa Electric Corporation 2-3 Hokuyodai, Kanazawa-shi, Ishikawa 920-0177, Japan Phone +81-76-258-7028 Fax +81-76-258-7029

E-mail CSU_livecell_imaging@cs.jp.yokogawa.com

Home page http://www.yokogawa.co.jp/scanner

14. General Specification

Model	CV7000				
Supported sample					
vessels	Wellplate (6, 24, 96, 384, 1536 wells), Slideglass				
	Confocal mode (simultaneous imaging of max.3 colors)				
Oh a smustism mass da	Epi-fluorescence mode (simultaneous imaging of max.3 colors)				
Observation mode	[Option]Bright field mode				
	Phase-contrast mode (The latter is available for wellplates of 6/24 wells)				
Output data format	Image data: 16 bit TIFF, PNG; Numerical data: CSV, Original format				
Excitation					
wavelength	405/488/532/640nm, all solid laser [Option] 365nmLED				
White light					
illumination	[Option] 100W halogen lamp				
Autofocus	Laser-based mode, Image-based mode				
	Max. 6 lenses are available: Automatically switchable				
Ohiaatiya lawa	Dry 4X, 10X, 20X, 40X Water immersion 60X				
Objective lens	Long working distance 20X Phase-contrast 10X, 20X (Only for 6/24				
	wellplates)				
Confocal unit	Microlens-enhanced wide-view dual nipkow disk confocal scanner				
Comoro	Max. 3 cameras				
Camera	(sCMOS, Number of pixels:2560 X 2160, pixel size:6.5μm)				
	[Option] Capable of live cell imaging.				
Stage incubator	Temperature range: 35 up to 40°C, CO ₂ supply box (CO ₂ : 5%, humidifier)				
Dispenser	[Option] Disposable tip type				
Bar-code reader	[Option] Target codes: 1 up to 2 dimension				
Markatation	Dual-monitor workstation for system control				
Workstation	Dual-monitor workstation for data analysis				
Operating	45 up to 20° 0. 40 up to 700 DU (Ne condensation)				
environment	15 up to 30°C 10 up to 70%RH (No condensation)				
	Measurement unit: Single-phase 200VAC, 2KVAmax				
Power supply	Workstation for system control:100-240VAC, 1.5KVAmax				
	Workstation for data analysis: 100-240VAC, 1.5KVAmax				
Dimension	Measurement unit: 1496W X 1160D X 1650H (mm)				
	Workstation for system control: 1000W X 700D X 1200H (mm)				
Weight	Measurement unit: 650kg, Workstation for system control: 50kg				
Noise level	Less than 70 dB				

14.1. MS Code

Main Unit

Madal				0				Ontion	
Model Code				Suffix Code				Option Code	Description
				Code				Code	High-throughput Cytological Discovery
CV7000									System
-C1									sCMOS one unit
Camera -C2									sCMOS two units
-C3									sCMOS three units
	F02								405, 488, 561, 640 nm
	F03								488 nm
	F04								405, 488 nm
Laser	F05								405, 488, 561 nm
	F06								488, 561, 640 nm
	F07								488, 561 nm
	U								With UV light source
UV	N								Without UV light source
		_							With Bright field/ Phase contrast
Bright Field		В							illumination
/ Phase contras	st	<u> </u>							Without Bright field/ Phase contrast
		Ν							illumination
		-	_						96 tip type dispenser,
			-D						loader (reagent plate and tip rack)
Dispenser		-	_						384 tip type dispenser,
			-E						loader (reagent plate and tip rack)
		ľ	-N						Without dispenser
									Full option
				F					(Temperature control, Humidifier and
									Water immersion)
				14/					Temperature control and Water
Stage Heater				W					immersion
/ Water Immers	sion		İ	Н					Temperature control and Humidifier
			ĺ	Т					Temperature control
				L					Water immersion
			ĺ	NI					Without Stage Heater and Water
				Ν					immersion
				С					With CO ₂ supply
CO ₂ Supply				Ν					Without CO ₂ supply
Sub code				-()0				Always "-00"
					-0				AC 200 V
Device Original					-1				AC 220 V
Power Supply				-2				AC 230 V	
					-3				AC 240 V
									Analysis workstation and Analysis
Market-time (-14			Software (1 st license)
Workstation for	Analysis	5				40			Without analysis workstation and
						-13			Analysis Software
							-J		Japanese
Language							-E		English
Onting							•	/R1	Single barcode reader
Option								/R2	Double barcode reader

Objective Lens

-			
Model	Suffix		Option Description
Code	Code		Code
CVLNS			Objective lens
-	7000		For CV7000
Position ^{^{%1}}		·P□	Objective position (without heater)
FUSILION		·H□	Objective position (with heater)
		-L04A	4 x dry (NA=0.16)
		-L10A	10 x dry (NA=0.40)
		-L20A	20 x dry (NA=0.75)
Objective Le		-L40A	40 x dry (NA=0.95)
Objective Le	:15	-L60W	60 x water immersion (NA=1.2)
		-20AL	20 x long working distance (NA=0.45)
		-L10P	10 x phase contrast (NA=0.30)
		-L20P	20 x phase contrast (NA=0.45)
Separate Or	der		/S Separate Order

[™] □: Position 1 - 6

CSU DM, PARA DM

Model Code	Suffix Code	Option Code	Description
CV7KDM			
	-D		Dummy code
Confoool	M (Diobroio Mirror)	/D∆01	DM D405/488/561/640
Confocal DM (Dichroic Mirror) ∆: Position 1 – 2		/D∆02	DM D405/488/561
		/D∆03	DM D405/488/640

Image Splitting DM

Model Code	Suffix Code	Option Code	Description
CV7KCA			
	-D		Dummy code
Imaga Cal	itting DM (Dichroid Mirror)	/D∆01	DM488
Image Splitting DM (Dichroic Mirror) △: Position 1 – 2		/D∆02	DM561
		/D∆03	DM640

Emission Filter

Model Code	Suffix Code	Option Code	Description
CV7KFW			
	-C 🗆		□: Camera port number 1 – 3
		/∆01	BP445/45 for 405nm
		/∆02	BP525/50 for 488nm
EM (Emissio	on Filter)	/∆03	BP600/37 for 561nm
∆: Position 1 – 5		/∆04	BP679/29 for 640nm
		/∆05	BP447/522 for 405nm and 488nm
		/∆06	BP488/568 for 488nm and 561nm

Option

Model Code	Suffix Code	Option Code	Description
CV7KPRT	Code	Code	
	-D		Dummy code
Option		/WS14	Analysis workstation and Analysis Software (2 nd license)